

REVIEW ARTICLE

Advances in understanding the mechanism and treatment strategies of radiation myelopathy

Yuhang Yu¹, Yukai Tang¹, Shengyi Liu¹, and Limin Liu^{1*}Department of Hematology and Oncology, The 921st Hospital of Joint Logistics Support Force, The Second Affiliated Hospital of Hunan Normal University, Changsha, Hunan, China

Abstract

Radiation myelopathy (RM) is a severe, late-onset complication of radiotherapy, involving complex pathological processes, such as vascular endothelial cell damage, disruption of the blood-spinal cord barrier, inflammation, demyelination, hypoxia, and tissue necrosis. Traditional treatments, including corticosteroids and immunoglobulins, can effectively alleviate acute symptoms, but their long-term use may cause side effects and offer limited efficacy, especially in advanced stages of the disease where significant neurological recovery remains challenging. In recent years, emerging therapeutic strategies for RM – such as neuromodulation technologies, stem cell transplantation, tissue engineering, and gene therapy – have gained increasing attention. These approaches promote spinal cord repair and functional recovery through mechanisms, such as neuroprotection, myelin regeneration, axonal regeneration, and immune modulation. In addition, the use of biomaterials, such as hydrogels and nanodelivery systems has enhanced the delivery efficiency and therapeutic efficacy of both drugs and cells. Future research should focus on optimizing intervention timing and developing combination treatment strategies – such as incorporating antifibrotic drugs, anti-inflammatory therapies, and hyperbaric oxygen therapy – to improve the microenvironment of injury and enhance therapeutic outcomes. This review evaluates the pathological mechanisms of RM, explores emerging therapeutic strategies, and highlights future research directions to improve clinical efficacy.

*Corresponding author:

Limin Liu
(liulm7080@163.com)

Citation: Yu Y, Tang Y, Liu S, Liu L. Advances in understanding the mechanism and treatment strategies of radiation myelopathy. *Eurasian J Med Oncol.* 2026;10(1):1-14.
doi: 10.36922/EJMO025100046

Received: March 7, 2025**Revised:** April 17, 2025**Accepted:** May 8, 2025**Published online:** May 29, 2025**Copyright:** © 2025 Author(s).

This is an Open-Access article distributed under the terms of the Creative Commons Attribution License, permitting distribution, and reproduction in any medium, provided the original work is properly cited.

Publisher's Note: AccScience Publishing remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Keywords: Pathological mechanisms; Radiation myelopathy; Review; Treatment strategies

1. Introduction

Radiotherapy is an essential treatment for both primary and metastatic tumors. However, its dosage is limited by late and irreversible damage to surrounding normal tissues and organs, known as late effects. Radiation myelopathy (RM) is a serious late complication of radiotherapy that requires special attention when treating spinal, spinal cord, and paraspinal tumors, as well as head, neck, and lung cancers. In clinical practice, radiotherapy is typically administered in small fractionated doses over several weeks. The cellular response to this fractionated approach is influenced by four key biological factors: Radiosensitivity, the ability to repair sublethal damage, cell cycle redistribution and reoxygenation, and the ability to repopulate after radiation.

Fractionated radiotherapy optimizes treatment efficacy by taking advantage of the differential responses of normal tissues and tumors. Based on extensive clinical experience, it is widely accepted that when the radiation dose to the spinal cord is controlled at 1.8 – 2.0 Gy per day, and the total dose remains within 45 – 50 Gy, the risk of permanent injury is low – estimated at 0.03 – 0.2%.¹ Stereotactic body radiation therapy (SBRT) has been widely used in the treatment of spine, spinal cord, and paraspinal tumors due to its precise and efficient dose delivery. However, the high-dose fractionation mode used in SBRT presents new challenges for the prevention and treatment of RM. A study analyzing data from patients with spinal metastases treated with SBRT identified new dosimetric constraints aimed at reducing RM risk, but further research is required to validate the efficacy and safety of these measures.² There is no evidence indicating that RM caused by SBRT differs significantly from that caused by standard fractionated radiotherapy in terms of pathobiological mechanisms. While permanent myelopathy is extremely rare with conventional fractionated radiotherapy, the incidence of RM has increased with the rise in spinal SBRT and re-irradiation treatments.³

The pathophysiological mechanisms of RM involve several complex processes, including vascular endothelial cell injury, blood-spinal cord barrier (BSCB) disruption, inflammation, demyelination, hypoxia, and angiogenesis.⁴ Present treatments for RM primarily include steroids, anticoagulants, anti-inflammatory therapies, and physical rehabilitation. Although these treatments are effective in alleviating acute symptoms and slowing disease progression, their long-term use may lead to side effects, such as opportunistic infections, endocrine abnormalities, and weight gain. In addition, their overall efficacy is limited, particularly in advanced stages of the disease, where significant neurological recovery is difficult to achieve. Furthermore, these treatments generally lack the specificity required to reverse spinal cord injury or promote regeneration. Emerging therapies, such as stem cell transplantation and gene therapy, have shown promising results in animal studies but face challenges including low transplantation efficiency, ethical concerns, and long-term safety in clinical applications. Therefore, exploring more precise and efficient therapeutic strategies – particularly those based on nerve repair and tissue regeneration – has become a key focus in RM research. This review aims to evaluate the pathological mechanisms and emerging treatment strategies for RM, with a focus on the potential of neuromodulation, stem cell transplantation, tissue engineering, and gene therapy, while also highlighting future research directions to improve clinical treatment effects.

2. Pathophysiologic mechanisms of RM

2.1. Vascular endothelial cell injury and BSCB disruption

The BSCB is a specialized structure that separates the blood and spinal cord tissues and plays a crucial role in maintaining spinal cord homeostasis. Radiation induces apoptosis in spinal cord vascular endothelial cells through the lipid second messenger ceramide, activating the acid sphingomyelinase-mediated apoptotic signaling pathway. The death of these endothelial cells disrupts BSCB integrity, impairing its function and allowing blood components to infiltrate the spinal cord parenchyma. This infiltration triggers an inflammatory response, exacerbating tissue damage,⁵ and represents an early critical stage in RM progression.⁶ Interestingly, radiation therapy does not significantly increase blood-brain barrier permeability following a total dose of 20 Gy delivered in two fractions,⁷ suggesting that vascular endothelial cell injury in the spinal cord is closely associated with cumulative radiation dose and regional susceptibility. While spinal cord vascular endothelial cell injury contributes significantly to late-stage damage in RM,⁸ it does not appear to be directly linked to long-term neuroinhibition.⁹

2.2. Demyelination

Demyelination is a key hallmark of advanced radiation injury in the spinal cord, indicating that oligodendrocytes (OLs) – the cells responsible for myelin formation – have been damaged by radiation. Animal model studies have demonstrated that OLs in the spinal cord of mice exhibit significant apoptosis, decreased density, and impaired function within hours of radiation exposure.¹⁰ This process is primarily driven by the activation of the p53 signaling pathway in response to radiation-induced DNA damage. In addition, oligodendrocyte progenitor cells (OPCs) undergo early apoptosis following radiation, leading to a reduced cell population. These progenitor cells begin to proliferate approximately 2 weeks post-radiation and typically recover to baseline density by approximately 6 weeks.¹¹ Both the rate of apoptosis in OL and the recovery of OPCs after 6 weeks are dose-dependent. The early reduction of OPCs following radiation may impair OL turnover, potentially leading to demyelination and neurological dysfunction. The ability of OPCs to proliferate and recover is crucial for preventing or mitigating these long-term injuries.¹² Neural stem cells (NSCs) – which can differentiate into neurons, astrocytes, and OLs – may support OL turnover when administered early post-radiation, offering a promising therapeutic strategy for RM.

2.3. Hypoxia and angiogenesis

Vascular endothelial cell injury and disruption of the BSCB can lead to localized hypoxia in the spinal cord.

Animal studies have shown that hypoxia stimulates the upregulation of hypoxia-inducible factor-1 α (HIF-1 α) and the expression of the vascular endothelial growth factor (VEGF) gene, with a positive correlation to radiation dose.^{13,14} VEGF, a transcriptional target gene of HIF-1 α , is upregulated under hypoxic conditions and plays crucial roles in angiogenesis and neurogenesis in RM. In RM models, hypoxia-induced VEGF has been shown to exacerbate secondary injury by increasing vascular permeability and disrupting the BSCB. This effect may be mediated either directly by VEGF's action on vascular endothelial cells or indirectly through other mediators, such as matrix metalloproteinase-9.¹⁵

VEGF has been shown to play multiple roles in RM (Table 1). VEGF increases BSCB permeability, leading to edema and contributing to secondary injury in RM.^{16,17} Conversely, VEGF may also exert neuroprotective effects by enhancing vascular density, restoring blood supply, and promoting neuronal survival, axon regeneration, and functional recovery.^{10,18-22} The diverse roles and underlying mechanisms of VEGF in RM warrant further investigation.

2.4. Tissue necrosis and neurodegeneration

The persistent progression of hypoxia, inflammation, and demyelination eventually leads to necrosis of the spinal cord white matter. Neurons within necrotic regions lack regenerative capacity, leading to permanent neurological deficits. In addition to direct tissue necrosis, radiation can also cause neuronal degeneration, potentially due to the increased sensitivity of neurons to hypoxic and inflammatory conditions, along with the impaired axonal transport mechanisms.

2.5. Oxidative stress and inflammation

Radiation therapy can induce oxidative stress – including the generation of reactive oxygen species and reactive nitrogen species – by damaging neuronal membranes,

mitochondria, and DNA. Oxidative stress not only directly harms neurons, impedes axonal regeneration, and hinders functional recovery, but also exacerbates vascular endothelial cell injury and disrupts the blood-brain barrier. In addition, it activates inflammatory signaling pathways (e.g., the nuclear factor kappa B pathway), leading to the release of inflammatory cytokines (e.g., tumor necrosis factor α , interleukin [IL]-1 β , and IL-6), which further aggravate tissue damage.

3. Present status of RM treatment

3.1. Traditional treatments

3.1.1. Glucocorticosteroid

Glucocorticoids play a crucial role in the treatment of RM by suppressing inflammatory responses, modulating immune activity, and reducing spinal cord edema, thereby effectively alleviating the neurological dysfunction caused by radiation injury. In the acute phase (within 8 h of trauma), high-dose shock therapy is typically administered during the first 24 h. This involves an initial loading dose of methylprednisolone at 30 mg/kg infused over 15 min, followed by a maintenance dose of 5.4 mg/kg/h infused for 45 min. After the loading dose, the same infusion rate is maintained for 23 h. This treatment is effective as an early intervention, helping to reduce pain and improve sensory as well as motor function. However, the dosage used far exceeds standard shock doses, increasing the risk of adverse effects such as infections.²⁴ In chronic or advanced RM patients, glucocorticoids show limited efficacy and are primarily used to control secondary inflammation and relieve residual symptoms. Moreover, long-term use of glucocorticoids can lead to side effects such as hyperglycemia, increased infection risk, and osteoporosis. Therefore, the risks and benefits of treatment must be carefully evaluated and tailored to each patient's specific condition. Overall, glucocorticoids are essential

Table 1. Summary of literature on the mechanisms of vascular endothelial growth factor in radiation myelopathy

Source of VEGF	Impact of VEGF	Summary mechanism	References
Endogenous	-	Arteriosclerosis.	16
Exogenous	-	Increased permeability of BSCB.	17
Endogenous	+	Relief of spinal cord infarction and apoptosis inhibition.	20
Endogenous	+	Improvement of ischemia and hypoxia.	22
Endogenous	+	Inhibit apoptosis.	23
Endogenous	+	Promotes angiogenesis in response to hypoxic environments.	19
Exogenous	+	Protects or repairs blood vessels and reduces apoptosis.	18
Exogenous	+	Promotes angiogenesis and contributes to neural support.	21

Note: “-” indicates negative impact of VEGF, while “+” indicates positive impact of VEGF.

Abbreviations: BSCB: Blood-spinal cord barrier; VEGF: Vascular endothelial growth factor.

in the treatment of RM and hold significant clinical value, especially during the acute phase.

Studies have shown that intravenous immunoglobulin (IVIG) is highly effective in treating hormone-resistant RM.²⁵ In cases of delayed RM, even when neurological symptoms fully resolve after adequate glucocorticoid shock therapy, imaging often reveals that the lesion persists, presenting a potential risk for delayed injury. However, combining IVIG with glucocorticoid therapy led to complete resolution of the lesion on imaging after 6 months, suggesting that IVIG may provide a therapeutic advantage in stabilizing delayed RM.²⁶ For RM induced by immunotherapy combined with radiotherapy, conventional treatments (such as glucocorticoids alone) have limited efficacy, with more than 80% of patients left with neurological deficits.²⁷⁻²⁹ However, a regimen combining high-dose glucocorticoids with IVIG has demonstrated better efficacy in treating immune-related RM. For patients with poor response or disease recurrence, further treatments – such as plasma exchange or mycophenolate mofetil – may be considered.³⁰

3.1.2. Adjunctive treatment

Adjunctive therapies for RM include antioxidant treatments, therapies that enhance the blood supply, and rehabilitative therapies. These are typically used as supplementary treatments alongside primary therapies – such as glucocorticoids – to alleviate symptoms, improve the spinal cord microenvironment, and promote functional recovery.

Antioxidant therapy can mitigate the pathological changes associated with RM, promote functional recovery, and improve patients' quality of life by scavenging free radicals, reducing oxidative stress and inflammation, and protecting neuronal and vascular function. Examples of antioxidant agents include oxygen radical scavengers, such as edaravone³¹; minerals such as magnesium, copper, and manganese; antioxidant vitamins such as vitamin E and vitamin C; polyphenol-rich botanicals, such as grape seed and raspberry seed; and saponin-rich botanical extracts from plants, such as *Panax ginseng*, *Rhizoma Ligustici Chuanxiong*, *Heptaphyllum* seed, alfalfa, and spinach. These agents primarily function by maintaining antioxidant enzyme activity, scavenging free radicals, and inhibiting inflammatory responses.³²⁻³⁷ For example, *Panax notoginseng* and Chuanqiong are well-known traditional Chinese medicinal herbs. *Panax notoginseng* is particularly recognized for its ability to promote blood circulation. Its saponins exhibit calcium antagonist properties, preventing calcium ions (Ca^{2+}) influx by blocking exogenous Ca^{2+} entry and inhibiting intracellular Ca^{2+} release. This mechanism

contributes to immune regulation and enhances the activity of antioxidant enzymes such as superoxide dismutase and reduced lipid peroxide. The early application of *Panax notoginseng* saponins has significant antioxidant effects, increases spinal cord blood flow, improves gray matter necrosis after injury, and creates favorable conditions for white matter survival.

Chuanqiong inhibits the expression of nitric oxide synthase following spinal cord injury, thereby preventing cell damage or apoptosis caused by excessive nitric oxide. It also inhibits the expression of c-fos, Bax, and caspase-3 proteins, which are involved in secondary pathological changes after spinal cord injury. In addition, Chuanqiong enhances the mitochondria's ability to scavenge free radicals, reduces lipid peroxidation, and prevents the reduction in calcium-magnesium adenosine triphosphatase activity. This helps reduce mitochondrial membrane structure and function damage caused by oxygen-free radicals, maintain mitochondrial integrity, inhibit neuronal apoptosis at the subcellular level, and protect the injured spinal cord. Furthermore, amphotericin may provide protective effects by stabilizing cell membranes.³⁸ However, the specific mechanisms and clinical value of antioxidant therapy in RM still require further research and validation.

The occurrence of RM is closely associated with local vascular injury and ischemia. Improving blood supply can enhance the delivery of oxygen and nutrients, promote tissue repair, and support nerve regeneration, thereby contributing to functional recovery. Common medications used to improve blood supply include angiotensin-converting enzyme inhibitors such as ramipril³⁹; anticoagulants, such as heparin and warfarin⁴⁰; antiplatelet agents such as disulfiram⁴¹; anti-VEGF receptor agents⁴²; and valproic acid.⁴³ These drugs exert their effects through mechanisms, such as reducing inflammatory cell infiltration, decreasing VEGF expression, protecting vascular endothelial cells, repairing the BSCB, and promoting myelin regeneration. For example, Bevacizumab, an anti-VEGF agent, has been explored based on the hypothesis that radiation-induced endothelial damage and subsequent vascular permeability contribute to spinal cord edema and necrosis. By inhibiting VEGF, bevacizumab may reduce vascular permeability and alleviate edema.

Rehabilitation for RM includes various approaches – such as physiotherapy, occupational therapy, psychological rehabilitation, spinal cord electrical stimulation, traditional Chinese medicine rehabilitation, and hyperbaric oxygen therapy (HBOT). Studies have shown that exoskeleton gait training significantly improves walking ability in patients with spinal cord injuries,⁴⁴ while robot-assisted gait training offers systematic rehabilitation. The Lokomat

robot has been proven effective in enhancing daily living abilities and muscle strength, wearable robots improve walking ability, and end-effector robots are particularly effective in balance enhancement.⁴⁵ In recent years, virtual reality (VR) technology combined with physical therapy (e.g., VR gait training) has emerged as a research focus, demonstrating promising potential for promoting motor function recovery.⁴⁶ In addition, occupational therapy primarily utilizes wearables and smart aids (e.g., robotic gloves) to improve hand function and mobility in daily life.⁴⁷ Remote occupational therapy – which integrates intelligent devices and VR technology – improves treatment accessibility and effectiveness, reduces healthcare costs, and facilitates personalized care and psychological support. This approach holds promising application potential.⁴⁸ Psychological interventions aim to address depression and anxiety in RM patients, enhancing both mental health and rehabilitation outcomes.⁴⁹

Interventions based on positive thinking are gaining increasing attention, with studies demonstrating their effectiveness in improving sleep quality and reducing perceived stress.⁵⁰ Electrical spinal cord stimulation aids motor function recovery by activating spinal cord neural networks, increasing spinal cord excitability, and promoting neuroplasticity.⁵¹ Individualized neurostimulation programs, when combined with rehabilitation training, have been shown to significantly improve both motor and autonomic functions in patients.^{52,53} Traditional Chinese medicine rehabilitation includes acupuncture, tuina, and Chinese herbal medicine, which help improve blood circulation, regulate nerve function, and alleviate pain. Acupuncture promotes functional recovery by modulating central nervous system plasticity through stimulation of specific acupuncture points.⁵⁴ Modern acupuncture techniques, such as electroacupuncture, have shown greater efficacy in pain relief.⁵⁵ HBOT is believed to exert therapeutic effects by improving oxygen delivery to hypoxic tissues, stimulating angiogenesis, and promoting wound healing. In the context of RM, HBOT may help counteract radiation-induced vascular endothelial cell injury by promoting capillary regeneration and reducing tissue hypoxia. Furthermore, enhanced oxygenation may facilitate remyelination and repair of radiation-damaged spinal cord tissue, though further research is needed to fully understand its efficacy. Therefore, HBOT has the potential to target both major pathophysiological mechanisms, which are vascular compromise and demyelination.⁵⁶ Recent studies suggest that HBOT is effective in early-stage RM patients and may be combined with glucocorticoids and bevacizumab to enhance efficacy, although this remains to be validated in clinical trials.^{57,58}

3.2. Emerging treatments

3.2.1. Neuromodulation and brain-spinal cord interface (BSCI) technology

The brain-machine-spinal cord interface is a technology that connects the brain to the spinal cord, aiming to decode brain signals and transmit them in real-time to an external device. This enables control of muscle movements using a neuromuscular electrical stimulation system, thereby improving motor function and enhancing patients' quality of life.⁵⁹ Recent studies have demonstrated that real-time control of muscle activation through cortical signaling can facilitate motor function recovery in patients with mechanical spinal cord injury.⁶⁰ However, the brain-machine-spinal cord interface system primarily targets functional rehabilitation, and research on its application in neurorehabilitation remains limited.

Recently, a new system called the BSCI has been proposed. This technique establishes a digital bridge directly between the brain and the spinal cord, aiming to restore the natural walking ability of patients paralyzed due to mechanical spinal cord injury. Studies have shown that with the BSCI, patients can regain neurological function and walk independently, even after the system is turned off.⁶¹ The BSCI technique holds promise for restoring ambulation in a broader group of spinal cord injury patients and is expected to have expanded applications in the treatment of RM and other neurological disorders.

3.2.2. Stem cell transplantation and tissue engineering therapy

Stem cell transplantation therapy for RM aims to promote neural repair and functional recovery by introducing stem cells into the damaged spinal cord region. The underlying mechanisms may involve various processes, including neuroprotection, immunomodulation, axon regeneration, synapse formation, myelin sheath formation, and angiogenesis, all of which contribute to spinal cord tissue repair and functional recovery. The primary cell types used in stem cell transplantation for spinal cord injury include Schwann cells, neural stem and progenitor cells, OPCs, olfactory ensheathing cells (OECs), and mesenchymal stem cells (MSCs). Schwann cells – which are myelin-forming glial cells of the peripheral nervous system – guide axon regeneration and support both axonal and myelin repair after transplantation. Neural stem and progenitor cells are pluripotent stem cells that can differentiate at the site of spinal cord injury into various cell types – including neurons, astrocytes, OLs, and OPCs – thereby promoting axon regeneration, myelin formation, and the restoration of neural connections. OPCs, also known as OL precursor cells, facilitate myelin

regeneration at the injury site, thereby restoring nerve conduction. OECs are glial cells that support the growth of olfactory receptor axons into the olfactory bulb. When transplanted into the injured spinal cord, OECs promote axonal regeneration and help reduce glial scar formation. MSCs are pluripotent progenitor cells found in various tissues and capable of self-renewal and differentiation into multiple cell types. After transplantation, MSCs may promote spinal cord injury repair, reduce inflammation, and mitigate scar formation by secreting neurotrophic and anti-inflammatory factors. Transplantable cells can be derived from adult or embryonic pluripotent stem cells through induced differentiation,⁶² or generated using direct transformation techniques.⁶³ Furthermore, their mechanisms of action may vary depending on the origin of allogeneic transplanted cells (Table 2).

The primary objectives of stem cell therapy for RM are to restore vascular function, inhibit fibrosis and chronic inflammation, and promote nerve regeneration. Vascular regeneration typically involves MSCs or endothelial progenitor cells, which possess vascular regenerative capabilities, as well as stem cells (e.g., NSCs or genetically modified stem cells) that secrete neurotrophic factors, such as brain-derived neurotrophic factor and VEGF. MSCs are the preferred choice for RM treatment due to their crucial roles in angiogenesis and antifibrosis.^{94,95} MSCs can secrete factors that regulate the gene expression in endothelial cells, thereby promoting vascular integrity recovery and repairing the BSCB. The interaction between endothelial

cells and pericytes is essential for the structural and functional maintenance of the BSCB following spinal cord injury. When pericytes separate from endothelial cells, it leads to BSCB destruction. MSCs transplantation has been shown to enhance pericyte coverage to endothelial cells, which helps promote BSCB repair by secreting bioactive molecules that stimulate pericyte recruitment and proliferation.⁹⁶ In contrast, NSCs, OPCs, OECs, and Schwann cells are more suitable for treating mechanical spinal cord injuries, as their primary therapeutic mechanism involves reducing secondary damage and promoting nerve regeneration. These cells are widely used in mechanical spinal cord injury and are expected to be applied to RM as well.

Fang *et al.*⁹⁷ developed a modified MSC therapy known as MSC-MM@LPHN, aimed at restoring the vascular microenvironment in a pulmonary fibrosis model. The therapy utilizes the inherent homing ability of MSCs to target lung tissue, promoting the dedifferentiation of myofibroblasts. This process reduces cytokine secretion responsible for endothelial cell damage, prevents endothelial cell transformation into a fibrotic phenotype, and ultimately restores vascular endothelial cell function. This approach is expected to be applicable in treating radiation-induced spinal cord fibrosis. Yusoff *et al.*⁹⁸ demonstrated that hypoxic pre-conditioning of MSCs enhances their proliferation and activity while increasing the expression of VEGF, CD31, HIF-1 α , and other factors. This process effectively promotes angiogenesis, as well as

Table 2. Possible mechanisms of action of transplanted cells from different origins and relevant references

Cell types	Origins	Possible mechanisms of action	References
Schwann cells	Skin pre-cursor cells	Neuroprotection, myelin formation, and axonal regeneration.	64,65
	Sciatic nerve	Neuroprotection, myelin formation, and axonal regeneration.	66,67
Neural stem and progenitor cells	Spinal cord	Neuroprotection, myelin formation, and axonal regeneration.	68-71
	Brain	Immunomodulation and synapse formation.	
Oligodendrocyte progenitor cells	Stem cells	Neuroprotection, myelin formation, and axonal regeneration.	72-75
	Peripheral blood mononuclear cells	Neuroprotection, myelin formation, and axonal regeneration.	76,77
Olfactory ensheathing cells	Peripheral blood mononuclear cells	Immunomodulation and synapse formation.	78
	Olfactory bulb	Neuroprotection and myelin formation.	79-83
	Lamina propria of spinal cord	Reduced inflammation and angiogenesis.	
	Lamina propria of olfactory mucosa	Axonal regeneration.	84
Mesenchymal stem cells	Tibial/femoral bone marrow	Axonal regeneration.	85
	Pelvic bone marrow	Neuroprotection, myelin formation, and axonal regeneration.	86-89
	Wharton's jelly of the umbilical cord	Neuroprotection and immunomodulation.	90,91
		Angiogenesis and immunomodulation.	92,93

Note: The mechanisms discussed in this review are limited to the main mechanisms proposed by most of the present studies, including: Neuroprotection, immunomodulation, axonal regeneration, synapse formation, myelin formation, attenuation of inflammation, and vascular regeneration.

improves blood flow and blood perfusion. The engineering of MSCs and other cell types to enhance their specific capabilities shows significant potential and warrants further investigation.

Regarding transplantation timing and methods, early intervention is crucial for inhibiting progressive fibrosis and preventing chronic damage. Intravenous or intrathecal injections are commonly used to ensure widespread coverage of the affected area. In terms of microenvironment regulation, stem cell transplantation for radiation injury often requires the use of antifibrotic drugs (e.g., pirfenidone)⁹⁹ or antioxidants (e.g., edaravone)¹⁰⁰ to improve stem cell survival. In addition, targeted inhibition of pro-fibrotic and inflammatory pathways, such as transforming growth factor β and nuclear factor κ B, is also crucial. For RM, the main objective of stem cell therapy is to slow disease progression and improve function; however, achieving full recovery remains challenging. Since radiation injury is a chronic and progressive condition, therapeutic effects tend to be gradual.

Tissue engineering therapy for spinal cord injury integrates stem cells, biomaterials, and bioactive factors to promote nerve regeneration, repair myelin, exert anti-inflammatory effects, and enhance the microenvironment. A key component of this approach is the use of scaffolds that provide structural support, facilitate stem cell differentiation into neurons and glial cells, and regulate the release of neurotrophic factors to restore the neural network. Despite challenges, such as low cell survival, poor biocompatibility, and difficulty in clinical translation, technological advancements show promising potential for functional recovery. Present research is focusing on controlled-release systems for biomaterials and bioactive factors, which are increasingly being applied in the treatment of mechanical spinal cord injuries. Chen *et al.*¹⁰¹ developed an injectable hydrogel that responds to early reactive oxygen species and late matrix metalloproteinases, enabling precise, on-demand drug delivery to the injury site. The accumulation of matrix metalloproteinases in the late stage of spinal cord injury can promote angiogenesis and NSC differentiation by triggering the release of VEGF, which can effectively resist the adverse effects caused by vascular endothelial cell injury.

For example, in the field of biomaterials, hydrogels¹⁰² and decellularized spinal cord scaffolds have been developed with mechanical properties that closely mimic those of damaged spinal cord tissue. These materials support neural regeneration and functional recovery by providing mechanical stability and fostering a neuroregenerative microenvironment following mechanical spinal cord injury.¹⁰³ In terms of controlled release systems for

bioactive factors, some studies have developed gamma-aminobutyric acid-ergic neuron nanomedicine, which has been shown to effectively regulate neuronal excitability, protect residual neural tissues, and promote functional recovery after spinal cord injury. This approach has emerged as an innovative method for enhancing functional recovery.¹⁰⁴ In addition, a highly drug-loaded microsphere technology has been developed using a nano-microsphere structure. This method significantly enhances drug loading capacity and therapeutic efficiency, reduces the frequency of administration and excipient usage, and consequently minimizes side effects.¹⁰⁵ Nanodelivery systems with controlled release characteristics play a crucial role in targeting spinal cord tissues and addressing the limitations of conventional drug delivery methods through strategic design and modification. However, tissue engineering therapy is currently primarily applied to mechanical spinal cord injuries, and its application and efficacy in RM still require further exploration.

3.2.3. Other emerging treatments

The treatment of RM is transitioning from single-modality interventions to multimodal integrated approaches, with numerous emerging treatments that still require further research and validation. Gene therapy aims to restore function by precisely regulating gene expression. For instance, viral vectors can be used to deliver neurotrophic factor genes (e.g., brain-derived neurotrophic factor and *NT-3*) to the injury site, thereby activating neuronal regeneration pathways and inhibiting apoptosis. Recent studies have demonstrated that gene editing techniques, such as clustered regularly interspaced short palindromic repeats-associated protein 9 (commonly known as CRISPR-Cas9), have successfully repaired spinal cord injury-related gene mutations in animal models; however, their clinical applicability still requires further validation.¹⁰⁶

Anti-inflammatory and immunomodulatory therapies aim to break the vicious cycle of chronic inflammation. New anti-inflammatory agents, such as monoclonal antibodies targeting IL-6 and small-molecule Janus kinase inhibitors, are currently undergoing clinical trials to attenuate secondary injury by blocking pro-inflammatory cytokine signaling pathways. Kong¹⁰⁷ reported that the use of poly lactic-co-glycolic acid scaffolds loaded with immunomodulators (e.g., fingolimod) and NSCs was effective in restoring locomotor ability and promoting the formation of new neurons in spinal cord-injured rats.

Metabolic intervention strategies aim to remodel microenvironmental homeostasis at the energy supply level. A study by Minhas *et al.*¹⁰⁸ revealed that indoleamine 2,3-dioxygenase 1 inhibitors (e.g., Epcadostat) restore

astrocyte glucose metabolism and reverse neuronal energy crises, showing significant effects in Alzheimer's disease models. These potential mechanisms could be extended to RM therapy. The development of targeted drugs is based on molecular mechanisms to enable personalized treatment. For example, inhibition of the transforming growth factor β signaling pathway reduces fibrotic scar formation,¹⁰⁹ and drugs targeting the glial cell water channel protein aquaporin-4 (e.g., TGN-020) alleviate radiation-induced edema.^{110,111}

While these advances are still in pre-clinical or early clinical stages, they have demonstrated promising synergistic potential. However, their safety and translational value require further validation through interdisciplinary studies in the future. Most of the emerging therapies discussed in this section are still under investigation in pre-clinical models, with limited data available on their optimal dosing or dose-response relationships.

4. Conclusion and outlook

RM, a severe late complication of radiotherapy, presents a significant challenge to present treatment strategies due to its complex pathological mechanisms and the progressive nature of injury. Traditional treatments – such as glucocorticoids and immunoglobulins – are effective in alleviating acute symptoms and controlling secondary inflammation, but they are insufficient for achieving substantial recovery of neurological function in the advanced stages of the disease. In recent years, emerging therapeutic approaches, including neuromodulation techniques, stem cell transplantation, tissue engineering, biomaterials, and gene therapy, have made notable advancements, offering new directions for RM treatment. These strategies have demonstrated promising potential through multiple mechanisms – including neuroprotection, myelin regeneration, axonal repair, immunomodulation, antifibrosis, and angiogenesis – that work synergistically.

However, several challenges remain in the treatment of RM, including low stem cell survival rates, the biocompatibility of tissue-engineered materials, treatment safety, and long-term efficacy. In addition, the optimization of multimodal combination therapies requires further investigation, and the development of personalized treatment strategies must be supported by more robust clinical data. Future research should focus on several key areas, including: (i) conducting in-depth analyses of the pathological mechanisms of RM to identify key therapeutic targets; (ii) optimizing the application of stem cell transplantation, nanodelivery systems, and gene editing technologies to enhance therapeutic efficacy; (iii) advancing neuromodulation technologies and tissue

engineering scaffolds to promote nerve regeneration and functional recovery; and (iv) validating the safety and efficacy of emerging therapies through large-scale clinical trials.

In conclusion, with ongoing advancements in basic research and clinical technology, the treatment of RM is shifting from single-modality interventions to multimodal integrated approaches. Across interdisciplinary collaboration, the integration of precision medicine, and the application of advanced technologies, significant recovery of neurological function in RM patients may become achievable – ultimately offering practical solutions to enhance patients' quality of life.

Acknowledgments

None.

Funding

This review was supported by the Hunan Provincial Natural Science Foundation of China (2024JJ9488)

Conflict of interest

The authors declare they have no competing interests.

Author contributions

Conceptualization: Limin Liu

Writing – original draft: Yuhang Yu, Yukai Tang, and Shengyi Liu

Writing – review & editing: Limin Liu

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of data

Material for this article is available from the corresponding author upon reasonable request.

References

- Schultheiss TE. The radiation dose-response of the human spinal cord. *Int J Radiat Oncol Biol Phys.* 2008;71(5):1455-1459.
doi: 10.1016/j.ijrobp.2007.11.075
- Jackson CB, Boe LA, Zhang L, *et al.* Radiation myelitis risk after hypofractionated spine stereotactic body radiation therapy. *JAMA Oncol.* 2025;11(2):128-134.
doi: 10.1001/jamaoncol.2024.5387

3. Wong CS, Fehlings MG, Sahgal A. Pathobiology of radiation myelopathy and strategies to mitigate injury. *Spinal Cord*. 2015;53(8):574-580.
doi: 10.1038/sc.2015.43
4. Schultheiss TE, Stephens LC, Maor MH. Analysis of the histopathology of radiation myelopathy. *Int J Radiat Oncol Biol Phys*. 1988;14(1):27-32.
doi: 10.1016/0360-3016(88)90046-6
5. Li YQ, Chen P, Haimovitz-Friedman A, Reilly RM, Wong CS. Endothelial apoptosis initiates acute blood-brain barrier disruption after ionizing radiation. *Cancer Res*. 2003;63(18):5950-5956.
6. Peña LA, Fuks Z, Kolesnick RN. Radiation-induced apoptosis of endothelial cells in the murine central nervous system: Protection by fibroblast growth factor and sphingomyelinase deficiency. *Cancer Res*. 2000;60(2):321-327.
7. Murrell DH, Zarghami N, Jensen MD, Chambers AF, Wong E, Foster PJ. Evaluating Changes to blood-brain barrier integrity in brain metastasis over time and after radiation treatment. *Transl Oncol*. 2016;9(3):219-227.
doi: 10.1016/j.tranon.2016.04.006
8. Stewart PA, Vinters HV, Wong CS. Blood-spinal cord barrier function and morphometry after single doses of x-rays in rat spinal cord. *Int J Radiat Oncol Biol Phys*. 1995;32(3):703-711.
doi: 10.1016/0360-3016(94)00594-B
9. Li YQ, Aubert I, Wong CS. Abrogation of early apoptosis does not alter late inhibition of hippocampal neurogenesis after irradiation. *Int J Radiat Oncol Biol Phys*. 2010;77(4):1213-1222.
doi: 10.1016/j.ijrobp.2010.01.015
10. Chow BM, Li YQ, Wong CS. Radiation-induced apoptosis in the adult central nervous system is p53-dependent. *Cell Death Differ*. 2000;7(8):712-720.
doi: 10.1038/sj.cdd.4400704
11. Atkinson SL, Li YQ, Wong CS. Apoptosis and proliferation of oligodendrocyte progenitor cells in the irradiated rodent spinal cord. *Int J Radiat Oncol Biol Phys*. 2005;62(2):535-554.
doi: 10.1016/j.ijrobp.2005.01.061
12. Hawryluk GW, Fehlings MG. The center of the spinal cord may be central to its repair. *Cell Stem Cell*. 2008;3(3):230-232.
doi: 10.1016/j.stem.2008.08.009
13. Tsao MN, Li YQ, Lu G, Xu Y, Wong CS. Upregulation of vascular endothelial growth factor is associated with radiation-induced blood-spinal cord barrier breakdown. *J Neuropathol Exp Neurol*. 1999;58(10):1051-1060.
doi: 10.1097/00005072-199910000-00003
14. Li YQ, Ballinger JR, Nordal RA, Su ZF, Wong CS. Hypoxia in radiation-induced blood-spinal cord barrier breakdown. *Cancer Res*. 2001;61(8):3348-3354.
15. Long HQ, Li GS, Cheng X, Xu JH, Li FB. Role of hypoxia-induced VEGF in blood-spinal cord barrier disruption in chronic spinal cord injury. *Chin J Traumatol*. 2015;18(5):293-295.
doi: 10.1016/j.cjtee.2015.08.004
16. Vaquero J, Zurita M, De Oya S, Coca S. Vascular endothelial growth/permeability factor in spinal cord injury. *J Neurosurg*. 1999;90 2 Suppl:220-223.
doi: 10.3171/spi.1999.90.2.0220
17. Benton RL, Whitemore SR. VEGF165 therapy exacerbates secondary damage following spinal cord injury. *Neurochem Res*. 2003;28(11):1693-1703.
doi: 10.1023/a:1026013106016
18. Widenfalk J, Lipson A, Jubran M, *et al*. Vascular endothelial growth factor improves functional outcome and decreases secondary degeneration in experimental spinal cord contusion injury. *Neuroscience*. 2003;120(4):951-960.
doi: 10.1016/s0306-4522(03)00399-3
19. Xiaowei H, Ninghui Z, Wei X, Yiping T, Linfeng X. The experimental study of hypoxia-inducible factor-1alpha and its target genes in spinal cord injury. *Spinal Cord*. 2006;44(1):35-43.
doi: 10.1038/sj.sc.3101813
20. Kao CH, Chen SH, Chio CC, Lin MT. Human umbilical cord blood-derived CD34+ cells may attenuate spinal cord injury by stimulating vascular endothelial and neurotrophic factors. *Shock*. 2008;29(1):49-55.
doi: 10.1097/shk.0b013e31805cddce
21. Facchiano F, Fernandez E, Mancarella S, *et al*. Promotion of regeneration of corticospinal tract axons in rats with recombinant vascular endothelial growth factor alone and combined with adenovirus coding for this factor. *J Neurosurg*. 2002;97(1):161-168.
doi: 10.3171/jns.2002.97.1.0161
22. Sakanaka M, Zhu P, Zhang B, *et al*. Intravenous infusion of dihydrogeninoside Rb1 prevents compressive spinal cord injury and ischemic brain damage through upregulation of VEGF and Bcl-XL. *J Neurotrauma*. 2007;24(6):1037-1054.
doi: 10.1089/neu.2006.0182
23. Choi UH, Ha Y, Huang X, *et al*. Hypoxia-inducible expression of vascular endothelial growth factor for the treatment of spinal cord injury in a rat model. *J Neurosurg Spine*. 2007;7(1):54-60.
doi: 10.3171/SPI-07/07/054
24. Fehlings MG, Wilson JR, *et al*. A clinical practice guideline for the management of patients with acute spinal cord injury: Recommendations on the use of methylprednisolone sodium succinate. *Global Spine J*. 2017;7(3 Suppl):203S-211S.

- doi: 10.1177/2192568217703085
25. Marchioni E, Marinou-Aktipi K, Uggetti C, *et al.* Effectiveness of intravenous immunoglobulin treatment in adult patients with steroid-resistant monophasic or recurrent acute disseminated encephalomyelitis. *J Neurol.* 2002;249(1):100-104.
doi: 10.1007/pl00007836
 26. Naghavi S, Motahharynia A, Fatemi F, Ahmadi E, Mokhtari F, Adibi I. The benefit of intravenous immune globulin in the treatment of delayed radiation myelopathy. *Strahlenther Onkol.* 2024;200(9):827-831.
doi: 10.1007/s00066-023-02150-1
 27. Picca A, Berzero G, Bihan K, *et al.* Longitudinally extensive myelitis associated with immune checkpoint inhibitors. *Neurol Neuroimmunol Neuroinflamm.* 2021;8(3):e967.
doi: 10.1212/NXI.0000000000000967
 28. Möhn N, Beutel G, Gutzmer R, Ivanyi P, Satzger I, Skripuletz T. Neurological immune related adverse events associated with nivolumab, ipilimumab, and pembrolizumab therapy-review of the literature and future outlook. *J Clin Med.* 2019;8(11):1777.
doi: 10.3390/jcm8111777
 29. Chang VA, Simpson DR, Daniels GA, Piccioni DE. Infliximab for treatment-refractory transverse myelitis following immune therapy and radiation. *J Immunother Cancer.* 2018;6(1):153.
doi: 10.1186/s40425-018-0471-2
 30. Owen T, Fung AS. Combination intravenous immune globulin (IVIG) and high dose steroids for treatment of immune-related myelitis in a non-small cell lung cancer patient treated with pembrolizumab and palliative radiation treatment: A case report. *Clin Lung Cancer.* 2022;23(8):e563-e567.
doi: 10.1016/j.clcc.2022.08.012
 31. Zhang Y, Zou Z, Liu S, *et al.* Edaravone-loaded poly(amino acid) nanogel inhibits ferroptosis for neuroprotection in cerebral ischemia injury. *Asian J Pharm Sci.* 2024;19(2):100886.
doi: 10.1016/j.ajps.2024.100886
 32. Lu ZX, Wang JH, Xu N, Luan J, Pang XY, Xia YJ. Protective effect of compound raspberry seed powder on radiation-induced spinal cord injury in mice. *J Radiat Res Radiat Process.* 2020;38(1):46-53.
doi: 10.11889/j.1000-3436.2020.rrj.38.010303
 33. Zhang XY, Zhou P, Zhu C, Chu XD. Effects of Panax ginseng and Chuanxiong on behavioral changes and axonal regeneration after spinal cord injury. *Chin J Ethnomed Ethnopharmacol.* 2009;18(20):4-5.
 34. Enginar H, Cemek M, Karaca T, Unak P. Effect of grape seed extract on lipid peroxidation, antioxidant activity and peripheral blood lymphocytes in rats exposed to x-radiation. *Phytother Res.* 2007;21(11):1029-1035.
doi: 10.1002/ptr.2201
 35. Yalinkilic O, Enginar H. Effect of X-radiation on lipid peroxidation and antioxidant systems in rats treated with saponin-containing compounds. *Photochem Photobiol.* 2008;84(1):236-242.
doi: 10.1111/j.1751-1097.2007.00240.x
 36. Deger Y, Dede S, Belge A, Mert N, Kahraman T, Alkan M. Effects of X-ray radiation on lipid peroxidation and antioxidant systems in rabbits treated with antioxidant compounds. *Biol Trace Elem Res.* 2003;94(2):149-156.
doi: 10.1385/BTER.94:2:149
 37. Peker S, Abacioglu U, Sun I, Konya D, Yüksel M, Pamir NM. Prophylactic effects of magnesium and vitamin E in rat spinal cord radiation damage: Evaluation based on lipid peroxidation levels. *Life Sci.* 2004;75(12):1523-1530.
doi: 10.1016/j.lfs.2004.05.003
 38. Nieder C, Andratschke NH, Wiedenmann N, Molls M. Prevention of radiation-induced central nervous system toxicity: A role for amifostine? *Anticancer Res.* 2004;24(6):3803-3809.
 39. Saager M, Hahn EW, Peschke P, *et al.* Ramipril reduces incidence and prolongates latency time of radiation-induced rat myelopathy after photon and carbon ion irradiation. *J Radiat Res.* 2020;61(5):791-798.
doi: 10.1093/jrr/rraa042
 40. Glantz MJ, Burger PC, Friedman AH, Radtke RA, Massey EW, Schold SC Jr. Treatment of radiation-induced nervous system injury with heparin and warfarin. *Neurology.* 1994;44(11):2020-2027.
doi: 10.1212/wnl.44.11.2020
 41. Hornsey S, Myers R, Jenkinson T. The reduction of radiation damage to the spinal cord by post-irradiation administration of vasoactive drugs. *Int J Radiat Oncol Biol Phys.* 1990;18(6):1437-1442.
doi: 10.1016/0360-3016(90)90319-f
 42. Psimaras D, Tafani C, Ducray F, *et al.* Bevacizumab in late-onset radiation-induced myelopathy. *Neurology.* 2016;86(5):454-457.
doi: 10.1212/WNL.0000000000002345
 43. Lee JY, Kim HS, Choi HY, Oh TH, Ju BG, Yune TY. Valproic acid attenuates blood-spinal cord barrier disruption by inhibiting matrix metalloprotease-9 activity and improves functional recovery after spinal cord injury. *J Neurochem.* 2012;121(5):818-829.
doi: 10.1111/j.1471-4159.2012.07731.x

44. Gil-Agudo Á, Megía-García Á, Pons JL, *et al.* Correction: Exoskeleton-based training improves walking independence in incomplete spinal cord injury patients: Results from a randomized controlled trial. *J Neuroeng Rehabil.* 2023;20(1):160.
doi: 10.1186/s12984-023-01281-x
45. Park JM, Kim YW, Lee SJ, Shin JC. Robot-assisted gait training in individuals with spinal cord injury: A systematic review and meta-analysis of randomized controlled trials. *Ann Rehabil Med.* 2024;48(3):171-191.
doi: 10.5535/arm.230039
46. Villiger M, Bohli D, Kiper D, *et al.* Virtual reality-augmented neurorehabilitation improves motor function and reduces neuropathic pain in patients with incomplete spinal cord injury. *Neurorehabil Neural Repair.* 2013;27(8):675-683.
doi: 10.1177/1545968313490999
47. Demolder C, Molina A, Hammond FL 3rd, Yeo WH. Recent advances in wearable biosensing gloves and sensory feedback biosystems for enhancing rehabilitation, prostheses, healthcare, and virtual reality. *Biosens Bioelectron.* 2021;190:113443.
doi: 10.1016/j.bios.2021.113443
48. Lee S, Kim J, Kim J. Substantiating clinical effectiveness and potential barriers to the widespread implementation of spinal cord injury telerehabilitation: A systematic review and qualitative synthesis of randomized trials in the recent past decade. *Telemed Rep.* 2021;2(1):64-77.
doi: 10.1089/tmr.2020.0026
49. Craig A, Tran Y, Middleton J. Psychological morbidity and spinal cord injury: A systematic review. *Spinal Cord.* 2009;47(2):108-114.
doi: 10.1038/sc.2008.115
50. Cao Y, Wu H, Shi S, Xie D. Effects of mindfulness-based stress reduction therapy for sleep quality and perceived stress in patients with spinal cord injury. *Explore (NY).* 2024;20(5):103037.
doi: 10.1016/j.explore.2024.103037
51. Harkema S, Gerasimenko Y, Hodes J, *et al.* Effect of epidural stimulation of the lumbosacral spinal cord on voluntary movement, standing, and assisted stepping after motor complete paraplegia: A case study. *Lancet.* 2011;377(9781):1938-1947.
doi: 10.1016/S0140-6736(11)60547-3
52. Angeli CA, Boakye M, Morton RA, *et al.* Recovery of over-ground walking after chronic motor complete spinal cord injury. *N Engl J Med.* 2018;379(13):1244-1250.
doi: 10.1056/NEJMoa1803588
53. Wagner FB, Mignardot JB, Le Goff-Mignardot CG, *et al.* Targeted neurotechnology restores walking in humans with spinal cord injury. *Nature.* 2018;563(7729):65-71.
doi: 10.1038/s41586-018-0649-2
54. Cheng KJ. Neurobiological mechanisms of acupuncture for some common illnesses: A clinician's perspective. *J Acupunct Meridian Stud.* 2014;7(3):105-114.
doi: 10.1016/j.jams.2013.07.008
55. He K, Hu R, Huang Y, Qiu B, Chen Q, Ma R. Effects of acupuncture on neuropathic pain induced by spinal cord injury: A systematic review and meta-analysis. *Evid Based Complement Alternat Med.* 2022;2022:6297484.
doi: 10.1155/2022/6297484
56. Zhou Y, Liu XH, Qu SD, *et al.* Hyperbaric oxygen intervention on expression of hypoxia-inducible factor-1 α and vascular endothelial growth factor in spinal cord injury models in rats. *Chin Med J (Engl).* 2013;126(20):3897-3903.
57. Fernández E, Morillo V, Salvador M, *et al.* Hyperbaric oxygen and radiation therapy: A review. *Clin Transl Oncol.* 2021;23(6):1047-1053.
doi: 10.1007/s12094-020-02513-5
58. Bodensohn R, Haehl E, Belka C, Niyazi M. Fractionated radiotherapy for spinal tumors: A literature review regarding spinal glioma, ependymoma, and meningioma. *Neurooncol Adv.* 2024;6(Suppl 3):3101-3109.
doi: 10.1093/noajnl/vdad158
59. Lobel DA, Lee KH. Brain machine interface and limb reanimation technologies: Restoring function after spinal cord injury through development of a bypass system. *Mayo Clin Proc.* 2014;89(5):708-714.
doi: 10.1016/j.mayocp.2014.02.003
60. Bouton CE, Shaikhouni A, Annetta NV, *et al.* Restoring cortical control of functional movement in a human with quadriplegia. *Nature.* 2016;533(7602):247-250.
doi: 10.1038/nature17435
61. Lorach H, Galvez A, Spagnolo V, *et al.* Walking naturally after spinal cord injury using a brain-spine interface. *Nature.* 2023;618(7963):126-133.
doi: 10.1038/s41586-023-06094-5
62. Lu P, Woodruff G, Wang Y, *et al.* Long-distance axonal growth from human induced pluripotent stem cells after spinal cord injury. *Neuron.* 2014;83(4):789-796.
doi: 10.1016/j.neuron.2014.07.014
63. Yang N, Zuchero JB, Ahlenius H, *et al.* Generation of oligodendroglial cells by direct lineage conversion. *Nat Biotechnol.* 2013;31(5):434-439.
doi: 10.1038/nbt.2564
64. Sparling JS, Bretzner F, Biernaskie J, *et al.* Schwann cells generated from neonatal skin-derived precursors or neonatal peripheral nerve improve functional recovery

- after acute transplantation into the partially injured cervical spinal cord of the rat. *J Neurosci*. 2015;35(17):6714-6730.
doi: 10.1523/JNEUROSCI.1070-14.2015
65. Biernaskie J, Sparling JS, Liu J, *et al*. Skin-derived precursors generate myelinating Schwann cells that promote remyelination and functional recovery after contusion spinal cord injury. *J Neurosci*. 2007;27(36):9545-9559.
doi: 10.1523/JNEUROSCI.1930-07.2007
66. Takami T, Oudega M, Bates ML, Wood PM, Kleitman N, Bunge MB. Schwann cell but not olfactory ensheathing glia transplants improve hindlimb locomotor performance in the moderately contused adult rat thoracic spinal cord. *J Neurosci*. 2002;22(15):6670-6681.
doi: 10.1523/JNEUROSCI.22-15-06670.200
67. Williams RR, Henao M, Pearse DD, Bunge MB. Permissive Schwann cell graft/spinal cord interfaces for axon regeneration. *Cell Transplant*. 2015;24(1):115-131.
doi: 10.3727/096368913X674657
68. Hawryluk GW, Mothe A, Wang J, Wang S, Tator C, Fehlings MG. An *in vivo* characterization of trophic factor production following neural precursor cell or bone marrow stromal cell transplantation for spinal cord injury. *Stem Cells Dev*. 2012;21(12):2222-2238.
doi: 10.1089/scd.2011.0596
69. Cao Q, Xu XM, Devries WH, *et al*. Functional recovery in traumatic spinal cord injury after transplantation of multineurotrophin-expressing glial-restricted precursor cells. *J Neurosci*. 2005;25(30):6947-6957.
doi: 10.1523/JNEUROSCI.1065-05.2005
70. Cao Q, He Q, Wang Y, *et al*. Transplantation of ciliary neurotrophic factor-expressing adult oligodendrocyte precursor cells promotes remyelination and functional recovery after spinal cord injury. *J Neurosci*. 2010;30(8):2989-3001.
doi: 10.1523/JNEUROSCI.3174-09.2010
71. Lu P, Wang Y, Graham L, *et al*. Long-distance growth and connectivity of neural stem cells after severe spinal cord injury. *Cell*. 2012;150(6):1264-1273.
doi: 10.1016/j.cell.2012.08.020
72. Hawryluk GW, Spano S, Chew D, *et al*. An examination of the mechanisms by which neural precursors augment recovery following spinal cord injury: A key role for remyelination. *Cell Transplant*. 2014;23(3):365-380.
doi: 10.3727/096368912X662408
73. Plemel JR, Chojnacki A, Sparling JS, *et al*. Platelet-derived growth factor-responsive neural precursors give rise to myelinating oligodendrocytes after transplantation into the spinal cords of contused rats and dysmyelinated mice. *Glia*. 2011;59(12):1891-1910.
doi: 10.1002/glia.21232
74. Yasuda A, Tsuji O, Shibata S, *et al*. Significance of remyelination by neural stem/progenitor cells transplanted into the injured spinal cord. *Stem Cells*. 2011;29(12):1983-1994.
doi: 10.1002/stem.767
75. Cusimano M, Biziato D, Brambilla E, *et al*. Transplanted neural stem/precursor cells instruct phagocytes and reduce secondary tissue damage in the injured spinal cord. *Brain*. 2012;135(Pt 2):447-460.
doi: 10.1093/brain/awr339
76. Sharp J, Frame J, Siegenthaler M, Nistor G, Keirstead HS. Human embryonic stem cell-derived oligodendrocyte progenitor cell transplants improve recovery after cervical spinal cord injury. *Stem Cells*. 2010;28(1):152-163.
doi: 10.1002/stem.245
77. Keirstead HS, Nistor G, Bernal G, *et al*. Human embryonic stem cell-derived oligodendrocyte progenitor cell transplants remyelinate and restore locomotion after spinal cord injury. *J Neurosci*. 2005;25(19):4694-4705.
doi: 10.1523/JNEUROSCI.0311-05.2005
78. All AH, Gharibani P, Gupta S, *et al*. Early intervention for spinal cord injury with human induced pluripotent stem cells oligodendrocyte progenitors. *PLoS One*. 2015;10(1):e0116933.
doi: 10.1371/journal.pone.0116933
79. López-Vales R, García-Álías G, Forés J, Navarro X, Verdú E. Increased expression of cyclo-oxygenase 2 and vascular endothelial growth factor in lesioned spinal cord by transplanted olfactory ensheathing cells. *J Neurotrauma*. 2004;21(8):1031-1043.
doi: 10.1089/0897715041651105
80. Barbour HR, Plant CD, Harvey AR, Plant GW. Tissue sparing, behavioral recovery, supraspinal axonal sparing/regeneration following sub-acute glial transplantation in a model of spinal cord contusion. *BMC Neurosci*. 2013;14:106.
doi: 10.1186/1471-2202-14-106
81. Takeoka A, Jindrich DL, Muñoz-Quiles C, *et al*. Axon regeneration can facilitate or suppress hindlimb function after olfactory ensheathing glia transplantation. *J Neurosci*. 2011;31(11):4298-4310.
doi: 10.1523/JNEUROSCI.4967-10.2011
82. Fouad K, Schnell L, Bunge MB, Schwab ME, Liebscher T, Pearse DD. Combining schwann cell bridges and olfactory-ensheathing glia grafts with chondroitinase promotes locomotor recovery after complete transection of the spinal cord. *J Neurosci*. 2005;25(5):1169-1178.
doi: 10.1523/JNEUROSCI.3562-04.200
83. Barakat DJ, Gaglani SM, Neravetla SR, *et al*. Survival,

- integration, and axon growth support of glia transplanted into the chronically contused spinal cord. *Cell Transplant.* 2005;14(4):225-240.
doi: 10.3727/000000005783983106
84. Richter MW, Fletcher PA, Liu J, Tetzlaff W, Roskams AJ. Lamina propria and olfactory bulb ensheathing cells exhibit differential integration and migration and promote differential axon sprouting in the lesioned spinal cord. *J Neurosci.* 2005;25(46):10700-10711.
doi: 10.1523/JNEUROSCI.3632-05.2005
85. Ramer LM, Au E, Richter MW, Liu J, Tetzlaff W, Roskams AJ. Peripheral olfactory ensheathing cells reduce scar and cavity formation and promote regeneration after spinal cord injury. *J Comp Neurol.* 2004;473(1):1-15.
doi: 10.1002/cne.20049
86. Gu W, Zhang F, Xue Q, Ma Z, Lu P, Yu B. Transplantation of bone marrow mesenchymal stem cells reduces lesion volume and induces axonal regrowth of injured spinal cord. *Neuropathology.* 2010;30(3):205-217.
doi: 10.1111/j.1440-1789.2009.01063.x
87. Ritfeld GJ, Patel A, Chou A, et al. The role of brain-derived neurotrophic factor in bone marrow stromal cell-mediated spinal cord repair. *Cell Transplant.* 2015;24(11):2209-2220.
doi: 10.3727/096368915X686201
88. Lu P, Blesch A, Graham L, et al. Motor axonal regeneration after partial and complete spinal cord transection. *J Neurosci.* 2012;32(24):8208-8218.
doi: 10.1523/JNEUROSCI.0308-12.2012
89. Lu P, Yang H, Jones LL, Filbin MT, Tuszynski MH. Combinatorial therapy with neurotrophins and cAMP promotes axonal regeneration beyond sites of spinal cord injury. *J Neurosci.* 2004;24(28):6402-6409.
doi: 10.1523/JNEUROSCI.1492-04.2004
90. DePaul MA, Palmer M, Lang BT, et al. Intravenous multipotent adult progenitor cell treatment decreases inflammation leading to functional recovery following spinal cord injury. *Sci Rep.* 2015;5:167195.
doi: 10.1038/srep16795
91. Nakajima H, Uchida K, Guerrero AR, et al. Transplantation of mesenchymal stem cells promotes an alternative pathway of macrophage activation and functional recovery after spinal cord injury. *J Neurotrauma.* 2012;29(8):1614-1625.
doi: 10.1089/neu.2011.2109
92. Sharma P, Maurya DK. Wharton's jelly mesenchymal stem cells: Future regenerative medicine for clinical applications in mitigation of radiation injury. *World J Stem Cells.* 2024;16(7):742-759.
doi: 10.4252/wjsc.v16.i7.742
93. You H, Wei L, Zhang J, Wang JN. Vascular endothelial growth factor enhanced the angiogenesis response of human umbilical cord-derived mesenchymal stromal cells in a rat model of radiation myelopathy. *Neurochem Res.* 2015;40(9):1892-1903.
doi: 10.1007/s11064-015-1684-0
94. Wang XZ, Xu WR, Zhu W, et al. Fluorescence labeling for human bone marrow mesenchymal stem cells with PHK26. *Chin J Lab Med.* 2006;29(9):834-837.
95. Awidi A, Al Shudifat A, El Adwan N, et al. Safety and potential efficacy of expanded mesenchymal stromal cells of bone marrow and umbilical cord origins in patients with chronic spinal cord injuries: A phase I/II study. *Cytotherapy.* 2024;26(8):825-831.
doi: 10.1016/j.jcyt.2024.03.480
96. Nakazaki M, Yokoyama T, Lankford KL, Hirota R, Kocsis JD, Honmou O. Mesenchymal stem cells and their extracellular vesicles: Therapeutic mechanisms for blood-spinal cord barrier repair following spinal cord injury. *Int J Mol Sci.* 2024;25(24):13460.
doi: 10.3390/ijms252413460
97. Fang YF, Zhang C, Han MM, et al. Engineered MSCs break endothelial-myofibroblast crosstalk in pulmonary fibrosis: Reconstructing the vascular niche. *Adv Mater.* 2025;37(13):e2414601.
doi: 10.1002/adma.202414601
98. Yusoff FM, Nakashima A, Kawano KI, et al. Implantation of Hypoxia-induced mesenchymal stem cell advances therapeutic angiogenesis. *Stem Cells Int.* 2022;2022:6795274.
doi: 10.1155/2022/6795274
99. Chen C, Zeng BW, Xue D, et al. Preliminary report of pirfenidone for the prevention of radiation pneumonitis in patients with esophageal cancer: Analysis using inverse probability of treatment weighting. *Chin J Clin Oncol.* 2021;48(15):772-776.
doi: 10.1183/13993003.01484-2024
100. Lai J, Lin PX, Huang J. Research progress of radiation-induced brain injury for nasopharyngeal carcinoma. *Cancer Res Prev Treat.* 2023;50(11):1133-1138.
101. Chen H, Wang W, Yang Y, et al. A sequential stimuli-responsive hydrogel promotes structural and functional recovery of severe spinal cord injury. *Biomaterials.* 2025;316:122995.
doi: 10.1016/j.biomaterials.2024.122995
102. Peng H, Liu Y, Xiao F, et al. Research progress of hydrogels as delivery systems and scaffolds in the treatment of secondary spinal cord injury. *Front Bioeng Biotechnol.* 2023;11:1111882.
doi: 10.3389/fbioe.2023.1111882
103. Ma YH, Shi HJ, Wei QS, et al. Developing a mechanically

- matched decellularized spinal cord scaffold for the *in situ* matrix-based neural repair of spinal cord injury. *Biomaterials*. 2021;279:121192.
doi: 10.1016/j.biomaterials.2021.121192
104. Zuo Y, Ye J, Cai W, *et al.* Controlled delivery of a neurotransmitter-agonist conjugate for functional recovery after severe spinal cord injury. *Nat Nanotechnol*. 2023;18(10):1230-1240.
doi: 10.1038/s41565-023-01416-0
105. Li W, Chen J, Zhao S, *et al.* High drug-loaded microspheres enabled by controlled in-droplet precipitation promote functional recovery after spinal cord injury. *Nat Commun*. 2022;13(1):1262.
doi: 10.1038/s41467-022-28787-7
106. Wang ZM, Bi MY, He JF, Ren BX, Liu DJ. Development of CRISPR/Cas9 system and its application in animal gene editing. *China Biotechnol*. 2020;40(10):43-50.
107. Kong WJ. *Electrospun PLGA Scaffolds Co-loaded with Neural Stem Cells and fingolimod for Repairing Spinal Cord Injury*. China: Jilin University; 2020.
108. Minhas PS, Jones JR, Latif-Hernandez A, *et al.* Restoring hippocampal glucose metabolism rescues cognition across Alzheimer's disease pathologies. *Science*. 2024;385(6711):eabm6131.
doi: 10.1126/science.abm6131
109. Li M, Fang Y. Research progresses of TGF- β /Smads signaling pathway-related therapeutic strategies for pathological scar. *J Shanghai Jiaotong Univ (Med Sci)*. 2016;36(4):594.
doi: 10.3969/j.issn.1674-8115.2016.04.027
110. Li, J. *Mechanism of TGN-020 Promoting Motorfunction Recovery After Spinal Cord Injury in Rat*. China: Jinzhou Medical University; 2019.
111. Li J, Li G, Guo WD, *et al.* Effects of TGN-020 on secondary edema and astrocyte proliferation after spinal cord injury in rats. *J Xi'an Jiaotong Univ (Med Sci)*. 2018;39(5):685-690+718.