

RESEARCH ARTICLE

Collagen bioinks redefined: Optimizing ionic strength and growth factor delivery for cartilage tissue engineering

Supplementary File

Purity of the extract

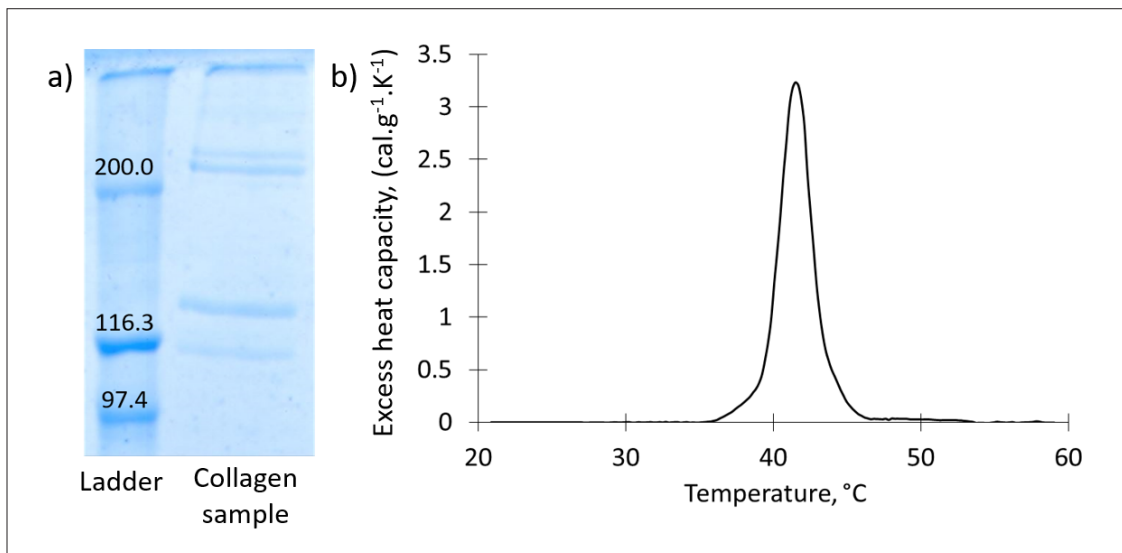


Figure S1. High quality of the extracted collagen was ascertained by (a) sodium dodecyl-sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and (b) differential scanning calorimetry (DSC).

Rheology of collagen gelation

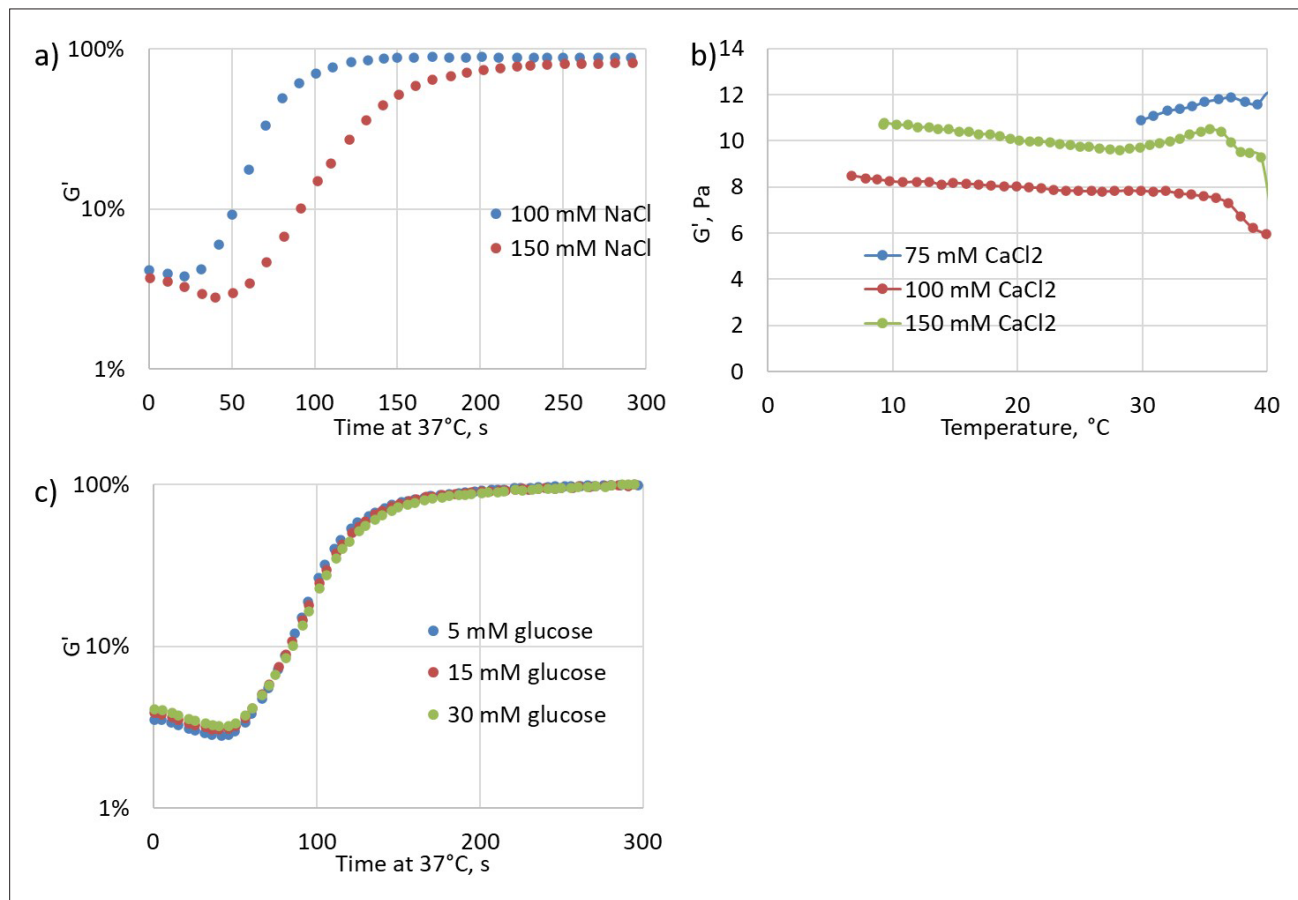


Figure S2. Results from rheological studies in oscillation mode, demonstrating the effect of (a) Cl^- ions, (b) Ca^{2+} ions, and (c) glucose in the presence of 1× phosphate-buffered saline (PBS) on storage modulus (G'), which corresponds to collagen gelation.

Proliferation of cells bioprinted in collagen inks with subphysiological ionic strength

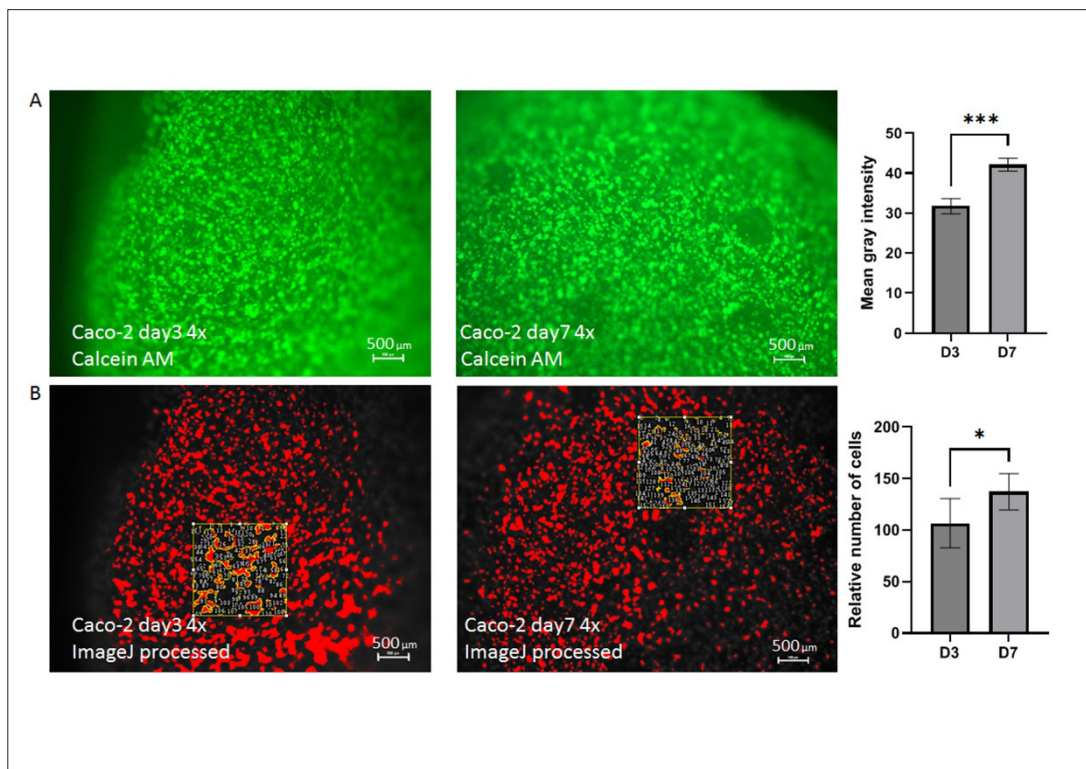


Figure S3. Assessment of proliferation of Caco-2 cells. (a) Representative images of calcein AM-stained Caco-2 cells on days 3 (left) and 7 (right) at 4x magnification. (b) Processing of the images in (a) with ImageJ for assessing the mean intensity and number of cells (particles) per ~2 mm². Images were processed in the same way (i.e., adjusting contrast and threshold values to the same parameters). Mean grey intensity and number of cells (particles) were estimated from at least two different regions of interest (~2 mm² each) from two images taken on day 3 and at least two different regions of interest from images taken on day 7. Scale bars:500 µm. Graphs represent the mean of the measurements with error bars (standard deviation). Student’s t-test values of * denotes p < 0.05, and *** annotates p < 0.0005.

Collagen printability

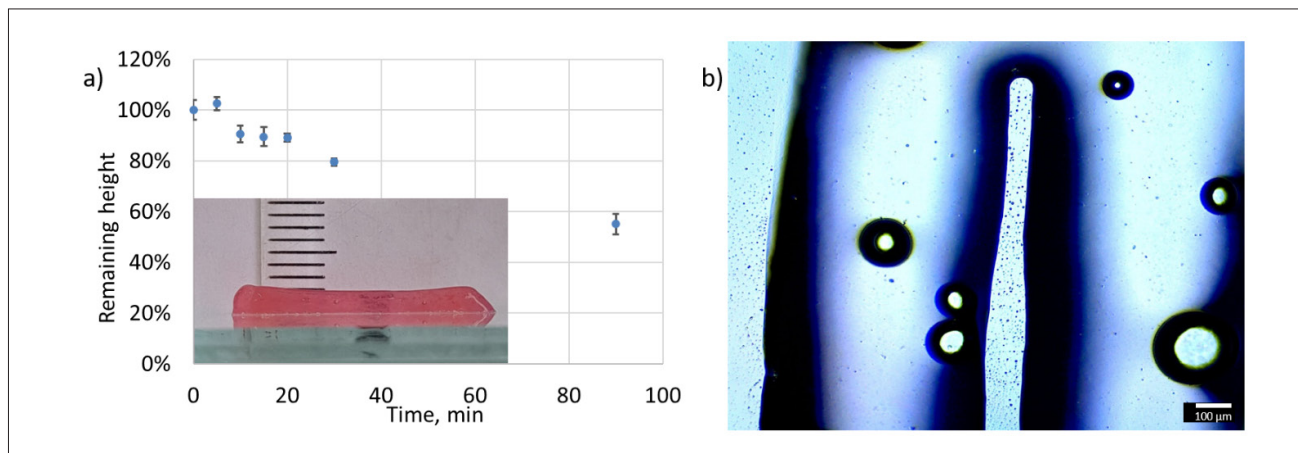


Figure S4. Collagen printability: (a) stability of a 10-layer “wall” at ambient conditions over 90 min (*n* = 5), and inset image displays a representative as-deposited 10-layer “wall”; and (b) collagen strands printed as close as 80 µm apart. Scale bar: 100 µm (b).

Comparison of KEGG pathways for glycosaminoglycan synthesis

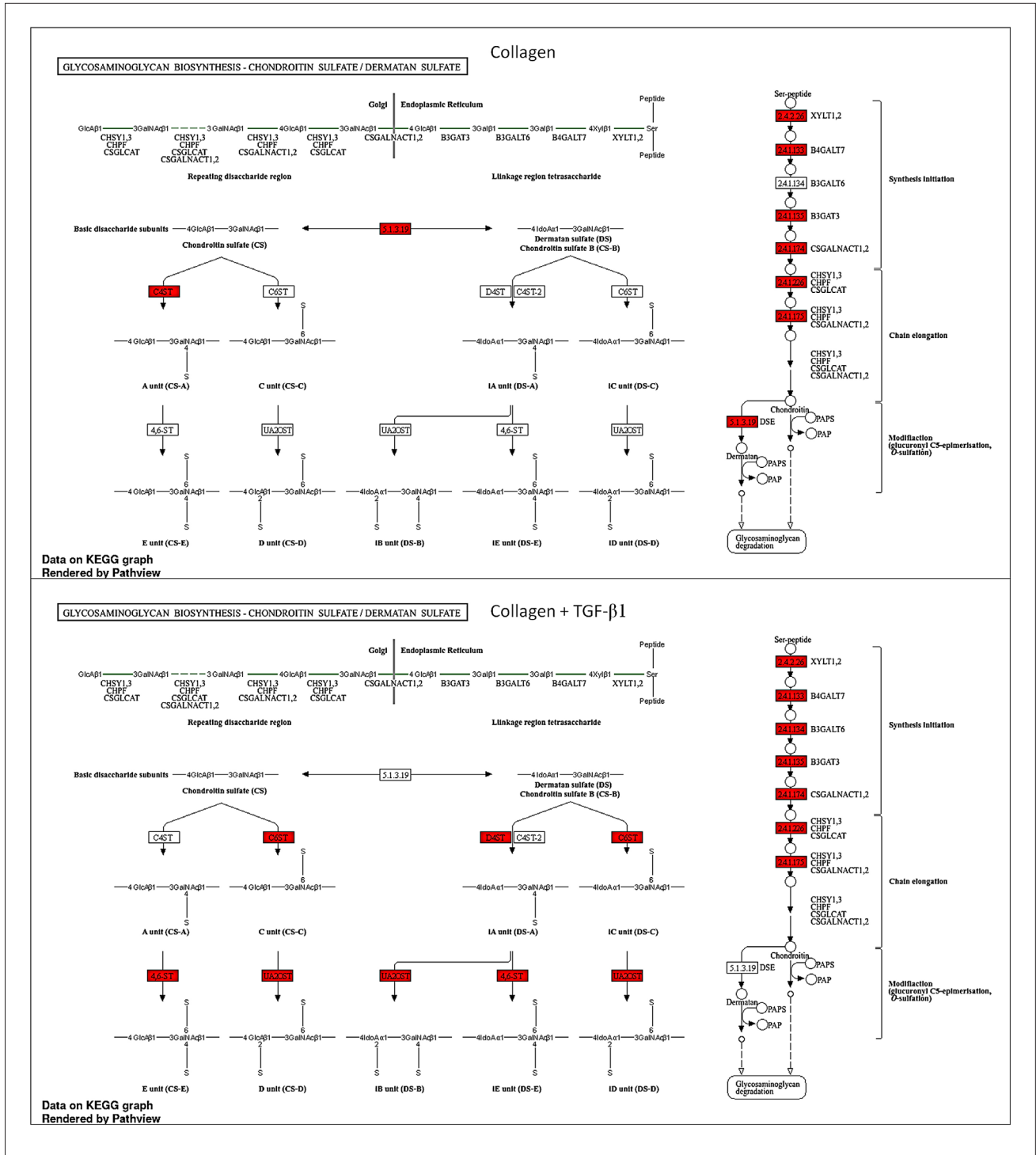


Figure S5. Comparison of upregulated genes in stromal vascular fraction (SVF) cells bioprinted in pure collagen (top) and biofunctionalized collagen (bottom) involved in cartilage extracellular matrix (ECM) biosynthesis.

Gene ontology analysis confirmation with StringDB

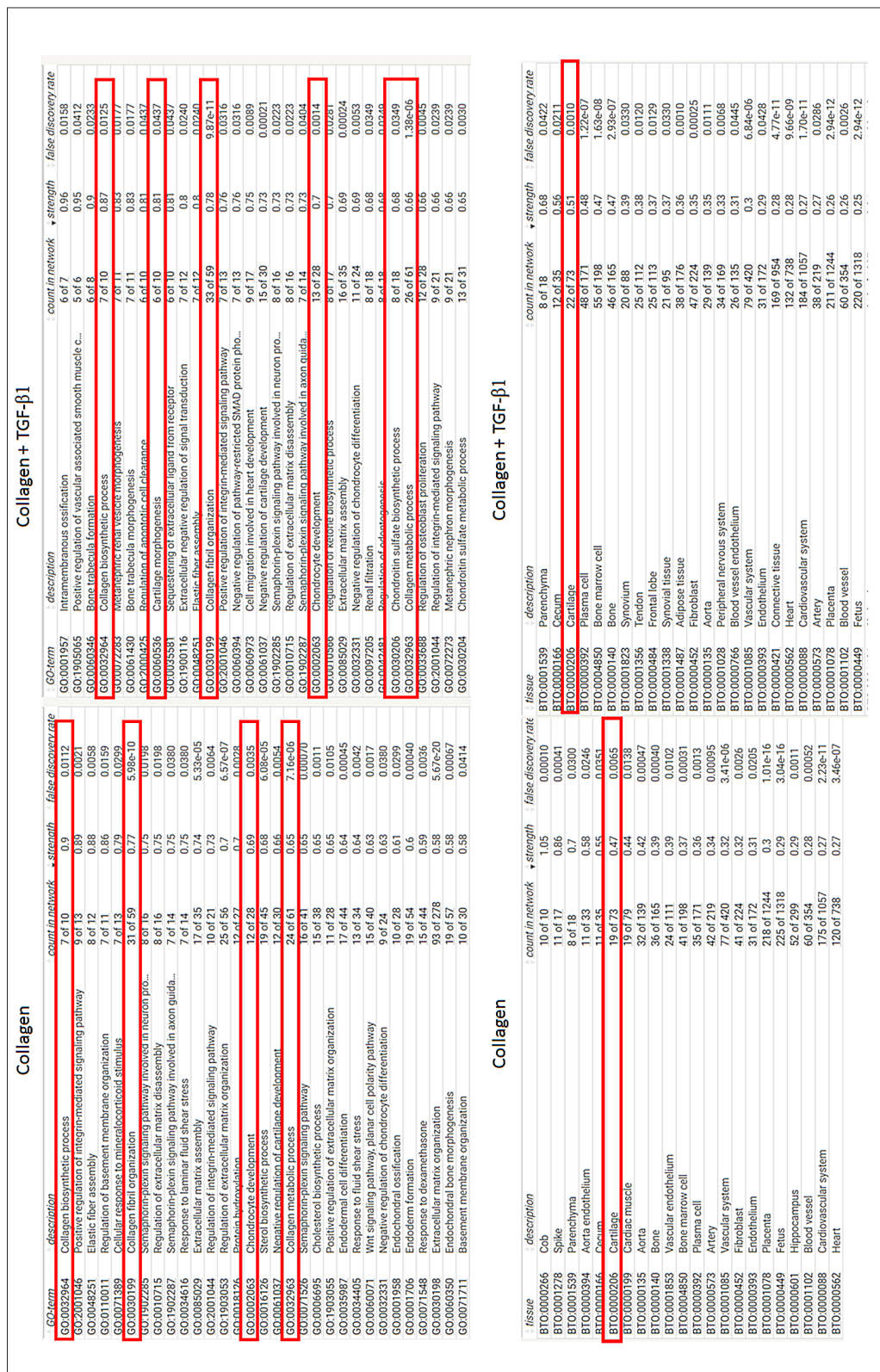


Figure S6. Gene ontology analysis with StringDB for biological processes and tissue expression. Red boxes highlight annotations of interest related to chondrogenesis.

Analysis of genes that are distinctly upregulated in 3D-bioprinted, biofunctionalized collagen

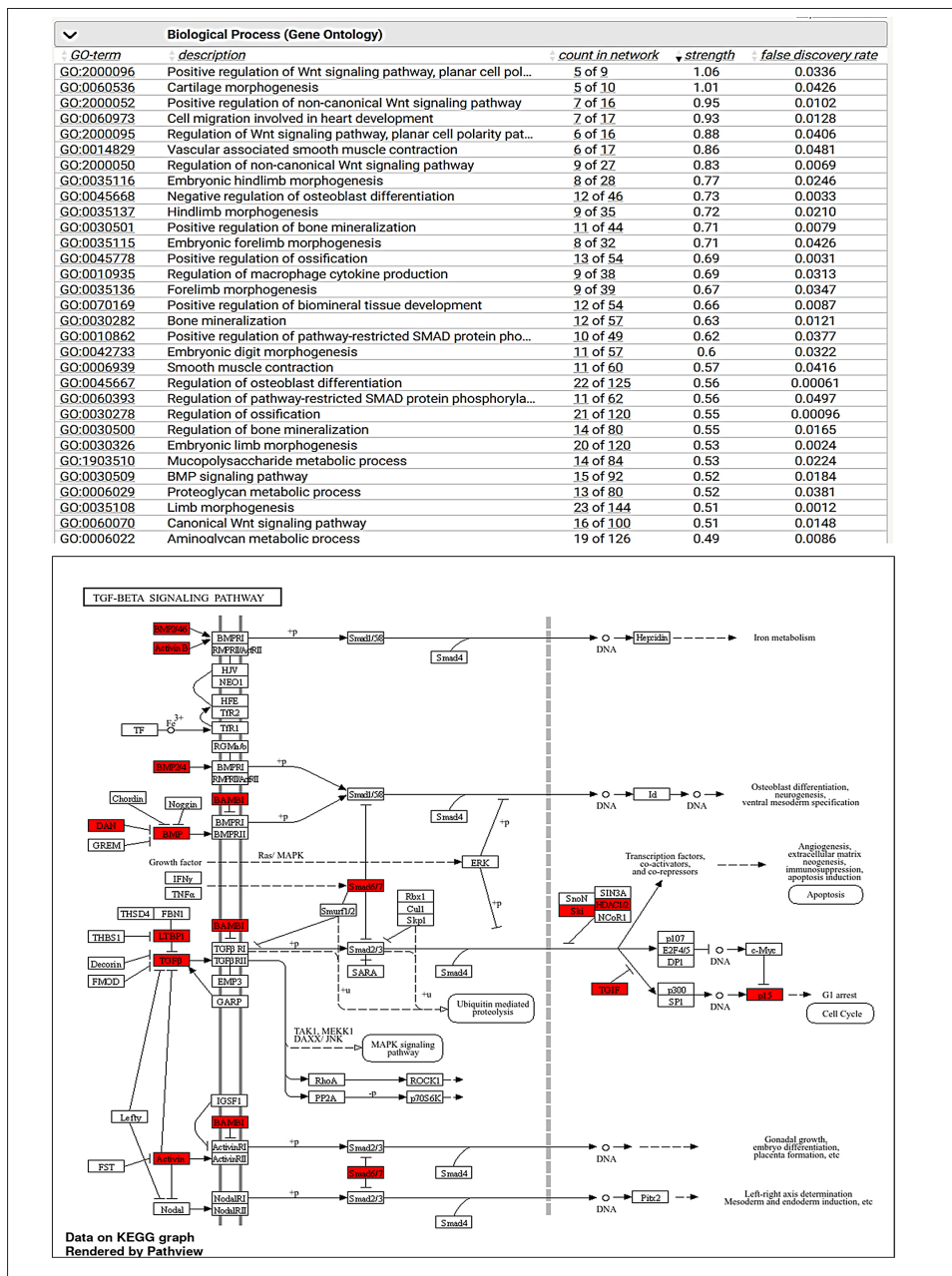


Figure S7. Gene ontology analysis with StringDB for biological processes and KEGG TGF-beta signaling pathway. Only genes that are exclusively upregulated in stromal vascular fraction (SVF) cells printed in collagen and TGF-beta are considered in this analysis.

Video S1. Time lapse microscopy of 3D bioprinted CHON-001 cells one day after bioprinting. The video reveals the biocompatibility of the hydrogel with respect

to cell viability and cell motility of chondrocytes within the bioprints.