

REVIEW ARTICLE

Smart hydrogel bioinks integrated with two-dimensional nanomaterials for cardiovascular bioprinting and atherosclerosis modeling

Jiangzhou Chu¹ , Zhi Liu² , Zhongshan He^{3,4} , and Shengbin Liu^{2*} 

¹School of Medicine and Health, College of Medicine and Health, Urban Vocational College of Sichuan, Chengdu, Sichuan, China

²Department of Critical Care Medicine, Frontiers Science Center for Disease-related Molecular Network, State Key Laboratory of Biotherapy and Cancer Center, West China Hospital, Sichuan University, Chengdu, Sichuan, China

³Department of Molecular Physiology and Biological Physics, The Robert M. Berne Cardiovascular Research Center, University of Virginia, Charlottesville, Virginia, United States of America

⁴Department of Biomedical Engineering, The Robert M. Berne Cardiovascular Research Center, University of Virginia, Charlottesville, Virginia, United States of America

Abstract

Cardiovascular diseases remain the leading cause of mortality worldwide, and atherosclerosis underlies most ischemic cardiovascular events. However, conventional *in vitro* and *in vivo* models fail to fully reproduce the complex cellular interactions, extracellular matrix remodeling, oxidative stress, and hemodynamic microenvironment of human vascular lesions. This review summarizes recent advances in stimuli-responsive hydrogel bioinks for three-dimensional bioprinting of atherosclerosis models. We discuss how disease-responsive biomaterials can dynamically regulate the cellular microenvironment and improve the physiological relevance of engineered vascular tissues. Particular emphasis is placed on the integration of two-dimensional nanomaterials. Graphene-based materials provide electrical conductivity, mechanical reinforcement, and sensing capabilities; black phosphorus offers biodegradability, photothermal responsiveness, and reactive oxygen species-sensitive behavior; and emerging Xene materials such as germanene present additional opportunities for multifunctional bioink design. We compare their respective advantages and limitations for cardiovascular bioprinting. Finally, we discuss key translational challenges, including material standardization, long-term biosafety, biological validation, manufacturing scalability, and regulatory approval. We propose that multifunctional, disease-responsive bioinks combined with advanced biofabrication technologies will accelerate the development of physiologically relevant cardiovascular disease models and precision medicine platforms.

Keywords: Cardiovascular bioprinting; Atherosclerosis modeling; Stimuli-responsive hydrogel bioinks; 2D nanomaterials; Organ-on-a-chip

*Corresponding author:

Shengbin Liu
(liushengbin88@scu.edu.cn)

Citation: Chu J, Liu Z, He Z, Liu S. Smart hydrogel bioinks integrated with two-dimensional nanomaterials for cardiovascular bioprinting and atherosclerosis modeling. *Int J Bioprint.* 2026;12(4):026230240. doi: 10.36922/IJB026230240

Received: June 6, 2026

Revised: July 4, 2026

Accepted: July 14, 2026

Published online: July 15, 2026

Copyright: © 2026 Author(s). This is an Open-Access article distributed under the terms of the Creative Commons Attribution License, permitting distribution, and reproduction in any medium, provided the original work is properly cited.

Publisher's Note: AccScience Publishing remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

1. Introduction

Atherosclerosis is the principal pathological process underlying most cardiovascular

diseases (CVDs), including coronary artery disease, myocardial infarction, ischemic stroke, and peripheral arterial disease.¹⁻³ Although traditionally viewed as a lipid-storage disorder, atherosclerosis is now recognized as a multifactorial chronic inflammatory disease involving endothelial dysfunction, oxidative stress, immune-cell recruitment, vascular smooth muscle cell (VSMC) phenotypic modulation, extracellular matrix (ECM) remodeling, and biomechanical disturbances associated with altered blood flow.⁴⁻⁷ These interconnected biological and mechanical processes create a highly heterogeneous vascular microenvironment that continuously evolves during disease initiation, progression, and plaque destabilization. Consequently, reproducing the complexity of the atherosclerotic niche remains one of the major challenges in cardiovascular research.^{8,9}

Current experimental models only partially recapitulate the pathophysiological features of human atherosclerosis. Conventional two-dimensional (2D) cell cultures lack the three-dimensional (3D) architecture, multicellular interactions, ECM organization, and mechanical stimuli that regulate vascular homeostasis and disease progression. Animal models have substantially advanced our understanding of atherosclerotic mechanisms; however, species-specific differences in lipid metabolism, immune responses, and vascular biology often limit their translational relevance. More recently, microphysiological systems and organ-on-a-chip (OOC) technologies have emerged as promising platforms for modeling vascular diseases under controlled conditions. Nevertheless, many existing *in vitro* systems remain limited in their ability to simultaneously reproduce the structural complexity, cellular heterogeneity, and dynamic pathological microenvironment characteristic of human atherosclerotic lesions. These limitations underscore the need for advanced biofabrication strategies capable of reconstructing physiologically relevant and disease-specific vascular models.¹⁰⁻¹²

Recent advances in tissue engineering and biofabrication have positioned 3D bioprinting as a powerful approach for generating biomimetic vascular tissues and disease models. By enabling the spatially controlled deposition of living cells, biomaterials, and bioactive factors, 3D bioprinting offers unprecedented opportunities to recreate multicellular architectures and microenvironmental organization that more closely resemble native tissues.¹³ Furthermore, the integration of bioprinting with OOC platforms, organoid technologies, and perfusion systems has expanded the capacity of engineered models to incorporate vascular flow, mechanical stimulation, and tissue-specific structural features.¹³⁻¹⁶ Despite these

advances, the success of bioprinted cardiovascular constructs remains fundamentally dependent on the design and functionality of the bioinks used to fabricate them.

Hydrogel-based bioinks have become the predominant class of materials used in cardiovascular bioprinting owing to their high water content, biocompatibility, tunable physicochemical properties, and structural resemblance to native ECM.¹³ Natural biomaterials such as gelatin, collagen, fibrin, alginate, hyaluronic acid, and decellularized ECM have been widely investigated because they provide favorable environments for cell viability, proliferation, migration, and differentiation.^{17,18} However, most conventional hydrogel bioinks behave as passive structural matrices and possess limited capacity to mimic the dynamic nature of pathological vascular microenvironments.¹⁷ This limitation is particularly important in atherosclerosis, where disease progression is driven by continuously changing biochemical and biomechanical signals, including oxidative stress, inflammatory cytokines, lipid accumulation, matrix remodeling, and cyclic mechanical stimulation.¹² Consequently, there is growing interest in developing smart bioinks capable of actively responding to disease-relevant stimuli and dynamically regulating cellular behavior.

Stimuli-responsive hydrogels have emerged as a promising class of smart biomaterials capable of altering their physicochemical properties in response to environmental cues such as temperature, pH, reactive oxygen species (ROS), enzymes, light, electrical stimulation, and magnetic fields.¹⁹⁻²¹ Unlike conventional static matrices, these adaptive materials enable spatiotemporal regulation of matrix mechanics, degradation, molecular delivery, and cell-matrix interactions. Such dynamic responsiveness offers unique opportunities to reconstruct pathological microenvironments more faithfully and to model disease progression in a controllable manner.²²⁻²⁴ Among the various pathological hallmarks of atherosclerosis, oxidative stress has attracted particular attention because excessive ROS production contributes to endothelial dysfunction, lipoprotein oxidation, chronic inflammation, macrophage activation, VSMC phenotypic switching, and plaque instability.²⁵⁻²⁶ Therefore, ROS-responsive bioinks represent an especially attractive strategy for developing disease-adaptive vascular models that more closely mimic *in vivo* pathological conditions.

In parallel with the evolution of smart hydrogels, 2D nanomaterials have emerged as powerful functional components for next-generation bioinks.²⁷ Owing to their unique structural, electrical, mechanical, optical, and surface properties, these materials can significantly

enhance biofabrication performance while introducing disease-relevant biological functionalities. Rather than serving solely as passive reinforcing agents, 2D nanomaterials increasingly function as active regulators of cellular behavior and microenvironmental cues.²⁸ Graphene-family nanomaterials have been extensively explored for improving mechanical stability, electrical conductivity, printability, and biosensing capabilities in cardiovascular tissue engineering.²⁹⁻³¹ Black phosphorus (BP), in contrast, offers distinctive advantages including intrinsic biodegradability, photothermal responsiveness, and ROS-sensitive degradation behavior.³²⁻³³ Importantly, accumulating evidence suggests that BP-containing biomaterials can modulate oxidative stress, regulate inflammatory responses, and promote angiogenesis, characteristics that closely align with critical pathological processes involved in atherosclerosis.^{27,32} Beyond graphene and BP, other emerging Xene-family materials such as germanene are beginning to attract attention owing to their unique electronic structures, surface reactivity, and multifunctional responsiveness.^{27,34,35,36} Collectively, these advances substantially expand the design space of smart bioinks for cardiovascular biofabrication and provide new opportunities for engineering disease-responsive vascular microenvironments.^{27,35}

Despite significant progress in cardiovascular bioprinting, hydrogel engineering, OOC technologies, and nanomaterial-based biomaterials, important knowledge gaps remain. Existing reviews have largely examined these research areas independently, focusing on bioprinting platforms, hydrogel bioinks, microphysiological systems, or specific classes of nanomaterials.¹¹⁻¹³ Comparatively little attention has been devoted to understanding how bioink design can be tailored to the unique pathophysiological requirements of atherosclerosis. In particular, the integration of stimuli-responsive hydrogels and functional 2D nanomaterials has not been systematically evaluated as a unified strategy for addressing key disease-specific challenges, including oxidative stress, chronic inflammation, vascular remodeling, and dynamic microenvironmental regulation.^{26,37} Moreover, the emerging convergence of disease-responsive biomaterials, advanced biofabrication technologies, and vascular disease modeling has yet to be critically assessed from the perspective of constructing physiologically relevant and pathologically adaptive atherosclerosis platforms.^{11,12}

In this review, we propose an atherosclerosis-oriented framework for the design of smart bioinks integrating stimuli-responsive hydrogels and functional 2D nanomaterials. We first discuss the pathophysiological requirements for reconstructing atherosclerotic

microenvironments and the corresponding design criteria for advanced bioinks. We then summarize recent advances in stimuli-responsive hydrogel systems and evaluate the functional contributions of graphene-family nanomaterials, BP, and emerging germanene-based platforms. Subsequently, we examine their applications in vascular bioprinting and atherosclerosis-on-a-chip systems and discuss how these technologies may facilitate more physiologically relevant disease reconstruction. Finally, we highlight current challenges, translational barriers, and future opportunities toward the development of dynamic, disease-responsive, and patient-specific cardiovascular models for disease investigation, drug screening, and precision medicine.

2. Pathophysiological requirements for atherosclerosis modeling

The development of physiologically relevant atherosclerosis models requires more than simply reproducing individual cellular phenotypes or isolated biochemical pathways. Atherosclerotic plaques evolve through dynamic interactions among vascular cells, immune cells, ECM components, circulating lipoproteins, and biomechanical forces. Consequently, successful disease reconstruction depends on the ability to recapitulate multiple pathological features simultaneously within a controllable microenvironment.^{2,7,8} Traditional *in vitro* models typically reproduce only one or several aspects of disease progression, such as endothelial inflammation or macrophage lipid uptake, whereas native plaques represent highly heterogeneous and continuously evolving ecosystems.⁶ Therefore, understanding the key pathophysiological requirements of atherosclerosis is essential for guiding the rational design of next-generation bioinks and bioprinted vascular disease models.^{11,12}

2.1. Endothelial dysfunction as the primary disease trigger

The vascular endothelium serves as a dynamic interface between circulating blood and the arterial wall. Under physiological conditions, endothelial cells maintain vascular homeostasis by regulating vascular permeability, thrombosis, inflammation, and vascular tone through the production of nitric oxide (NO), prostacyclin, and other vasoprotective mediators.⁹

Atherosclerosis is initiated when endothelial homeostasis becomes disrupted by cardiovascular risk factors, including hyperlipidemia, hypertension, diabetes mellitus, smoking, and disturbed blood flow. Dysfunctional endothelial cells exhibit reduced NO bioavailability, increased oxidative stress, elevated permeability to circulating lipoproteins,

and enhanced expression of adhesion molecules such as vascular cell adhesion molecule-1 (VCAM-1), intercellular adhesion molecule-1 (ICAM-1), and E-selectin.^{7,9} These alterations promote leukocyte recruitment and lipoprotein retention within the arterial intima, thereby establishing a pro-inflammatory microenvironment that drives plaque initiation and progression.⁷

Most conventional endothelial culture systems fail to reproduce the 3D vascular architecture, physiological shear stress, and cell–matrix interactions that regulate endothelial behavior *in vivo*. Consequently, endothelial responses observed in static monolayer cultures frequently differ from those occurring in native arteries under physiological or pathological flow conditions.^{11,12}

For bioprinted atherosclerosis models, the reconstruction of a functional endothelial barrier therefore represents a fundamental design requirement. Bioinks should support endothelial adhesion, junction formation, mechanosensing, and long-term barrier integrity while permitting dynamic responses to inflammatory and hemodynamic stimuli. Furthermore, endothelialized vascular constructs should enable real-time assessment of endothelial permeability, inflammatory activation, leukocyte adhesion, and barrier dysfunction as key indicators of disease initiation and progression.^{11,12}

2.2. Vascular smooth muscle cell plasticity and arterial remodeling

Vascular smooth muscle cells are the predominant cellular component of the arterial media and are responsible for maintaining vascular contractility and structural integrity. In healthy vessels, VSMCs exhibit a contractile phenotype characterized by expression of α -smooth muscle actin (α -SMA), smooth muscle myosin heavy chain (SM-MHC), and calponin.^{38,39}

During atherosclerosis progression, VSMCs undergo extensive phenotypic switching toward synthetic, inflammatory, osteogenic, and macrophage-like states. This phenotypic plasticity contributes to ECM production, plaque growth, vascular calcification, and fibrous cap formation.^{40,41} Traditional 2D culture systems often fail to maintain physiological VSMC phenotypes because cell behavior is strongly influenced by matrix stiffness, dimensionality, and cell–cell interactions. Consequently, VSMCs cultured on rigid substrates frequently exhibit artificial activation profiles that do not accurately reflect disease progression.⁴² Next-generation bioprinted vascular models should therefore provide tunable mechanical microenvironments capable of regulating VSMC phenotype in a controlled manner. Bioinks should enable spatial localization of VSMCs within arterial wall-like

structures while supporting phenotypic transitions that occur during plaque development. Such capabilities are essential for reproducing arterial remodeling and fibrous cap formation.^{38,42}

2.3. Immune cell recruitment and macrophage polarization

Atherosclerosis is increasingly recognized as a chronic inflammatory vascular disease driven by both innate and adaptive immune responses.^{43,44} Among immune cell populations, macrophages play a central role in plaque initiation, progression, and destabilization.^{44,45} Following endothelial activation, circulating monocytes infiltrate the arterial wall and differentiate into macrophages.^{43,45} These macrophages internalize oxidized low-density lipoprotein (oxLDL), forming lipid-laden foam cells that constitute one of the earliest hallmarks of plaque formation.⁴⁴ As disease progresses, macrophages secrete inflammatory cytokines, matrix-degrading enzymes, and ROS that contribute to tissue remodeling and plaque instability.^{44,46}

Importantly, macrophages exhibit remarkable phenotypic heterogeneity.^{45,47} Pro-inflammatory macrophages promote lesion progression through secretion of tumor necrosis factor alpha (TNF- α), interleukin (IL)-1 β , and IL-6, whereas reparative macrophages participate in inflammation resolution and tissue repair.^{45,47} The balance between these phenotypes significantly influences plaque stability and therapeutic outcomes.^{45,47} Most existing vascular models either omit immune cells entirely or incorporate them only transiently. As a result, critical inflammatory processes remain poorly represented. Future bioprinted atherosclerosis models must therefore support immune-cell recruitment, macrophage infiltration, foam-cell formation, and dynamic polarization.^{15,48} This requirement creates a strong demand for bioinks capable of regulating immune-cell behavior through biochemical and biophysical cues.^{15,48}

2.4. Oxidative stress as a disease-defining microenvironment

Oxidative stress has emerged as one of the most important regulators of atherosclerosis progression. Excessive production of ROS contributes to endothelial dysfunction, low-density lipoprotein (LDL) oxidation, inflammatory activation, VSMC migration, and ECM degradation.^{25,49} Unlike many pathological signals, oxidative stress exhibits significant spatial and temporal heterogeneity within plaques. Regions enriched with activated macrophages, inflammatory cell clusters, and necrotic-core-associated microenvironments frequently show enhanced oxidative and inflammatory signatures, whereas neighboring plaque regions may remain biologically distinct.^{45,50–52} Such

heterogeneity is difficult to reproduce using conventional *in vitro* models.

The inability to model dynamic redox microenvironments represents a major limitation of current atherosclerosis platforms. Consequently, ROS-responsive biomaterials have attracted increasing attention because they offer opportunities to engineer disease-relevant feedback mechanisms directly into bioinks.²⁶ For cardiovascular bioprinting, ideal bioinks should not only tolerate oxidative environments but also dynamically respond to ROS accumulation through matrix remodeling, cargo release, signal amplification, or biosensing functions.^{26,53} These requirements provide a strong rationale for incorporating disease-responsive nanomaterials such as BP into hydrogel systems.^{54,55}

2.5. Extracellular matrix remodeling and plaque stability

The ECM provides structural support and biochemical cues that regulate vascular-cell behavior. During atherosclerosis progression, extensive remodeling occurs within the ECM, including collagen accumulation, elastin fragmentation, proteoglycan deposition, and matrix degradation mediated by matrix metalloproteinases (MMPs).^{5,56}

The composition and organization of the ECM strongly influence plaque stability. Stable plaques generally contain thick collagen-rich fibrous caps, whereas vulnerable plaques are characterized by increased proteolytic activity, inflammatory infiltration, and thinning of the fibrous cap.^{38,57} Plaque rupture, which underlies most acute cardiovascular events, is closely associated with ECM destabilization and excessive matrix degradation driven by inflammatory cells and MMP activity.^{58,59}

Most hydrogel-based models provide static extracellular matrices that cannot reproduce progressive matrix remodeling. Therefore, future bioinks should support controlled degradation, matrix deposition, and mechanical evolution over time. Dynamic ECM remodeling capabilities will be essential for modeling plaque progression and vulnerability.^{5,56}

2.6. Hemodynamic regulation and mechanobiological signaling

One of the defining characteristics of atherosclerosis is its non-random localization within the vasculature. Plaques preferentially develop at arterial bifurcations, branch points, and curved regions exposed to disturbed blood flow and oscillatory shear stress.^{9,60-62} Hemodynamic forces regulate endothelial function through mechanotransduction pathways involving Krüppel-like

factor 2, nuclear factor kappa B, yes-associated protein/transcriptional coactivator with PDZ-binding motif, platelet endothelial cell adhesion molecule-1, and integrin signaling.^{60,63} Laminar shear stress generally promotes anti-inflammatory and atheroprotective phenotypes, whereas disturbed flow induces endothelial activation, oxidative stress, and inflammatory signaling.^{9,60,63} Conventional static cultures fail to reproduce these critical biomechanical influences. Consequently, integrating perfusion systems and microfluidic technologies with bioprinted constructs has become increasingly important for CVD modeling.^{64,65} Future bioprinted atherosclerosis platforms should therefore incorporate controllable flow conditions, physiologically relevant shear stress profiles, and patient-specific vascular geometries. Such systems would allow investigation of the complex interplay between mechanical forces and biochemical disease cues.^{64,65}

2.7. Design criteria for next-generation bioprinted atherosclerosis models

To summarize the relationship between atherosclerotic pathology and bioink engineering requirements, [Figure 1](#) presents a disease-oriented framework linking key pathological features with the functional criteria required for next-generation smart bioink design. Collectively, current understanding of atherosclerosis pathogenesis reveals several essential requirements for physiologically relevant disease reconstruction.^{9,44,66} An ideal bioprinted model should integrate: (i) a functional endothelial barrier; (ii) spatially organized VSMCs with phenotype plasticity; (iii) dynamic immune-cell recruitment and macrophage polarization; (iv) heterogeneous oxidative stress microenvironments; (v) tunable ECM remodeling; and (vi) physiologically relevant hemodynamic stimulation.^{9,46,56,66,67} Meeting these requirements exceeds the capabilities of conventional biomaterials and highlights the need for multifunctional bioinks capable of actively interacting with disease-associated signals.^{48,68} Stimuli-responsive hydrogels offer dynamic control over matrix properties and cellular behavior,^{69,70} whereas 2D nanomaterials introduce conductivity, photothermal responsiveness, biosensing capability, and redox regulation.^{71,72} The convergence of these technologies provides a promising foundation for constructing next-generation CVD models with unprecedented physiological relevance.^{48,68,73} The major pathological features of atherosclerosis and their corresponding bioink design requirements are summarized in [Table 1](#). The following section therefore focuses on stimuli-responsive hydrogel bioinks and examines how smart biomaterials can be rationally designed to satisfy the biological and engineering requirements outlined above.

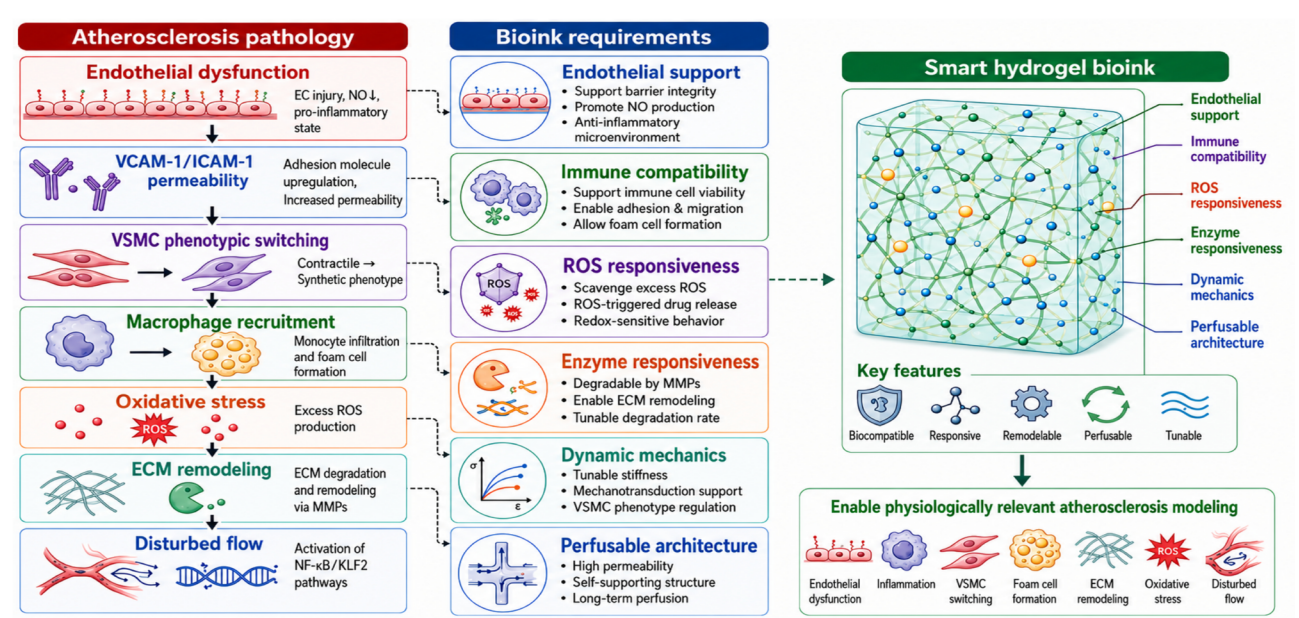


Figure 1. Pathophysiological requirements and bioink design criteria for atherosclerosis modeling. The progression of atherosclerosis involves endothelial dysfunction, oxidative stress, macrophage recruitment, foam-cell formation, VSMC phenotypic switching, ECM remodeling, and disturbed flow. These pathological features define the functional requirements of next-generation bioinks, including dynamic matrix remodeling, ROS responsiveness, tunable mechanics, immune-cell compatibility, and perfusion stability. Created in BioRender. Chu, R. (2026). <https://BioRender.com/ygc93cd>, with assistance from ChatGPT (OpenAI, GPT-5.5 Thinking) for preliminary figure drafting, visual refinement, and layout optimization. Abbreviations: EC: Endothelial cell; ECM: Extracellular matrix; ICAM-1: Intercellular adhesion molecule 1; KLF2: Krüppel-like factor 2; MMPs: Matrix metalloproteinases; NF-κB: Nuclear factor kappa B; NO: Nitric oxide; ROS: Reactive oxygen species; VCAM-1: Vascular cell adhesion molecule 1; VSMC: Vascular smooth muscle cell.

Table 1. Disease-specific pathological features of atherosclerosis and corresponding bioink design requirements

Pathological feature	Representative readouts	Bioink design requirement	Representative implementation
Endothelial dysfunction ^{3,9,65}	TEER, permeability, eNOS, NO	Barrier-supportive matrix	Endothelialized lumen
Oxidative stress ^{25,26,68,74}	ROS, oxidative biomarkers	ROS-responsive matrix	Thioketal/BP systems
Inflammation ^{75,76}	VCAM-1, ICAM-1, cytokines	Immune-compatible bioink	Cytokine-responsive systems
ECM remodeling ^{56,59,77,78}	MMP activity, collagen organization	Enzyme-responsive degradation	MMP-cleavable hydrogels
Disturbed flow ^{7,60,79}	KLF2, NF-κB	Perfusable vascular architecture	Microfluidics/OOC
VSMC phenotypic switching ^{40,80}	α-SMA, SM-MHC	Dynamic mechanical adaptability	Mechanoadaptive bioinks

Abbreviations: α-SMA: Alpha-smooth muscle actin; BP: Black phosphorus; ECM: Extracellular matrix; eNOS: Endothelial nitric oxide synthase; ICAM-1: Intercellular adhesion molecule 1; KLF2: Krüppel-like factor 2; MMP: Matrix metalloproteinase; NF-κB: Nuclear factor kappa B; NO: Nitric oxide; OOC: Organ-on-a-chip; ROS: Reactive oxygen species; SM-MHC: Smooth muscle myosin heavy chain; TEER: Transendothelial electrical resistance; VCAM-1: Vascular cell adhesion molecule 1; VSMC: Vascular smooth muscle cell.

3. Stimuli-responsive hydrogel bioinks for atherosclerosis modeling and cardiovascular applications

3.1. From passive scaffolds to disease-responsive bioinks

Hydrogels have become the predominant biomaterial platform for cardiovascular bioprinting owing to their high water content, tissue-like mechanical properties, and ability to support cellular viability and function. Conventional hydrogel bioinks primarily serve as passive structural matrices that provide physical support for encapsulated cells during and after the printing process. While these materials have enabled substantial advances in vascular tissue engineering, they are often unable to reproduce the dynamic and evolving microenvironments characteristic of cardiovascular tissues and vascular diseases.^{81,82} In native vessels, cells continuously respond to biochemical signals, mechanical forces, oxidative stress, inflammatory mediators, and metabolic alterations, while the ECM undergoes constant degradation and remodeling. Consequently, static biomaterials are insufficient to fully capture the adaptive nature of vascular pathophysiology.⁸³

This limitation is particularly relevant for atherosclerosis, a multifactorial disease involving chronic inflammation, oxidative stress, endothelial dysfunction, ECM remodeling, and progressive plaque development. Successful disease modeling therefore requires bioinks that can actively interact with and respond to pathological cues rather than merely providing structural support. Stimuli-responsive hydrogels offer a promising solution by dynamically altering their physicochemical properties in response to environmental triggers such as ROS, enzymes, light, electrical signals, mechanical stress, temperature, or pH. Through such responsiveness, bioinks can regulate matrix stiffness, degradation behavior, cellular interactions, and therapeutic delivery in a spatiotemporally controlled manner.^{26,70,84}

Rather than functioning solely as passive scaffolds, smart bioinks actively participate in disease-relevant microenvironmental regulation. This transition from structural support to dynamic pathological modulation represents a key step toward next-generation vascular disease models and forms an important foundation for emerging four-dimensional bioprinting strategies.^{21,82}

3.2. Atherosclerosis-specific design requirements for smart bioinks

Unlike many tissue-engineering applications that primarily focus on tissue regeneration, atherosclerosis

modeling requires bioinks capable of reproducing complex pathological microenvironments. Therefore, the design of smart bioinks should be guided not only by printability and biocompatibility but also by disease-specific biological requirements.⁸⁵

Oxidative stress is a hallmark of atherosclerosis and contributes to endothelial dysfunction, LDL oxidation, VSMC phenotypic switching, macrophage activation, and plaque progression. Bioinks capable of sensing and responding to ROS are therefore highly desirable for reproducing disease-associated oxidative microenvironments.

Chronic vascular inflammation drives all stages of atherosclerotic development. The recruitment of immune cells and sustained production of pro-inflammatory cytokines create complex biochemical gradients within plaques. Smart bioinks should therefore support the establishment of inflammatory microenvironments and enable dynamic regulation of immune-related signaling pathways.

Atherosclerotic lesions undergo continuous ECM remodeling mediated by MMPs, elastases, and other proteolytic enzymes. These processes contribute to fibrous cap formation, plaque destabilization, and vascular structural changes. Bioinks capable of cell-mediated degradation and remodeling are essential for reproducing disease progression.⁸⁶

Blood vessels are continuously exposed to pulsatile pressure, cyclic strain, and shear stress. Moreover, vascular stiffening and altered biomechanics are characteristic features of atherosclerosis. Therefore, bioinks should possess adaptive mechanical properties that enable dynamic responses to physiological and pathological mechanical stimuli.⁸¹

Collectively, these requirements highlight the need for disease-responsive bioinks capable of recapitulating the multifactorial nature of atherosclerosis rather than simply supporting vascular cell growth.⁶⁸

3.3. Reactive oxygen species-responsive bioinks

Among the various stimuli investigated for cardiovascular applications, ROS have attracted particular attention because oxidative stress is a central pathological feature of atherosclerosis, myocardial infarction, ischemia-reperfusion injury, and vascular aging.⁸⁷⁻⁸⁹ Excessive ROS production contributes to endothelial dysfunction, LDL oxidation, inflammatory activation, VSMC phenotypic modulation, and plaque instability, making oxidative stress one of the most representative hallmarks of atherosclerotic disease progression.⁹⁰⁻⁹²

Compared with conventional hydrogels, ROS-responsive systems offer several advantages for atherosclerosis modeling. First, they enable disease-specific matrix remodeling that directly reflects local oxidative conditions. Second, they facilitate controlled release of therapeutic molecules in response to disease progression. Third, they create feedback-regulated microenvironments capable of dynamically adapting to pathological severity.^{78,90} In cardiovascular tissue engineering, ROS-responsive and ROS-scavenging hydrogels have demonstrated beneficial effects in modulating oxidative microenvironments, reducing inflammatory responses, and supporting tissue repair following myocardial infarction and ischemia-reperfusion injury.^{78,93} These findings highlight the potential of ROS-responsive biomaterials for constructing disease-relevant vascular models.

Importantly, ROS-responsive bioinks are particularly attractive for atherosclerosis research because the triggering stimulus originates directly from disease progression itself. Such systems may enable localized simulation of inflamed plaque regions while preserving relatively healthy vascular compartments within the same construct. Although direct ROS-responsive bioink platforms specifically designed for atherosclerosis remain relatively limited, this area is emerging as a promising direction for next-generation vascular disease modeling.^{68,94}

3.4. Enzyme-responsive bioinks for dynamic matrix remodeling

Extracellular matrix remodeling is a defining characteristic of CVD progression. Dysregulated interactions between vascular cells and the ECM contribute to alterations in vascular structure, mechanical properties, and inflammatory signaling, thereby driving disease development and progression. MMPs, elastases, and other proteolytic enzymes play critical roles in ECM degradation and reorganization during vascular remodeling, atherosclerotic plaque progression, plaque destabilization, aneurysm formation, and post-myocardial infarction repair processes.^{95,96}

To reproduce these dynamic pathological processes, enzyme-responsive hydrogels incorporate protease-cleavable peptide sequences that can be selectively degraded by disease-associated enzymes. Such systems enable localized matrix degradation, controlled release of bioactive factors, and adaptive structural evolution in response to cellular activity.^{97,98}

Compared with conventional hydrolytically degradable hydrogels, enzyme-responsive materials provide greater biological specificity because degradation occurs predominantly in regions exhibiting active remodeling

and elevated protease expression. This feature allows cell-mediated matrix adaptation and closely mimics native tissue dynamics. Consequently, enzyme-responsive hydrogels have emerged as promising platforms for modeling fibrous cap remodeling, plaque destabilization, angiogenesis, vascular regeneration, and post-infarction tissue repair.^{97,99}

3.5. Electrically responsive bioinks

Electrical signaling plays an essential role in cardiovascular physiology. Although electrical responsiveness has been extensively investigated in cardiac tissue engineering, increasing evidence suggests that electrical cues can also influence endothelial behavior, vascular maturation, NO signaling, and mechanotransduction pathways relevant to vascular function.¹⁰⁰

Traditional hydrogel bioinks generally exhibit poor electrical conductivity, limiting electrophysiological communication and bioelectronic integration. Electrically responsive hydrogels address this limitation through incorporation of conductive components such as graphene-based materials, conductive polymers, carbon nanomaterials, metallic nanostructures, and other conductive additives.^{28,101,102} These materials improve electrical signal transmission and facilitate interactions between engineered tissues and external bioelectronic systems. In addition to supporting cellular communication, conductive bioinks may enable integration with biosensors, soft bioelectronics, and real-time monitoring platforms. Recent advances in conductive hydrogel electrodes and bioelectronic interfaces further highlight their potential for dynamic disease monitoring and functional assessment.¹⁰³

However, compared with cardiac applications, direct evidence supporting electrically responsive bioinks for atherosclerotic vascular disease modeling remains relatively limited. Nevertheless, emerging studies indicate that electrical stimulation may influence vascular formation, endothelial function, and mechanobiological responses, suggesting a potentially valuable but still underexplored direction for vascular bioengineering.^{104,105}

3.6. Photoresponsive bioinks for spatiotemporal control

Photoresponsive hydrogels provide unique advantages because light can be precisely controlled in terms of wavelength, intensity, duration, and spatial localization. As a result, photoresponsive systems enable highly controllable modulation of biomaterial properties and biological functions after construct fabrication.^{106,107} Current photoresponsive bioinks are commonly based on photocrosslinkable polymers, photoswitchable molecules,

photothermal agents, and light-triggered delivery systems. Through external optical stimulation, these materials can regulate matrix stiffness, degradation behavior, cellular interactions, and bioactive factor release with high spatial and temporal precision.^{106,107}

For vascular tissue engineering and disease modeling, photoresponsive bioinks may facilitate controlled vascular network formation, regulation of angiogenic signaling, and dynamic remodeling of engineered microenvironments.¹⁰⁸ Furthermore, incorporation of photothermal nanomaterials can enable remote regulation of cellular behavior and tissue remodeling through near-infrared (NIR) irradiation. Although most current studies remain focused on regenerative applications, photoresponsive systems offer substantial potential for constructing programmable vascular disease models with externally controlled functionality. Emerging 2D nanomaterials may further expand opportunities for photoresponsive biofabrication and dynamic vascular microenvironment regulation.^{36,109,110}

3.7. Toward multifactorial atherosclerosis-mimetic bioinks

While individual responsive mechanisms provide important advantages, atherosclerosis is fundamentally a multifactorial disease involving oxidative stress, chronic inflammation, ECM remodeling, endothelial dysfunction, biomechanical abnormalities, and disturbed hemodynamics.^{44,111,112} Consequently, single-stimulus bioinks are unlikely to fully reproduce the complexity of disease progression.

Future bioink design will therefore require integration of multiple responsive functions within a single platform. Multifunctional systems capable of simultaneously sensing oxidative stress, responding to enzymatic remodeling, adapting to mechanical alterations, and regulating therapeutic delivery may provide more physiologically relevant disease models.^{53,55,68,69} Such disease-responsive bioinks move beyond traditional biomaterials toward active microenvironmental regulators capable of sensing, interpreting, and responding to pathological signals. In this context, advanced nanomaterial-enabled hydrogels have attracted increasing interest because they offer opportunities to combine conductivity, biosensing, photothermal responsiveness, ROS regulation, and mechanical reinforcement within a unified platform.¹⁰⁸⁻¹¹¹ More broadly, recent advances in smart nanomaterials have demonstrated a clear shift toward multifunctional platforms that integrate diagnostic, therapeutic, and regenerative capabilities, providing an important

conceptual foundation for the development of next-generation disease-responsive bioinks.¹¹³⁻¹¹⁶

The development of multifunctional, disease-responsive bioinks is therefore expected to become a central direction in next-generation atherosclerosis modeling and vascular tissue engineering. The major classes of stimuli-responsive bioinks and their potential roles in cardiovascular tissue engineering and disease modeling are summarized in Figure 2.

3.8. Transition toward nanomaterial-enabled smart bioinks

The design principles discussed above demonstrate that effective atherosclerosis models require bioinks capable of responding to oxidative stress, inflammatory signaling, matrix remodeling, and dynamic vascular microenvironments. Achieving these functions within a single material system remains a significant challenge. A comparison of the major classes of stimuli-responsive bioinks and their cardiovascular relevance is provided in Table 2.

Two-dimensional nanomaterials have emerged as particularly promising candidates for addressing this challenge because they can simultaneously provide mechanical reinforcement, electrical conductivity, photothermal responsiveness, ROS regulation, and biosensing functionality. As summarized in Figure 3, hydrogel matrices provide the structural and biological foundation of bioinks, whereas 2D nanomaterials function primarily as physicochemical and bioactive enhancers that improve mechanical stability, conductivity, cellular regulation, and printability. Consequently, the following sections examine how graphene-family nanomaterials, BP, and emerging Xene-based materials can be incorporated into smart hydrogel bioinks to construct next-generation platforms for vascular disease modeling and cardiovascular bioprinting.

4. Graphene-based smart bioinks: Functions, evidence, and design boundaries

4.1. Graphene derivatives and formulation logic

Graphene-family nanomaterials are among the most mature and extensively investigated 2D materials in bioprinting and tissue engineering. Since the discovery of graphene, numerous derivatives—including graphene oxide (GO), reduced GO (rGO), graphene nanoplatelets, and graphene quantum dots—have been developed to improve processability, dispersibility, conductivity, and biocompatibility.²⁹

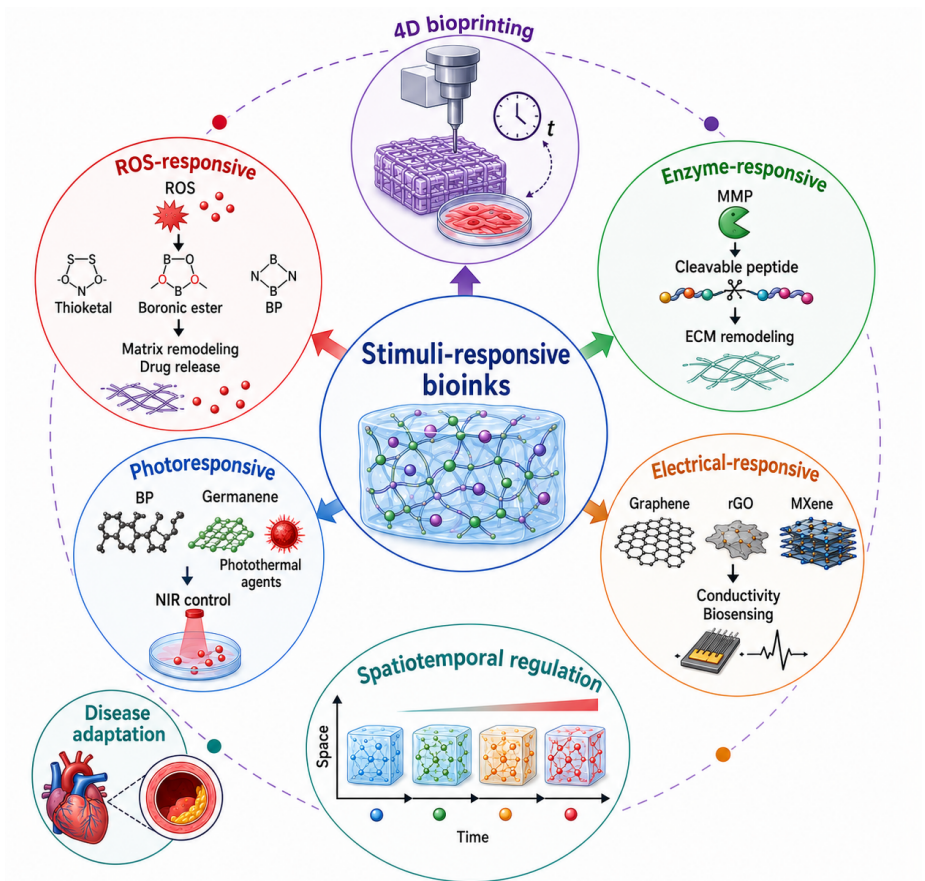


Figure 2. Stimuli-responsive hydrogel bioinks for cardiovascular applications. Smart hydrogel bioinks respond to disease-associated stimuli including ROS, enzymes, electrical signals, light, and mechanical stress. These responses regulate hydrogel degradation, stiffness, drug release, cell behavior, and tissue maturation, enabling dynamic reconstruction of cardiovascular microenvironments. Created in BioRender. Chu, R. (2026). <https://BioRender.com/5813oka>, with assistance from ChatGPT (OpenAI, GPT-5.5 Thinking) for preliminary figure drafting, visual refinement, and layout optimization. Abbreviations: 4D: Four-dimensional; BP: Black phosphorus; ECM: Extracellular matrix; MMP: Matrix metalloproteinase; MXene: Two-dimensional transition-metal carbide, nitride, or carbonitride; NIR: Near-infrared; rGO: Reduced graphene oxide; ROS: Reactive oxygen species.

Table 2. Stimulus–response mechanisms and cardiovascular relevance of smart hydrogel bioinks

Bioink type	Trigger	Representative materials	Cardiovascular application	Atherosclerosis relevance
ROS-responsive ^{25,26,68,74}	ROS	Thioketal polymers, BP	Controlled degradation, drug release	High
Enzyme-responsive ^{98,117}	MMPs, elastases	Protease-cleavable hydrogels	ECM remodeling	High
Electrically responsive ^{101,118–121}	Electrical stimulation	Graphene, rGO, conductive polymers	Signal transmission, sensing	Moderate
Photoresponsive ^{101,107}	NIR/light	BP, germanene, photothermal agents	Remote regulation	Moderate
Multifunctional ^{53,68,114,122}	Multiple stimuli	Hybrid nanomaterial systems	Disease-adaptive regulation	High

Abbreviations: BP: Black phosphorus; ECM: Extracellular matrix; MMPs: Matrix metalloproteinases; NIR: Near-infrared; rGO: Reduced graphene oxide; ROS: Reactive oxygen species.

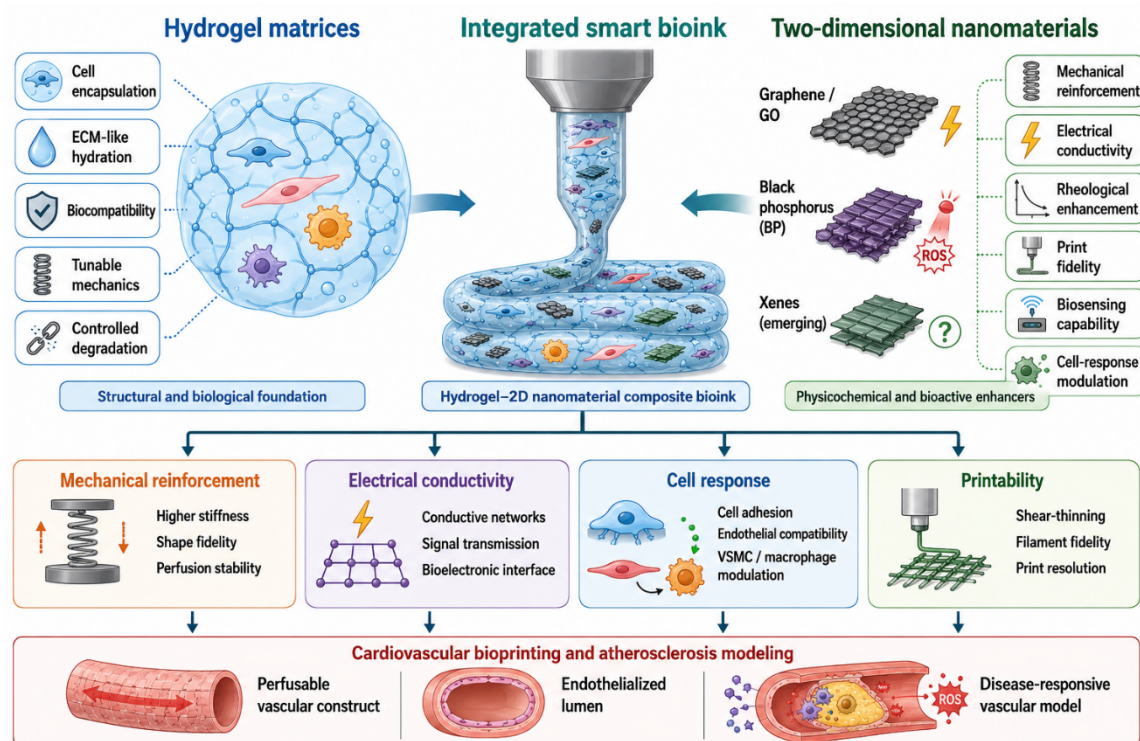


Figure 3. Complementary roles of hydrogel matrices and 2D nanomaterials in smart bioink systems. Hydrogel matrices provide the foundational ECM-like framework for cell encapsulation, hydration, biocompatibility, tunable mechanics, and controlled degradation. 2D nanomaterials serve as physicochemical and bioactive enhancers that improve mechanical reinforcement, electrical conductivity, rheological behavior, print fidelity, biosensing capability, and cell-response modulation. Their integration enables multifunctional smart bioinks with enhanced structural stability, bioelectronic functionality, cellular regulation, and printability for cardiovascular bioprinting and atherosclerosis modeling. Created in BioRender. Chu, R. (2026). <https://BioRender.com/ia11euo>, with assistance from ChatGPT (OpenAI, GPT-5.5 Thinking) for preliminary figure drafting, visual refinement, and layout optimization. All figure concepts, scientific labels, annotations, and final visual content were reviewed, revised, and approved by the authors.

Abbreviations: ECM: Extracellular matrix; GO: Graphene oxide.

For cardiovascular bioink design, graphene is attractive not because of a single material property but because it can simultaneously influence multiple aspects of biofabrication. Depending on the derivative selected, graphene-based additives may improve rheological behavior, mechanical reinforcement, electrical conductivity, surface bioactivity, and biosensing capability.^{28,123} Consequently, different graphene derivatives are typically chosen according to the intended function of the bioink rather than conductivity alone.

Graphene oxide is the most commonly used derivative for hydrogel bioinks because its oxygen-containing functional groups improve aqueous dispersibility and facilitate interactions with polymer networks through hydrogen bonding, electrostatic interactions, and physical entanglement. These characteristics make GO particularly suitable for enhancing printability and structural stability.¹²³ In contrast, rGO partially restores the conductive sp^2 carbon network and is therefore preferred

for applications requiring electrical signal transmission and electrophysiological coupling. rGO-gelatin methacryloyl (GelMA) hydrogels have been widely investigated as conductive cardiac scaffolds that improve electrical and mechanical properties.¹²⁴ This distinction is important for bioink design because vascular applications may prioritize dispersion, printability, and cytocompatibility, whereas cardiac applications often require higher conductivity and electromechanical coupling.¹¹⁹

From a formulation perspective, graphene derivatives can be viewed as functional nanofillers that provide distinct advantages depending on the target application. For vascular bioinks, GO is primarily utilized to improve printability, support endothelial-cell compatibility, and enhance scaffold stability. For cardiac bioinks, rGO is commonly incorporated to enhance conductivity and tissue maturation. For biosensing-enabled platforms, graphene derivatives may serve as conductive interfaces

that facilitate real-time monitoring of cellular and biochemical activities. Therefore, rational selection of graphene derivatives should be guided by the specific requirements of the target cardiovascular application rather than by a generalized assumption that all graphene materials provide identical functions.

4.2. Rheology, mechanics, and printability

One of the most established contributions of graphene to bioink development is the enhancement of rheological properties and mechanical performance. Conventional hydrogels often suffer from insufficient mechanical strength, poor shape fidelity, and structural instability during long-term culture or perfusion. These limitations are particularly problematic for cardiovascular applications, where constructs must withstand extrusion, handling, fluid flow, and cyclic mechanical loading.^{81,125} Graphene nanosheets possess exceptionally large surface areas and high aspect ratios, allowing extensive interactions with polymer chains through hydrogen bonding, electrostatic attraction, van der Waals forces, and π - π interactions. As a result, graphene incorporation can increase viscosity, improve shear-thinning behavior, enhance post-shear recovery, and reinforce hydrogel networks.^{126,127}

Numerous studies have demonstrated improved printability and mechanical performance following graphene incorporation. GO-containing GelMA, collagen, alginate, and gelatin-based bioinks have shown enhanced shape fidelity, filament stability, compressive strength, and viscoelastic behavior.^{123-126,128} In extrusion-based bioprinting, graphene additives frequently improve filament formation and reduce structural collapse after deposition, facilitating fabrication of multilayered vascular constructs and perfusable architectures.

Despite these advantages, graphene concentration must be carefully optimized. Excessive loading may induce aggregation, heterogeneous mechanical properties, nozzle clogging, and reduced cellular compatibility.^{127,128} Furthermore, improvements in bulk mechanical properties do not necessarily translate into improved biological performance, emphasizing the need for simultaneous evaluation of rheological and cellular outcomes. Current evidence suggests that graphene is one of the most effective nanofillers available for improving bioink printability and structural stability. Consequently, its greatest immediate value in vascular bioprinting lies in mechanical reinforcement and fabrication support rather than disease-specific biological regulation.

4.3. Conductive cardiac bioinks

Electrical conductivity represents another defining

characteristic of graphene-based bioinks. Native myocardium relies on synchronized electrical signal propagation to coordinate contraction, whereas most conventional hydrogels are electrically insulating and therefore poorly suited for supporting electrophysiological communication between cardiomyocytes.^{119,129} By incorporating conductive graphene derivatives, hydrogel matrices can form interconnected conductive networks that facilitate electrical signal transmission and improve intercellular communication.

Graphene-containing hydrogels have been reported to enhance the expression of cardiac maturation markers such as connexin-43, cardiac troponin T, and α -actinin while improving calcium handling and synchronized beating behavior in engineered cardiac tissues.^{124,130,131} In particular, rGO-GelMA hydrogels have demonstrated improved sarcomere organization and electrophysiological coupling, supporting functional maturation of bioprinted cardiac constructs.¹²⁴

This conductive function is highly relevant for cardiovascular bioprinting because the maturation of engineered cardiac tissues remains a major bottleneck in the development of clinically relevant tissue models. Recent studies have highlighted that conductive graphene-based biomaterials can improve electrophysiological integration, promote anisotropic tissue organization, and generate more physiologically representative cardiac tissue models for disease modeling and drug screening applications.^{129,132} These findings have established graphene as one of the most extensively studied conductive additives for cardiac tissue engineering and bioprinting.

However, most available evidence originates from myocardial applications rather than vascular disease models. Conductivity alone cannot overcome other challenges associated with tissue maturation, including vascularization, nutrient transport, mechanical matching, and long-term tissue remodeling.^{119,133} Although conductive graphene bioinks provide valuable proof-of-concept for electroactive biofabrication, their direct relevance to atherosclerosis modeling remains limited. Their primary contribution lies in demonstrating how nanomaterial-enabled conductivity can be integrated into multifunctional cardiovascular bioinks.¹²⁹

4.4. Vascular endothelialization and angiogenesis

Beyond cardiac applications, graphene-based materials can influence endothelial behavior through modulation of surface chemistry, nanotopography, mechanical properties, and electrical characteristics. Endothelial cells are highly responsive to these environmental cues, making graphene an attractive component for vascular bioinks.¹³⁴

Previous studies have reported enhanced endothelial adhesion, proliferation, migration, and spreading on graphene-containing substrates and hydrogels.¹³⁵ A collagen/GO bioink developed for 3D-bioprinted vascular graft fabrication demonstrated improved mechanical performance and favorable endothelial-cell compatibility.¹²⁸ In addition, graphene-based systems have been associated with vascular endothelial growth factor-related angiogenic activity and phosphoinositide 3-kinase/protein kinase B-mediated signaling pathways that regulate endothelial survival, migration, and vascular formation.¹³⁶ Such effects may facilitate microvascular network formation and improve oxygen and nutrient transport within thick bioprinted tissues.¹³⁷

Most current studies evaluate endothelial viability and proliferation rather than disease-relevant endothelial dysfunction. However, endothelial dysfunction is a central event in atherosclerosis and involves alterations in barrier integrity, inflammatory activation, leukocyte recruitment, and NO signaling. Therefore, favorable endothelial growth does not necessarily indicate successful disease modeling. Future atherosclerosis-oriented bioinks should assess endothelial barrier function, VCAM-1 and ICAM-1 expression, monocyte adhesion, permeability changes, and responses to disturbed flow rather than relying solely on conventional cell viability measurements.¹³⁸

4.5. Biosensing-enabled disease monitoring

Among the various applications of graphene in cardiovascular biofabrication, biosensing may represent its most distinctive and underexploited capability. Graphene exhibits high electrical conductivity, large surface area, excellent surface functionalization potential, and remarkable sensitivity to interfacial chemical changes, making it highly suitable for bioelectronic sensing.^{121,127,139} Graphene-based sensing platforms have been explored for monitoring cellular activity, metabolic changes, inflammatory signaling, oxidative stress, and endothelial barrier function.¹⁴⁰⁻¹⁴²

In CVD models, continuous assessment of endothelial dysfunction, cytokine secretion, thrombosis-related responses, and inflammatory activation is increasingly recognized as essential for understanding disease progression.⁷⁹ Despite considerable interest, fully integrated sensing bioinks remain relatively uncommon. Most current graphene-containing hydrogels focus primarily on mechanical reinforcement, printability, or conductivity rather than embedded sensing functionality.^{127,139,143} Consequently, real-time monitoring remains largely

separated from the biofabrication process itself. For atherosclerosis modeling, biosensing may represent the most unique contribution of graphene. Sensor-integrated vascular constructs could transform disease modeling from endpoint-based analysis toward continuous and non-destructive monitoring of pathological progression.

4.6. Design boundaries and future directions

Despite its versatility, graphene should not be regarded as a universal solution for atherosclerosis bioink design. Graphene performs exceptionally well as a mechanical stabilizer, conductive component, endothelial-supportive additive, and biosensing interface.^{123,128,144,145,146} These properties directly address several challenges associated with vascular bioprinting, including printability, structural integrity, bioelectronic integration, and real-time monitoring.⁸¹ However, graphene exhibits important limitations when viewed from the perspective of disease-responsive bioink design.

Unlike ROS-responsive materials, graphene does not intrinsically respond to oxidative stress, inflammatory signaling, or disease-associated biochemical changes.¹⁴⁷⁻¹⁴⁹ Furthermore, many graphene derivatives exhibit limited biodegradability and may persist within biological environments for extended periods.^{148,150} Consequently, graphene alone cannot adequately reproduce the dynamic oxidative and inflammatory microenvironments characteristic of atherosclerotic plaques.⁹⁰

From an application standpoint, graphene is most suitable for four major scenarios: (i) mechanically stable perfusable vascular constructs, (ii) conductive cardiovascular tissues, (iii) biosensor-integrated vascular disease models, and (iv) multifunctional hybrid bioinks. Among these, hybrid systems are likely to be particularly important.^{54,55} Combining graphene with ROS-responsive hydrogels, degradable matrices, or complementary nanomaterials may overcome the limitations of single-component designs while integrating structural stability, conductivity, biosensing capability, and disease responsiveness within a single platform.^{149,151}

Therefore, graphene should be viewed primarily as a structural, conductive, and sensing component in atherosclerosis bioinks, whereas disease-specific responsiveness will likely need to be provided by complementary biomaterials. This distinction is critical for rational material selection and provides a useful framework for evaluating other emerging nanomaterials discussed in subsequent sections.

5. Black phosphorus-based disease-responsive bioinks: Opportunities and translational challenges

5.1. Black phosphorus as a disease-responsive nanomaterial

Among currently available 2D nanomaterials, BP possesses a material profile that is particularly relevant to CVD microenvironments. Unlike graphene-family materials, which are primarily valued for mechanical reinforcement, electrical conductivity, and biosensing capabilities,^{30,31} BP exhibits intrinsic biodegradability, oxidative sensitivity, photothermal responsiveness, and redox-regulatory activity.^{152,153}

This distinction is especially important for atherosclerosis. The disease is characterized by chronic inflammation, oxidative stress, endothelial dysfunction, macrophage activation, ECM remodeling, and progressive plaque destabilization. These pathological features generate disease-specific biochemical signals that may be directly recognized or modulated by BP-containing biomaterials.^{32,55} Recent studies have demonstrated that BP nanosheets possess ROS-scavenging capability, regulate inflammatory signaling pathways, and influence macrophage polarization in CVD models.^{55,154}

Consequently, BP should not be regarded merely as a reinforcing nanofiller. Instead, it represents a potentially active disease-responsive component capable of interacting with pathological microenvironments.^{55,154} This property distinguishes BP from most currently used nanomaterials and makes it particularly attractive for the development of atherosclerosis-oriented smart bioinks.^{85,132}

5.2. Physicochemical properties and bioink formulation considerations

Black phosphorus consists of phosphorus atoms arranged in a puckered layered structure, giving rise to unique optical, electronic, and physicochemical properties.^{155,156} When exfoliated into nanosheets or quantum dots, BP exhibits high surface area, NIR responsiveness, and thickness-dependent electronic behavior.^{157,158} These characteristics provide a physicochemical foundation for constructing photoresponsive and multifunctional hydrogel bioinks.^{122,159}

A major advantage of BP is its biodegradability. Under physiological conditions, BP gradually oxidizes and degrades into phosphate-containing products that may participate in normal biological processes and tissue remodeling.^{158,160,161} This characteristic potentially reduces long-term accumulation concerns associated with nondegradable nanomaterials.^{160,161}

From a bioink formulation perspective, BP nanosheets can interact with polymer networks through hydrogen bonding, electrostatic interactions, physical adsorption, and coordination effects, enabling incorporation into commonly used hydrogel systems such as GelMA, hyaluronic acid, alginate, chitosan, and composite polymer matrices.^{115,122,159} However, the same oxidative sensitivity that makes BP biologically responsive also creates important formulation challenges. BP readily undergoes degradation during preparation, storage, and processing. Variations in nanosheet size, oxidation state, surface chemistry, and storage conditions may significantly influence biological performance and reproducibility.^{159,162} Therefore, BP-based bioinks require careful optimization of exfoliation methods, stabilization strategies, polymer encapsulation, and storage protocols. Importantly, these characteristics simultaneously create opportunities for disease responsiveness and challenges for bioink standardization.

5.3. Reactive oxygen species responsiveness and atherosclerosis-relevant microenvironment regulation

Reactive oxygen species are among the most important pathological drivers of atherosclerosis. Excessive ROS production contributes to endothelial dysfunction, LDL oxidation, macrophage activation, foam-cell formation, VSMC phenotypic switching, inflammatory amplification, and plaque destabilization.²⁵

Because BP is intrinsically sensitive to oxidative environments, its degradation behavior is closely linked to disease-associated redox conditions. In contrast to conventional nanomaterials that remain relatively inert under oxidative stress, BP undergoes oxidation and gradual degradation in the presence of ROS, generating phosphate-containing degradation products.¹⁶³ This property creates a unique opportunity to couple material behavior directly to pathological microenvironments.

Recent studies have demonstrated that BP-containing hydrogels can regulate oxidative stress, attenuate inflammatory responses, and promote tissue repair in CVD models. For example, ROS-responsive Gel-pBP@Mg hydrogels enabled ROS-triggered degradation, sustained BP release, reduced oxidative stress, suppressed inflammatory signaling, and promoted angiogenesis following myocardial infarction.⁵⁵ Similar BP-based systems have been reported to regulate macrophage polarization and improve tissue regeneration through redox-dependent mechanisms.^{154,164}

Although most available evidence originates from myocardial repair studies,¹⁶⁵ the underlying mechanisms

are highly relevant to atherosclerosis because oxidative stress and inflammation represent common pathological pathways. Consequently, BP-integrated bioinks may enable the construction of vascular models in which oxidative stress gradients dynamically regulate matrix behavior, cellular responses, and therapeutic release. Among currently available 2D nanomaterials, BP is arguably the material whose intrinsic responsiveness most closely aligns with the pathological microenvironment of atherosclerosis. This disease-matched responsiveness represents its greatest advantage for smart bioink development.

5.4. Photothermal regulation and external control

Black phosphorus nanosheets exhibit strong NIR absorption and efficient photothermal conversion, enabling localized heat generation under external optical stimulation. This property has attracted considerable interest in drug delivery, tissue repair, and regenerative medicine applications.¹⁶⁶ BP-based nanospheres/nanosheets have been reported as biodegradable photothermal agents for *in vivo* cancer photothermal therapy and cancer theranostics, while recent reviews further summarize their antibacterial and tissue-regenerative applications.¹⁵⁸

Within hydrogel systems, photothermal responsiveness provides an opportunity for remote control of matrix degradation, molecular release, and cellular activity. Compared with chemically triggered systems, light-mediated regulation offers superior temporal and spatial precision. BP-containing hydrogels have demonstrated NIR-triggered release of bioactive molecules and externally controlled modulation of material behavior.^{74,167} However, the relevance of photothermal regulation to atherosclerosis modeling remains less clear than its relevance to ROS-responsive regulation. Excessive photothermal stimulation may impair endothelial viability, compromise vascular barrier integrity, and induce inflammatory responses. Recent studies have shown that BP nanosheets may influence endothelial permeability, transendothelial electrical resistance (TEER) values, and junctional protein expression under certain conditions.¹⁶⁸

Therefore, endothelial safety must be rigorously evaluated when developing photoresponsive BP bioinks. Future studies should assess endothelial viability, barrier integrity, vascular endothelial cadherin (VE-cadherin) expression, zonula occludens-1 (ZO-1) organization, inflammatory activation, and macrophage responses under photothermal conditions rather than relying solely on standard cytocompatibility assays.¹⁶⁷ At present, photothermal responsiveness should be regarded as a potentially useful auxiliary function rather than the primary rationale for BP incorporation into atherosclerosis-oriented bioinks.

5.5. Angiogenesis and vascular remodeling

Black phosphorus-containing biomaterials have demonstrated significant pro-angiogenic potential in multiple regenerative medicine studies. Controlled BP degradation, redox modulation, and interactions with endothelial cells have been associated with enhanced endothelial migration, neovascularization, and tissue repair.^{55,169}

For tissue engineering applications, angiogenesis is generally viewed as a beneficial outcome because vascularization improves oxygen and nutrient transport within thick engineered tissues. BP-integrated hydrogels have therefore been widely investigated for promoting vascular regeneration in myocardial repair and other ischemic conditions.^{55,169,170} However, angiogenesis has a dual role in atherosclerosis. While vascularization may support tissue regeneration, intraplaque neovascularization is strongly associated with plaque progression, hemorrhage, inflammation, and plaque vulnerability.^{171,172} Consequently, increased angiogenesis should not automatically be interpreted as a favorable outcome in atherosclerosis-oriented bioink design.

This distinction highlights the importance of disease-specific evaluation criteria. Rather than simply measuring vascular growth, future BP-based atherosclerosis models should investigate how BP influences pathological neovascularization, endothelial dysfunction, inflammatory activation, and plaque remodeling.

5.6. Translational gap between therapeutic hydrogels and true bioinks

A major limitation of the current BP literature is that most reported systems have been developed as injectable therapeutic hydrogels rather than bioprintable bioinks. Existing studies provide substantial evidence supporting the biological activity of BP-containing hydrogels, including ROS scavenging, inflammatory regulation, macrophage modulation, angiogenesis promotion, and tissue repair.^{55,154,164,173} However, these studies typically focus on therapeutic efficacy after injection and rarely evaluate the material properties required for bioprinting.

Consequently, critical biofabrication parameters remain insufficiently characterized, including viscosity, shear-thinning behavior, yield stress, filament formation, extrusion fidelity, crosslinking kinetics, post-printing cell viability, long-term construct stability, and perfusion compatibility.¹⁷⁴⁻¹⁷⁶ This distinction is important because therapeutic performance should not be equated with biofabrication suitability. A hydrogel that performs well as an injectable matrix may fail to satisfy the rheological and biological requirements necessary for successful

bioprinting. Therefore, the current evidence base supports BP primarily as a promising therapeutic hydrogel component rather than as a fully validated cardiovascular bioink additive.

5.7. Design requirements for black phosphorus-based atherosclerosis bioinks

To realize the full potential of BP in cardiovascular bioprinting, future studies must simultaneously address disease relevance and biofabrication performance. From a biofabrication perspective, BP-based systems should be evaluated for extrusion rheology, shear-thinning behavior, viscosity recovery, filament fidelity, print resolution, post-printing viability, and long-term structural stability under perfusion. These parameters remain largely absent from the current literature.¹⁷⁷⁻¹⁷⁹

From a disease-modeling perspective, evaluation should extend beyond conventional cytocompatibility measurements. Future BP-containing bioinks should be validated under oxidative stress, inflammatory stimulation, oxLDL exposure, macrophage co-culture, disturbed-flow conditions, and long-term vascular perfusion.^{65,180,181}

Particular attention should be paid to functional endpoints directly associated with disease progression, including ROS-responsive degradation behavior, endothelial barrier integrity and permeability, expression of junctional proteins such as VE-cadherin and ZO-1, endothelial activation markers including VCAM-1 and ICAM-1, macrophage polarization profiles, inflammatory signaling pathways, VSMC phenotypic switching, and construct stability under physiologically relevant flow conditions.^{9,44,55,65,180} These parameters collectively provide a more rigorous framework for evaluating the biological relevance and translational potential of BP-enabled vascular disease models.

Ultimately, BP-containing hydrogels should not be considered atherosclerosis-specific bioinks solely on the basis of their antioxidant or photothermal properties. Rather, only materials that simultaneously satisfy stringent biofabrication requirements and faithfully reproduce disease-associated vascular microenvironments can be regarded as validated BP-containing bioinks for atherosclerosis modeling and cardiovascular bioprinting.

5.8. Summary and comparison with graphene

The distinct roles of graphene and BP in cardiovascular bioink design are summarized in [Figure 4](#) and are further compared with other representative 2D nanomaterials in [Section 6.4](#). Overall, graphene primarily contributes structural stability, conductivity, and biosensing functionality,^{29,118,123,182} whereas BP contributes disease-

responsive, redox-regulatory, and immunomodulatory capabilities.^{55,115,154} Rather than functioning as competing nanomaterial platforms, graphene and BP provide complementary functionalities that may be integrated within multifunctional bioinks.

For atherosclerosis modeling, graphene may be most useful for constructing mechanically stable and sensor-integrated vascular platforms, whereas BP may be more suitable for reproducing or modulating redox-active plaque-like microenvironments, given the central role of oxidative stress and ROS-responsive biomaterials in cardiovascular pathology.^{65,92,183} Therefore, graphene and BP should not be viewed as competing materials, but as complementary modules within multifunctional bioinks. Hybrid graphene-BP systems may integrate conductivity, mechanical reinforcement, biosensing capability, ROS responsiveness, biodegradability, and photothermal regulation; this concept is supported by recent GO/BP composite scaffold studies, although direct validation in atherosclerosis-on-a-chip bioinks remains an important future direction.¹⁸⁴ Such systems could be especially powerful for atherosclerosis-on-a-chip platforms requiring both real-time monitoring and disease-responsive remodeling.^{65,185}

6. Emerging Xenes beyond graphene and black phosphorus

6.1. Why look beyond graphene and black phosphorus?

To clarify the distinct functional roles of different 2D nanomaterials, [Figure 5](#) compares graphene, BP, and emerging Xenes in terms of disease responsiveness, multifunctionality, and translational maturity. Graphene and BP currently represent the two most extensively investigated 2D nanomaterials for cardiovascular bioink development.^{29,119,154} As discussed in the preceding sections, graphene primarily contributes mechanical reinforcement, electrical conductivity, and biosensing capability, whereas BP provides biodegradability, ROS responsiveness, and disease-adaptive functionality. Together, these materials address many of the structural and biological challenges associated with cardiovascular bioprinting and atherosclerosis modeling.

Nevertheless, neither material fully satisfies all requirements of next-generation smart bioinks. Graphene exhibits limited disease-specific responsiveness, while BP faces challenges related to instability, reproducibility, and bioprinting validation. Consequently, increasing attention has been directed toward emerging members of the Xene family as potential candidates for future multifunctional bioink systems.

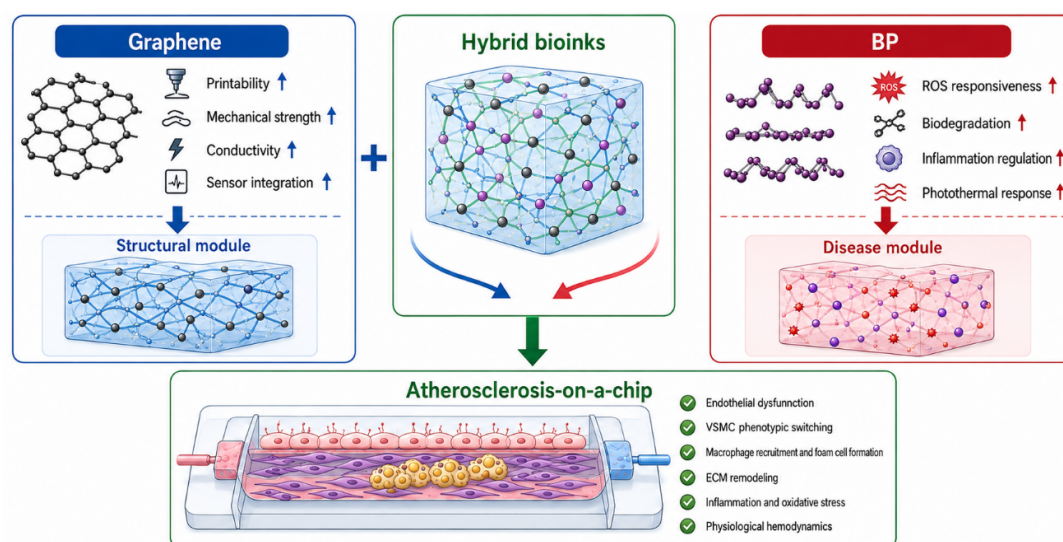


Figure 4. Complementary rather than competitive roles of graphene and BP in smart bioinks. Graphene- and BP-based nanomaterials contribute distinct but complementary functionalities to cardiovascular bioink design. Graphene primarily serves as a structural and bioelectronic component by enhancing mechanical reinforcement, printability, electrical conductivity, and biosensing capability, thereby supporting construct stability, electrophysiological integration, and real-time monitoring. In contrast, BP functions predominantly as a disease-responsive component owing to its biodegradability, ROS-sensitive behavior, photothermal responsiveness, and potential immunomodulatory effects. These properties enable dynamic regulation of oxidative stress, inflammation, and tissue remodeling within pathological vascular microenvironments. Rather than representing competing nanomaterial platforms, graphene and BP provide complementary modules that may be integrated within multifunctional smart bioinks to simultaneously improve biofabrication performance and disease-specific responsiveness for advanced cardiovascular tissue engineering and atherosclerosis modeling. Created in BioRender. Chu, R. (2026). <https://BioRender.com/4iakeoo>, with assistance from ChatGPT (OpenAI, GPT-5.5 Thinking) for preliminary figure drafting, visual refinement, and layout optimization.

Abbreviations: BP: Black phosphorus; ECM: Extracellular matrix; ROS: Reactive oxygen species; VSMC: Vascular smooth muscle cell.

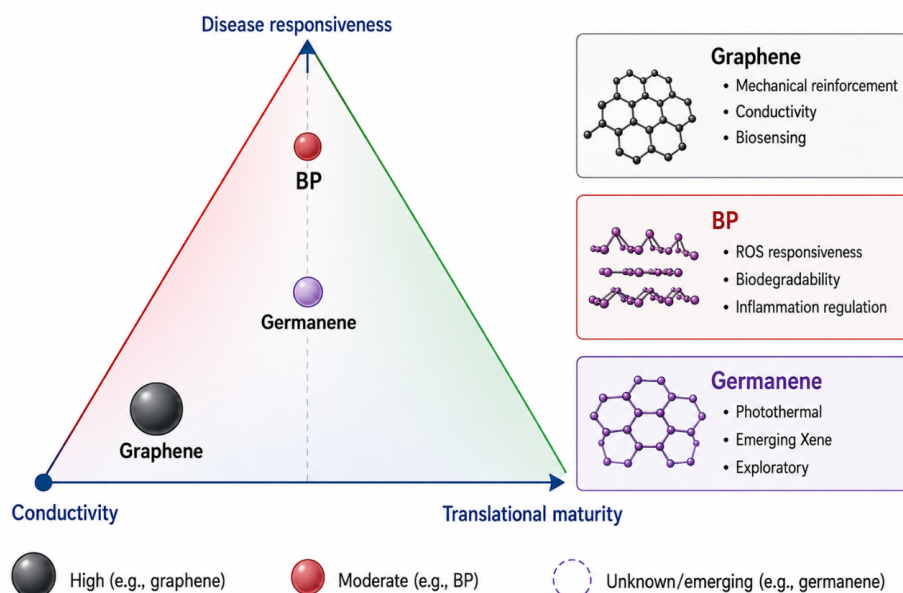


Figure 5. Functional positioning of graphene, BP, and emerging Xenes in cardiovascular bioink design. Graphene primarily contributes conductivity, mechanical reinforcement, and biosensing functionality. BP provides biodegradability, ROS responsiveness, photothermal activity, and angiogenic potential. Germanene represents an emerging Xene platform with photothermal and programmable functionalities but currently has limited cardiovascular evidence. Created in BioRender. Chu, R. (2026). <https://BioRender.com/5lvir0n>, with assistance from ChatGPT (OpenAI, GPT-5.5 Thinking) for preliminary figure drafting, visual refinement, and layout optimization.

Abbreviations: BP: Black phosphorus; ROS: Reactive oxygen species.

Among these emerging materials, germanene has attracted particular interest because of its photothermal properties, surface reactivity, and potential compatibility with hydrogel-based biomaterials.^{186–188} However, compared with graphene and BP, its biomedical development remains at a much earlier stage.^{189,190} Therefore, germanene should currently be regarded as a frontier material that may expand future bioink-design strategies rather than as an established component of cardiovascular bioprinting platforms.

6.2. Germanene and emerging Xenes: Exploratory opportunities

Germanene is a 2D germanium-based nanomaterial belonging to the broader Xene family. Although originally investigated for electronic and photonic applications, recent studies have suggested potential biomedical utility owing to its photothermal responsiveness, surface functionalization capability, and compatibility with hydrogel systems.^{186–188} From a bioink-design perspective, the most relevant features of germanene are not its electronic structure alone but its potential contribution to externally regulated and multifunctional hydrogel systems. First, germanene exhibits efficient NIR photothermal conversion, enabling remote modulation of material behavior.¹⁹¹ Second, its chemically active surface may facilitate conjugation with polymers, peptides, therapeutic molecules, or targeting ligands.^{192–194} Third, emerging studies suggest that germanene may eventually support sensing or bioelectronic applications if stable and biocompatible interfaces can be established.¹⁹⁵

Collectively, these properties suggest that germanene and related Xenes could contribute to future hydrogel systems requiring remote regulation, multifunctional surface modification, or integrated sensing functions. However, these possibilities remain largely conceptual and have not yet been validated in cardiovascular biofabrication.^{81,196,197}

Importantly, the currently available germanene-containing hydrogel evidence should be interpreted cautiously. The strongest biomedical evidence remains derived mainly from antibacterial and wound-healing applications rather than vascular tissue engineering or atherosclerosis modeling. For example, germanene-modified chitosan hydrogels have demonstrated NIR-triggered photothermal antibacterial activity and accelerated healing in infected skin wound models.³⁶ Although such studies provide useful proof-of-concept evidence that germanene can be incorporated into hydrogel matrices, they do not establish germanene

as a vascular bioink component. Critical parameters required for cardiovascular bioprinting—including bioink rheology, shear-thinning behavior, extrusion fidelity, endothelial-cell compatibility, immune-cell interactions, and long-term perfusion stability—remain insufficiently investigated.^{81,86,198}

Therefore, the discussion of germanene in this review is intentionally positioned as exploratory rather than confirmatory. At present, there is no direct evidence demonstrating germanene-enabled vascular bioinks, cardiac bioinks, atherosclerosis-on-a-chip platforms, or bioprinted CVD models. This distinction is important to avoid overextending conclusions from non-cardiovascular hydrogel systems to vascular biofabrication.

6.3. Evidence gaps and bioink validation requirements

Several major barriers must be addressed before germanene or other emerging Xenes can be considered practical components of cardiovascular bioinks. Material stability remains one of the most significant challenges. Germanene is susceptible to oxidation and structural degradation under ambient and aqueous conditions,^{191,199,200} potentially affecting reproducibility, biological activity, and long-term functionality. Stable formulations compatible with hydrogel precursor solutions have not yet been systematically established.

Uniform dispersion within hydrogel precursor solutions is also essential for maintaining consistent mechanical and biological performance.^{188,189,201} Aggregation of germanene nanosheets may result in heterogeneous material properties, nozzle clogging, uneven photothermal responses, and concentration-dependent cytotoxicity. Effective stabilization and surface-modification strategies will therefore be required before any cell-laden printing application can be realistically considered.

Because current germanene-based hydrogel systems have not been validated as vascular bioinks, cytocompatibility should not be presented as an established cardiovascular function. Instead, future studies should first evaluate basic vascular-cell compatibility using endothelial cells, VSMCs, macrophages, and cardiomyocytes, together with functional endpoints such as endothelial barrier integrity, inflammatory activation, oxidative-stress response, macrophage polarization, and long-term cellular functionality.^{17,81} These assays should be interpreted as validation requirements rather than evidence of current applicability.

Perhaps the most important limitation is the complete lack of bioprinting validation. To date, no germanene-containing system has been systematically characterized as a cardiovascular bioink. Essential biofabrication parameters—including viscosity, shear-thinning behavior, viscosity recovery, filament fidelity, extrusion pressure, crosslinking compatibility, post-printing cell viability, and long-term perfusion stability—remain unknown.^{81,86,202,203} Accordingly, germanene currently remains a hydrogel-compatible nanomaterial with exploratory potential rather than a validated bioink additive for cardiovascular bioprinting.^{187,204}

Future research should therefore proceed in a stepwise manner: first establishing stable and dispersible germanene formulations, then confirming compatibility with cell-laden hydrogels, and finally evaluating vascular-relevant functions under oxidative stress, inflammatory stimulation, endothelial dysfunction, and dynamic perfusion conditions.^{204,205} Only after these criteria are met can germanene-containing systems be reasonably assessed for atherosclerosis modeling or cardiovascular biofabrication.

6.4. Comparison with graphene and black phosphorus

Graphene, BP, and germanene occupy distinct positions in the development landscape of 2D nanomaterial-integrated bioinks. Among these materials, graphene represents the most mature platform. Extensive studies have demonstrated its ability to improve printability, mechanical strength, electrical conductivity, and cellular responses in biofabricated constructs, supporting applications in tissue engineering, regenerative medicine, and biosensing. Recent reviews have highlighted graphene-based nanomaterials as one of the most extensively investigated 2D material families for 3D bioprinting and bioink development.²⁹ Furthermore, conductive graphene- and GO-based bioinks have shown promise for enhancing cell adhesion, electrical signal propagation, and functional maturation in cardiac tissue engineering applications.^{123,206}

Compared with graphene, BP is less mature from a translational perspective but possesses stronger disease-oriented relevance owing to its intrinsic biodegradability, ROS-responsive behavior, photothermal properties, and potential pro-angiogenic effects. Recent studies have emphasized BP as a multifunctional biomaterial capable of regulating oxidative stress, promoting tissue regeneration, and facilitating controlled therapeutic delivery.¹¹⁵ BP-incorporated hydrogels have demonstrated beneficial effects in diabetic wound healing through

combined antibacterial and antioxidant mechanisms,²⁰⁷ while ROS-responsive BP-based hydrogel systems have shown promising therapeutic efficacy in myocardial infarction repair by enhancing angiogenesis and cardiac tissue regeneration.⁵⁵

In contrast, germanene remains the least developed of these three categories of 2D materials for biomedical applications. Current evidence is largely limited to photothermal antibacterial hydrogel systems, particularly germanene-modified chitosan hydrogels designed for infected wound treatment.³⁶ Although germanene may offer unique surface reactivity and optoelectronic characteristics relative to graphene due to its buckled atomic structure and semiconductor-like behavior, it currently lacks direct evidence in vascular bioinks, CVD models, or atherosclerosis-on-a-chip platforms. Consequently, germanene should not be considered a direct competitor to graphene or BP at the present stage of development. Its more realistic role is that of a long-term frontier material that may broaden the functional diversity of smart bioinks only after physicochemical stability, aqueous dispersibility, cytocompatibility, bioprintability, and perfusion compatibility have been systematically addressed.²⁷ The functional positioning, evidence maturity, and translational readiness of the principal 2D nanomaterials discussed in this review are compared in Table 3.

7. Integrated atherosclerosis-on-a-chip and patient-specific bioprinted models

7.1. From biomaterials to integrated disease models

Atherosclerosis is a spatially heterogeneous and temporally progressive vascular disease involving coordinated interactions among endothelial cells, VSMCs, macrophages, lipoproteins, ECM components, and hemodynamic forces.³ Conventional 2D cell cultures and animal models have provided important mechanistic insights, but they cannot fully reproduce the 3D architecture, human-specific cellular interactions, or dynamic microenvironment of human atherosclerotic plaques.³

Recent advances in 3D bioprinting and OOC technologies have enabled the development of engineered vascular disease systems that more closely recapitulate human pathophysiology.^{14,79} These platforms allow spatially controlled positioning of vascular cells, immune cells, biomaterials, and disease-associated biochemical stimuli within perfusable architectures. Combined with microfluidic flow systems, they provide opportunities to reproduce endothelial mechanobiology, inflammatory

Table 3. Comparative physicochemical, biological, printability, and cardiovascular features of representative 2D nanomaterials for smart bioinks

2D nanomaterial	Key physicochemical properties	Biological functions	Printability enhancement	Cardiovascular/atherosclerosis applications	Evidence and limitations
GO ^{123,128,143,208}	Hydrophilic oxygenated sheets; large surface area; strong polymer interactions	Supports adhesion and endothelial-cell compatibility; may modulate inflammatory responses	Improves viscosity, shear-thinning behavior, filament fidelity, and scaffold stability	Perfusable vascular constructs; endothelialized scaffolds; sensor-compatible matrices	Strong bioink evidence; dose, purity, and long-term persistence require control
rGO/graphene nanoplatelets ^{101,124,209}	Partly restored sp ² network; higher conductivity; lower dispersibility than GO	Enhances electroactive cell communication and bioelectronic interfaces	Reinforces hydrogels but aggregation can impair homogeneity and cell viability	Conductive cardiac bioinks; vascular biosensing; real-time monitoring	Mature in cardiac models; direct atherosclerosis evidence remains limited
Graphene quantum dots ^{210,211}	Nanoscale carbon dots; high surface functionality; optical activity	Potential antioxidant, imaging, and biosensing functions	Limited reinforcement compared with sheets; may improve dispersion in hydrogels	Bioimaging, sensing interfaces, and adjunctive bioink additives	Promising but less validated as structural bioink components
BP ^{32,54,55,154}	Puckered degradable nanosheets; ROS-sensitive oxidation; NIR photothermal response	ROS regulation, anti-inflammatory activity, macrophage modulation, angiogenesis support	May tune mechanics and degradation, but bioprinting rheology remains underreported	Redox-responsive vascular models; myocardial repair hydrogels; disease-adaptive matrices	High atherosclerosis relevance; stability and reproducibility remain major barriers
MXenes/transition-metal dichalcogenides ²¹²⁻²¹⁴	Conductive or semiconductive layered materials; tunable surface chemistry	Potential photothermal, antibacterial, and sensing functions	Can improve conductivity and mechanics depending on formulation	Bioelectronic scaffolds, photothermal platforms, and sensing-enabled models	Emerging; cardiovascular bioink evidence is still sparse

Abbreviations: 2D: Two-dimensional; BP: Black phosphorus; GO: Graphene oxide; MXene: Two-dimensional transition-metal carbide, nitride, or carbonitride; NIR: Near-infrared; rGO: Reduced graphene oxide; ROS: Reactive oxygen species.

activation, lipid accumulation, and vascular remodeling under physiologically relevant conditions.^{79,215}

Importantly, the objective of next-generation atherosclerosis models is not simply to fabricate vessel-like structures but to reconstruct disease-relevant vascular microenvironments in a controllable and measurable manner.⁶⁵ This requires simultaneous integration of endothelial dysfunction, VSMC remodeling, macrophage infiltration, oxLDL exposure, oxidative stress, ECM remodeling, and disturbed flow within a unified experimental platform.⁶⁵

7.2. A model architecture for next-generation atherosclerosis platforms

An ideal bioprinted atherosclerosis model should integrate

structural, cellular, biochemical, mechanical, and sensing elements within a unified vascular platform. At the luminal interface, endothelial cells form a dynamic barrier that regulates vascular permeability, leukocyte recruitment, NO signaling, and overall vascular homeostasis. Because endothelial dysfunction is one of the earliest and most critical events in atherogenesis, faithful reconstruction of endothelial behavior is essential for disease modeling.^{3,9}

Beneath the endothelium, a VSMC-rich medial layer provides structural support while enabling investigation of key pathological processes, including VSMC phenotypic switching, migration, proliferation, and ECM remodeling during plaque development.^{39-41,77,216,217}

To recapitulate the complexity of atherosclerotic lesions, plaque-mimicking compartments may incorporate

macrophages, foam cells, ECM-like matrices, and lipid-rich microenvironments. These regions can be selectively exposed to oxLDL, pro-inflammatory cytokines, oxidative stress, or matrix-degrading enzymes to reproduce localized plaque progression and heterogeneity.^{66,180,218-220}

The incorporation of smart bioinks can further enhance physiological relevance. ROS-responsive hydrogels may recreate oxidative microenvironments;²⁶ enzyme-responsive matrices may support dynamic ECM remodeling;⁷⁷ graphene-containing systems may facilitate integration with sensing components;^{28,29,31} and BP-containing bioinks may provide redox-responsive regulation and disease-adaptive functionality.⁷⁴

Accordingly, a next-generation atherosclerosis-on-a-chip platform should ideally integrate an endothelialized vascular lumen, a VSMC-containing medial layer, a macrophage-rich plaque niche, oxLDL and inflammatory stimulation, disturbed-flow microfluidic systems, ROS-

responsive or enzyme-responsive matrices, and embedded biosensing modules for continuous monitoring.^{11,76,79} Such a platform would move beyond conventional vascular tissue engineering by reconstructing the key cellular, biochemical, and biomechanical drivers of atherosclerotic progression within a controllable and physiologically relevant *in vitro* system. Based on the concepts discussed above, Figure 6 illustrates an integrated atherosclerosis-on-a-chip platform incorporating multicellular vascular architecture, pathological stimuli, responsive biomaterials, and real-time sensing modules.

To provide a clearer overview of the current modeling landscape, Table 4 compares major atherosclerosis models in terms of their key features, advantages, limitations, representative matrix or bioink compositions, and key supporting references. This comparison highlights that no single model fully recapitulates the cellular complexity, lipid accumulation, inflammatory activation, ECM remodeling, and hemodynamic regulation of human atherosclerosis.

Table 4. Comparative overview of atherosclerosis models and representative bioink or matrix compositions

Model type	Key features	Associated bioink/ matrix composition	Main advantages	Major limitations	Best-fit applications
Conventional 2D vascular cell culture ^{221,222}	Endothelial cells, VSMCs, macrophages, or foam cells cultured on flat substrates; commonly stimulated with oxLDL, TNF- α , IL-1 β , or inflammatory cues	Tissue-culture plastic; collagen-, gelatin-, fibronectin-, or Matrigel-coated surfaces	Simple, low cost, high throughput, easy molecular analysis	Lacks 3D architecture, ECM organization, physiological stiffness, flow, and multicellular spatial interactions	Mechanistic screening of endothelial activation, macrophage lipid uptake, inflammatory signaling, and VSMC phenotypic switching
Static 3D hydrogel-based plaque models ^{223,224}	Vascular cells or immune cells embedded in 3D ECM-like matrices; may incorporate oxLDL, cytokines, ROS, or disease-related remodeling cues	Collagen, fibrin, GelMA, alginate, hyaluronic acid, Matrigel, or dECM-based hydrogels	Better mimics 3D cell-matrix interactions and local inflammatory niches than 2D culture	Limited control over perfusion, shear stress, spatial plaque geometry, and long-term disease progression	Modeling macrophage infiltration, foam-cell formation, ECM remodeling, and cell-matrix crosstalk
Spheroid/pseudo-plaque vascular models ²²⁵⁻²²⁸	Self-assembled or hanging-drop plaque-like aggregates containing inflammatory, stromal, or vascular components	Scaffold-free spheroids; hanging-drop aggregates; Matrigel, collagen, fibrin, or ECM-mimetic matrices	Captures compact multicellular organization and plaque-like cell-cell interactions	Poor control over vascular geometry, luminal structure, perfusion, and mechanical stimulation	Studying multicellular inflammatory responses, pseudo-plaque formation, immune recruitment, and drug screening

(Cont'd...)

Table 4. (Continued)

Model type	Key features	Associated bioink/ matrix composition	Main advantages	Major limitations	Best-fit applications
Microfluidic vascular models ^{12,79,229-231}	Endothelialized channels exposed to laminar, disturbed, or geometry-dependent flow; may include monocyte adhesion and permeability assays	PDMS/glass chips coated with collagen, fibronectin, gelatin, or ECM proteins; hydrogel-patterned channels	Enables controlled shear stress, real-time imaging, dynamic perfusion, and flow-dependent endothelial analysis	Often simplified wall structure; limited incorporation of VSMC-rich media, plaque ECM, and advanced lesion architecture	Studying endothelial dysfunction, leukocyte adhesion, permeability, and shear-stress-regulated inflammatory activation
Atherosclerosis-on-a-chip platform ^{86,76,232,233}	Integrated microfluidic systems combining endothelialized lumen, VSMCs, immune-cell recruitment, oxLDL exposure, and inflammatory stimulation	Collagen, fibrin, GelMA, hyaluronic acid, dECM, or composite hydrogel matrices integrated into microfluidic devices	Reproduces dynamic flow, inflammatory activation, lipid exposure, and multicellular crosstalk in a controllable platform	Technical complexity, limited standardization, variable material compatibility, and difficulty modeling advanced plaque rupture	Disease-mechanism studies, anti-inflammatory drug screening, plaque microenvironment modeling, and patient-specific testing
3D-bioprinted vascular constructs ²³⁴⁻²³⁸	Spatially patterned endothelial cells, VSMCs, fibroblasts, macrophages, and ECM-like matrices within tubular or multilayered vascular structures	GelMA, collagen, fibrin, alginate, hyaluronic acid, dECM, and hybrid hydrogel bioinks	Enables controlled multicellular organization, vessel-like architecture, and customizable geometry	Requires optimized bioink rheology, post-printing viability, lumen perfusion, and long-term construct stability	Engineering vascular wall-like structures, testing bioink performance, and reconstructing early plaque-like microenvironments
Nanomaterial-enabled smart bioink models ²³⁹⁻²⁴²	Hydrogel bioinks integrated with GO/rGO, BP, MXenes, or other 2D nanomaterials to enhance mechanics, conductivity, sensing, ROS regulation, or responsiveness	GelMA/GO, collagen/GO, alginate/GO, BP-containing hydrogels, conductive polymer-graphene composites, or multifunctional hybrid bioinks	Improves print fidelity, mechanical reinforcement, electrical conductivity, biosensing, and disease-responsive regulation	Potential cytotoxicity, aggregation, batch variability, incomplete biodegradation, and limited atherosclerosis-specific validation	Smart vascular bioinks, ROS-responsive disease models, biosensing-enabled constructs, and multifunctional cardiovascular platforms
Patient-specific/iPSC-derived models ^{243,244}	Uses imaging-derived vascular geometry, CFD-guided shear maps, patient-derived cells, or iPSC-derived vascular organoids	Patient-specific cell-laden GelMA, collagen, fibrin, hyaluronic acid, dECM, blood vessel organoids, or composite smart bioinks	High translational relevance; can capture interpatient heterogeneity, individualized geometry, and patient-specific cellular responses	High cost, complex fabrication, limited throughput, and lack of standardized validation criteria	Precision cardiovascular modeling, individualized drug screening, and disease-risk stratification

Abbreviations: 2D: Two-dimensional; 3D: Three-dimensional; BP: Black phosphorus; CFD: Computational fluid dynamics; dECM: Decellularized extracellular matrix; ECM: Extracellular matrix; GelMA: Gelatin methacryloyl; GO: Graphene oxide; IL-1 β : Interleukin 1 beta; iPSC: Induced pluripotent stem cell; MXenes: Two-dimensional transition-metal carbides, nitrides, or carbonitrides; oxLDL: Oxidized low-density lipoprotein; PDMS: Polydimethylsiloxane; rGO: Reduced graphene oxide; ROS: Reactive oxygen species; TNF- α : Tumor necrosis factor alpha; VSMC: Vascular smooth muscle cell.

7.3. Functional readouts and validation criteria

A major challenge in atherosclerosis modeling is determining whether a printed construct genuinely

reproduces disease-relevant phenotypes. Therefore, standardized functional readouts are required. Endothelial integrity should be evaluated using measurements of TEER, permeability, NO production, endothelial nitric

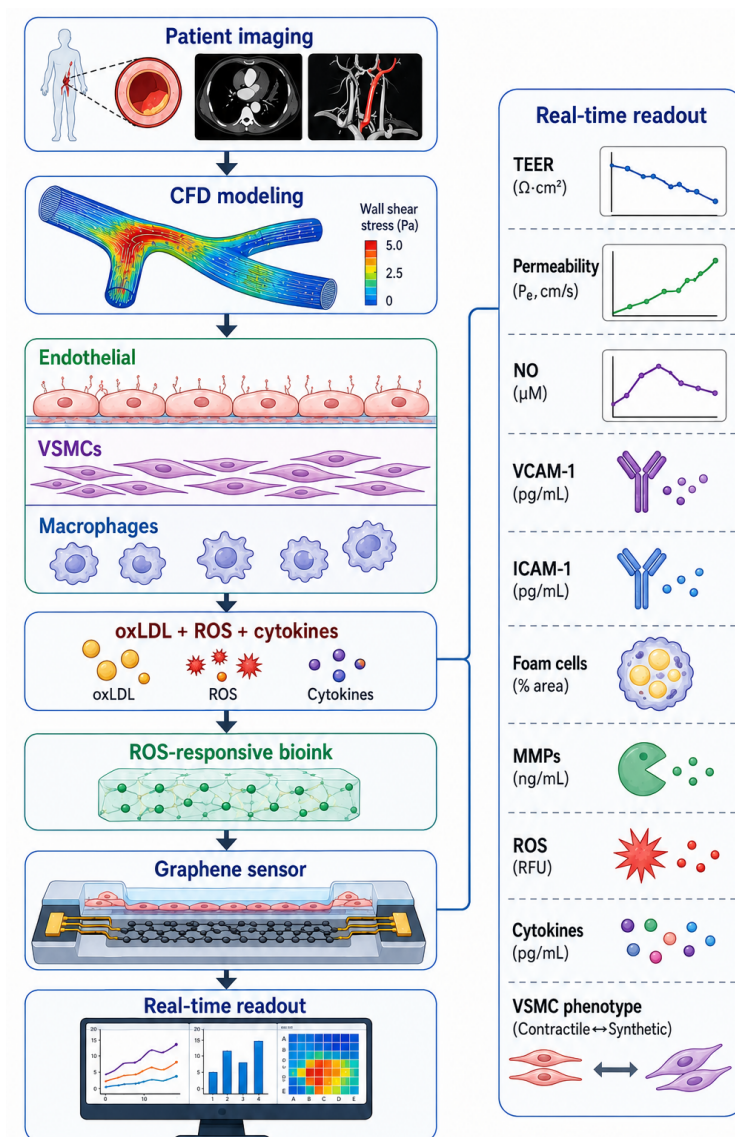


Figure 6. Integrated atherosclerosis-on-a-chip platform. Schematic illustration of a next-generation bioprinted atherosclerosis model integrating disease-relevant cellular, biochemical, mechanical, and sensing components within a perfusable platform. The model consists of an endothelialized lumen, a VSMC-containing medial layer, and a macrophage-rich plaque niche exposed to pro-atherogenic stimuli such as oxLDL, ROS, inflammatory cytokines, and disturbed flow conditions. Smart hydrogel bioinks incorporating stimuli-responsive materials and functional 2D nanomaterials provide dynamic regulation of matrix properties and pathological microenvironments. Integrated biosensing modules enable continuous monitoring of endothelial barrier integrity, inflammatory activation, oxidative stress, extracellular matrix remodeling, and cellular phenotypic changes. By combining 3D bioprinting, OOC technology, microfluidic perfusion, and real-time sensing, the platform enables mechanistic investigation of atherosclerotic progression, therapeutic screening, and the development of patient-specific vascular disease models for precision cardiovascular medicine. This framework represents the conceptual convergence of disease-responsive bioinks, functional nanomaterials, OOC systems, and patient-specific vascular modeling, which together define a future direction for predictive atherosclerosis-on-a-chip platforms. Created in BioRender. Chu, R. (2026). <https://BioRender.com/cgmdnyp>, with assistance from ChatGPT (OpenAI, GPT-5.5 Thinking) for preliminary figure drafting, visual refinement, and layout optimization.

Abbreviations: CFD: Computational fluid dynamics; ICAM-1: Intercellular adhesion molecule 1; MMPs: Matrix metalloproteinases; NO: Nitric oxide; OOC: Organ-on-a-chip; oxLDL: Oxidized low-density lipoprotein; RFU: Relative fluorescence units; ROS: Reactive oxygen species; TEER: Transendothelial electrical resistance; VCAM-1: Vascular cell adhesion molecule 1; VSMC: Vascular smooth muscle cell.

oxide synthase (eNOS) expression, and endothelial barrier function. Inflammatory endothelial activation should be assessed through VCAM-1, ICAM-1, E-selectin, and leukocyte adhesion assays, which represent established hallmarks of endothelial dysfunction.^{66,76,245,246} Macrophage behavior should be monitored through infiltration, lipid uptake, foam-cell formation, CD68 expression, M1/M2-associated markers, cytokine secretion, and ROS production.²⁴⁷⁻²⁵² Monocyte adhesion and transendothelial migration provide additional indicators of inflammatory progression and vascular immune activation.²⁵³

Plaque progression is strongly associated with matrix remodeling and oxidative stress. Relevant endpoints include MMP activity, collagen organization, ROS accumulation, antioxidant responses, and ECM degradation. VSMCs should be evaluated using markers associated with contractile and synthetic phenotypes, including α -SMA, SM-MHC, collagen production, migration, and proliferation.

Collectively, these readouts provide a standardized framework for determining whether bioprinted systems accurately reproduce key aspects of human atherosclerotic pathology.

7.4. Hemodynamic control and plaque localization

Atherosclerotic plaques develop preferentially at arterial bifurcations, curvatures, and branch points exposed to disturbed or oscillatory flow. Such hemodynamic conditions promote endothelial activation, lipid retention, oxidative stress, and leukocyte recruitment.^{9,246} Therefore, physiologically relevant atherosclerosis models must incorporate flow. OOC and microfluidic platforms can generate precisely controlled shear-stress profiles that regulate endothelial behavior. Laminar flow typically promotes an atheroprotective phenotype, whereas disturbed flow induces inflammatory activation and barrier dysfunction.^{245,254}

Patient-specific vascular geometries may further improve disease relevance. Medical imaging modalities such as computed tomography angiography, magnetic resonance angiography, intravascular ultrasound, and optical coherence tomography can provide individualized vascular architectures.^{9,254}

When combined with computational fluid dynamics (CFD), these imaging-derived geometries can generate patient-specific shear-stress maps that identify plaque-prone regions and local hemodynamic abnormalities.^{9,254} Such information can subsequently guide fabrication of bioprinted vascular constructs that reproduce clinically relevant disease localization and progression.

7.5. Real-time monitoring and closed-loop disease modeling

Most current vascular disease models rely heavily on endpoint analyses such as immunostaining, gene expression profiling, and biochemical assays. While informative, these approaches provide limited temporal information regarding disease evolution.^{181,255} Integration of biosensing technologies into bioprinted vascular constructs offers opportunities for continuous and non-destructive monitoring of disease progression. Graphene-based materials are particularly attractive because of their electrical conductivity, surface sensitivity, and compatibility with electrochemical sensing systems.^{28,29,121,256}

Real-time monitoring may enable dynamic assessment of endothelial barrier integrity, TEER, permeability, ROS generation, cytokine secretion, metabolic activity, monocyte recruitment, foam-cell formation, and matrix remodeling.⁶⁸ Importantly, future platforms may combine biosensing modules with responsive biomaterials. For example, graphene-based sensors could continuously monitor oxidative and inflammatory signals, while BP-integrated ROS-responsive matrices could dynamically regulate therapeutic release or material behavior in response to disease progression.^{26,74,92} Such integration may ultimately enable closed-loop disease models capable of both monitoring and regulating pathological processes in real time.^{65,255,257-259}

7.6. Patient-specific atherosclerosis models

Atherosclerosis exhibits substantial interpatient heterogeneity arising from differences in genetics, metabolism, vascular anatomy, immune responses, age, and environmental exposures. Consequently, personalized disease models are increasingly needed.⁹

Human induced pluripotent stem cells (hiPSCs) provide a promising source of patient-specific endothelial cells, VSMCs, and macrophage-like cells.^{255,257,260} Incorporation of these cells into bioprinted vascular constructs may enable individualized modeling of endothelial dysfunction, inflammatory responses, lipid metabolism, and therapeutic sensitivity.

In parallel, patient-specific vascular geometries derived from medical imaging can reproduce anatomical features associated with disease susceptibility, including carotid bifurcations, coronary branch points, and restenosis-prone vascular segments.²⁶¹ A comprehensive personalized platform may therefore integrate hiPSC-derived vascular and immune cells, medical imaging-derived vascular geometries, CFD-generated shear-stress distributions, disease-responsive bioinks, and real-time biosensing systems.^{255,257,260-262} Such systems could provide

unprecedented opportunities for investigating patient-specific disease mechanisms and therapeutic responses.

7.7. Current challenges and future directions

Despite significant progress, several challenges continue to limit translation of bioprinted atherosclerosis models. First, biological complexity remains difficult to reproduce. Advanced plaque features—including necrotic core formation, calcification, intraplaque hemorrhage, fibrous cap rupture, and long-term disease evolution—remain challenging to model *in vitro*.²⁶³ Second, bioink standardization remains an important obstacle. Smart bioinks must simultaneously support printability, multiple vascular cell populations, long-term perfusion, and disease-specific responsiveness while maintaining reproducibility and safety.⁸¹ Third, integration of biosensing and responsive biomaterials remains technically challenging. Although graphene-based sensing platforms and BP-containing responsive hydrogels show promise, fully integrated closed-loop disease systems remain largely experimental.^{26,27,29,121} Finally, clinical validation remains limited. Standardized benchmarking criteria and clinically relevant endpoints are required before widespread adoption in drug discovery and precision medicine.^{261,264}

Ultimately, the goal of cardiovascular biofabrication is not simply to fabricate vascular tissues but to construct predictive, patient-specific disease platforms capable of reproducing atherosclerotic progression, therapeutic responses, and clinical heterogeneity. The

convergence of smart bioinks, microfluidics, stem cell engineering, biosensing technologies, medical imaging, and computational modeling is expected to drive the next generation of atherosclerosis-on-a-chip systems for mechanistic investigation, drug screening, and precision cardiovascular medicine.^{257,260,264-266}

8. Challenges and translational perspectives

8.1. Distinguishing bioink from hydrogels: The need for bioprinting validation

A translational checklist for evaluating smart bioinks intended for atherosclerosis modeling is presented in Table 5. Stimuli-responsive hydrogels integrated with 2D nanomaterials have generated considerable interest for cardiovascular bioprinting and atherosclerosis modeling. However, a critical distinction must be made between a hydrogel and a bioink. Many reported systems are injectable hydrogels, wound dressings, or therapeutic matrices that have not been systematically validated for bioprinting applications.¹²⁵

A material should not be considered a functional bioink solely because it exhibits favorable biological activity. Instead, successful bioinks must simultaneously satisfy biological and manufacturing requirements, including cytocompatibility, shear-thinning behavior, viscosity recovery, extrusion fidelity, crosslinking controllability, post-printing shape stability, long-term cell viability, and compatibility with perfusion culture.^{125,198,267}

Table 5. Key translational barriers and validation criteria for nanomaterial-enabled smart bioinks

Challenge	Key question	Recommended evaluation	Current status
Bioink validation ^{15,18,81,125,128,143,176,178,264}	Can it be printed reliably?	Rheology, shear-thinning behavior, shape fidelity	Partially addressed
Material standardization ^{24,268-270}	Can results be reproduced?	Size, oxidation, zeta potential, purity	Poorly standardized
Disease relevance ^{198,267,271}	Does it mimic plaque biology?	VCAM-1, foam cells, ROS, MMPs	Limited validation
Safety ^{81,153,272-275}	Is the material biocompatible?	Inflammation, hemocompatibility	Incomplete
Long-term perfusion ¹⁷⁷⁻¹⁷⁹	Can it tolerate flow?	Perfusion stability, endothelialization	Emerging
Sensor integration ^{121,276-280}	Can it support real-time monitoring?	TEER, ROS, cytokines	Early-stage
Manufacturing ^{24,281-283}	Can it be scaled up?	Batch consistency, sterilization	Major challenge
Regulatory feasibility ^{24,264,267,282-285}	Can it be translated?	GMP compatibility, degradation products	Largely unexplored

Abbreviations: GMP: Good manufacturing practice; MMPs: Matrix metalloproteinases; ROS: Reactive oxygen species; TEER: Transendothelial electrical resistance; VCAM-1: Vascular cell adhesion molecule.1

This distinction is particularly important for nanomaterial-containing systems. Graphene, BP, and germanene can significantly alter hydrogel rheology, network formation, optical behavior, conductivity, degradation kinetics, and cellular responses. Consequently, each nanomaterial-integrated formulation should be evaluated as a complete printing system rather than as a simple hydrogel composite.²⁹

The translational gap between conceptual biomaterial platforms and validated bioprinting systems remains one of the most significant challenges facing the field. Materials that perform well as bulk hydrogels may exhibit poor printability, inadequate structural fidelity, or loss of biological function after printing. Therefore, future studies should prioritize comprehensive biofabrication validation rather than relying solely on hydrogel performance.

8.2. Nanomaterial standardization and reproducibility

A second major challenge is the lack of standardized characterization of 2D nanomaterials. The biological and physicochemical behavior of nanomaterials is highly dependent on parameters such as lateral size, thickness, oxidation state, surface charge, functionalization, aggregation state, purity, degradation behavior, and endotoxin contamination.^{268,269,286}

This issue is particularly relevant for graphene-family materials, BP, and emerging Xenes. GO and rGO exhibit substantial differences in conductivity, surface chemistry, dispersion stability, and biological interactions.^{27,268} BP nanosheets are highly sensitive to oxidation and environmental conditions, which may significantly influence degradation behavior and biological activity.^{287,288} Germanene remains even less standardized because reproducible synthesis, stabilization, and formulation strategies are still under development.²⁶⁹

Without rigorous physicochemical characterization, results cannot be reliably compared across laboratories. Future studies should report material size distribution, layer number, zeta potential, oxidation degree, conductivity, surface chemistry, degradation kinetics, endotoxin levels, and batch-to-batch variability.^{268,270} Standardization of these parameters will be essential for reproducibility, cross-study comparison, and eventual regulatory acceptance.

8.3. Disease relevance and model validation

A major concern in the current literature is that many vascular constructs are evaluated primarily on the basis of cell viability, proliferation, or morphology. While these measurements are important, they do not necessarily demonstrate successful disease modeling. A

bioprinted vascular construct should not automatically be considered an atherosclerosis model. The defining feature of a disease model is its ability to reproduce disease-relevant biological processes rather than merely support vascular cell survival.^{3,9} Current systems often reproduce selected early pathological events such as endothelial inflammation, monocyte adhesion, or oxLDL-induced foam-cell formation. However, advanced features of human atherosclerosis—including necrotic core formation, fibrous cap thinning, calcification, cholesterol crystal deposition, intraplaque hemorrhage, and plaque instability—remain difficult to recapitulate *in vitro*.^{3,248}

Therefore, disease validation should extend beyond conventional cell-based assays. Future models should be evaluated using standardized pathological endpoints. For endothelial dysfunction, these include endothelial permeability, eNOS expression, nitric oxide production, VCAM-1, ICAM-1, E-selectin, and monocyte adhesion.⁹ For VSMC remodeling, α -SMA, SM-MHC, calponin expression, migration, matrix production, and osteogenic differentiation should be assessed.³ For macrophage-driven pathology, relevant endpoints include CD68 expression, macrophage polarization, oxLDL uptake, foam-cell formation, cytokine secretion, ROS production, and matrix-degrading enzyme activity.^{45,248}

Advanced models should additionally incorporate spatial measurements of lipid distribution, ROS gradients, ECM organization, and multicellular interactions within layered vascular constructs.^{257,266} Such validation frameworks are essential for determining whether a model faithfully reproduces atherosclerotic pathology rather than simply representing an inflamed hydrogel.

8.4. Safety, degradation, and immune compatibility

The incorporation of nanomaterials introduces unique safety considerations that extend beyond conventional hydrogel systems. 2D nanomaterials can interact directly with cell membranes, proteins, immune receptors, and intracellular organelles, potentially influencing oxidative stress, inflammatory signaling, mitochondrial function, and cellular homeostasis.^{272,273}

This issue is particularly relevant for atherosclerosis modeling because inflammation and oxidative stress are central disease mechanisms. If a nanomaterial independently induces inflammatory activation, it may confound interpretation of experimental results and obscure disease-specific responses.²⁷²

Graphene-based materials have raised concerns regarding persistence, accumulation, and chronic inflammatory effects.^{269,286} BP offers improved biodegradability, but uncontrolled oxidation may alter

local phosphate concentrations, redox balance, or pH.^{272,273} Germanene remains largely unexplored in cardiovascular systems, and its long-term biodegradation, biodistribution, and immunological consequences remain unclear.

Consequently, future studies should evaluate nanomaterial safety under both baseline and disease-relevant conditions. Cytotoxicity alone is insufficient. Comprehensive assessment should include oxidative stress responses, inflammatory cytokine production, macrophage activation, endothelial barrier integrity, hemocompatibility, thrombogenicity, and long-term degradation behavior.²⁷⁵

8.5. Long-term perfusion and immune-cell compatibility

Physiologically relevant CVD models require long-term perfusion. Static culture systems cannot adequately reproduce shear stress, convective transport, endothelial mechanotransduction, immune-cell recruitment, or hemodynamic regulation.^{79,289,290} However, perfusion introduces substantial engineering challenges. Printed vascular channels must remain patent, resist collapse, maintain endothelialization, and tolerate continuous flow while preserving matrix remodeling and cellular migration.^{81,291,292}

For atherosclerosis models, additional complexity arises from the need to support circulating immune cells, macrophage recruitment, inflammatory signaling, lipid transport, and chronic oxidative stress over extended culture periods. Nanomaterial-containing bioinks must also maintain homogeneous distribution during perfusion while avoiding aggregation, leaching, or loss of functionality.^{142,260}

Importantly, short-term static culture is insufficient for validating atherosclerosis-relevant bioinks. Long-term perfusion studies incorporating immune-cell interactions and hemodynamic stimulation will be essential for assessing physiological relevance and predictive value.^{79,142,289}

8.6. Real-time monitoring and closed-loop disease platforms

Most current disease models rely on endpoint analyses, providing limited information regarding the temporal evolution of disease processes. Because atherosclerosis develops gradually through dynamic interactions among endothelial dysfunction, inflammation, oxidative stress, lipid accumulation, and matrix remodeling, longitudinal monitoring is highly desirable.^{215,293-295} Recent advances in graphene-based sensors, electrochemical interfaces, optical reporters, and microfluidic biosensing technologies provide opportunities for continuous measurement of

barrier integrity, cytokine secretion, ROS production, metabolic activity, and matrix degradation.²⁷⁶⁻²⁸⁰

Looking forward, the integration of biosensing technologies with responsive biomaterials may enable closed-loop disease platforms. For example, a graphene-based sensor could detect inflammatory activation or oxidative stress, while a BP-containing ROS-responsive hydrogel dynamically regulates therapeutic release in response to local disease progression.²⁹⁶⁻²⁹⁸ Such systems would transform bioprinted disease models from passive observation platforms into adaptive experimental systems capable of sensing, interpreting, and responding to pathological signals in real time.^{277,299}

8.7. Regulatory and manufacturing feasibility

Clinical and industrial translation requires more than biological performance. Manufacturing reproducibility, scalability, sterilization, storage stability, quality control, and regulatory classification represent equally important challenges.^{267,281} Nanomaterial-integrated bioinks are particularly complex because variations in synthesis, processing, storage, and degradation may significantly alter material properties and biological responses.²⁸² Regulatory agencies will likely require comprehensive characterization of nanomaterial composition, degradation products, manufacturing workflows, and biological safety profiles before clinical use can be considered.

Sterilization presents another important challenge. Heat, radiation, filtration, or chemical sterilization may alter hydrogel chemistry, nanomaterial structure, conductivity, photothermal properties, or degradation behavior.²⁸³ Consequently, sterilization protocols must be validated individually for each formulation. Although *in vitro* disease models face fewer regulatory barriers than implantable products, reproducibility remains essential for drug screening, interlaboratory comparison, industrial adoption, and future regulatory acceptance.^{267,284}

8.8. Perspective: From material innovation to predictive disease platforms

The major developmental stages and validation requirements for translating nanomaterial-enabled smart bioinks into clinically relevant disease models are summarized in Figure 7. Despite substantial progress in nanomaterial-enabled bioinks, future advances should focus less on material novelty and more on disease-oriented functionality and translational relevance. Historically, bioink development has often been driven by improvements in printability, mechanical properties, conductivity, or photothermal behavior. While these characteristics remain important, the ultimate objective of

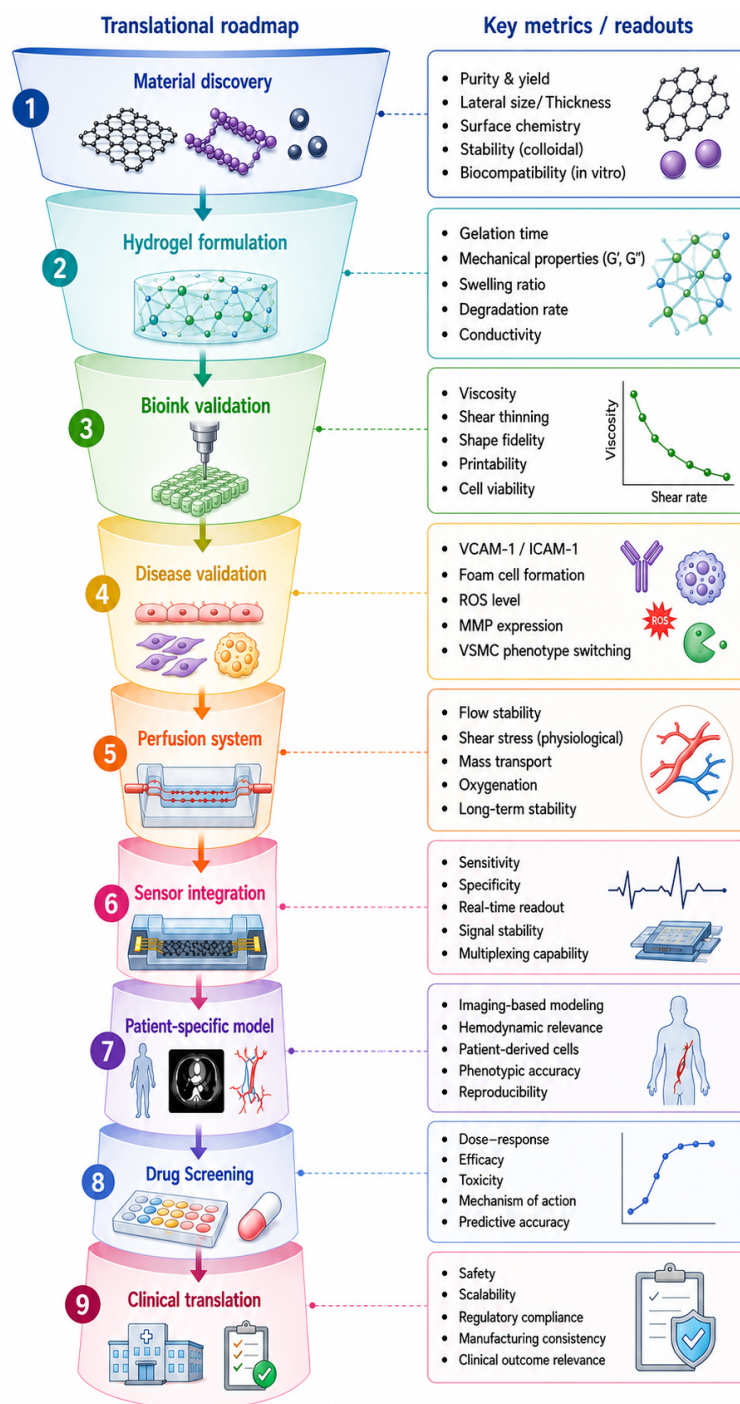


Figure 7. Translational roadmap for nanomaterial-enabled smart bioinks. Conceptual framework illustrating the progression from material development to clinically relevant cardiovascular disease modeling. The translational pathway includes nanomaterial discovery, hydrogel formulation, bioink validation, disease-specific model construction, long-term perfusion culture, biosensing integration, and patient-specific platform development. At each stage, critical validation criteria—including physicochemical characterization, rheological performance, printing fidelity, cellular functionality, disease relevance, immune compatibility, and long-term stability—must be systematically addressed. The roadmap highlights the transition from material-centered innovation toward disease-oriented and validation-driven biofabrication strategies, ultimately enabling predictive atherosclerosis-on-a-chip systems for drug screening, mechanistic investigation, and precision cardiovascular medicine. Created in BioRender. Chu, R. (2026). <https://BioRender.com/h9k8aej>, with assistance from ChatGPT (OpenAI, GPT-5.5 Thinking) for preliminary figure drafting, visual refinement, and layout optimization.

Abbreviations: ICAM-1: Intercellular adhesion molecule 1; MMP: Matrix metalloproteinase; ROS: Reactive oxygen species; VCAM-1: Vascular cell adhesion molecule 1; VSMC: Vascular smooth muscle cell.

cardiovascular biofabrication is to improve the accuracy and predictive value of disease models.^{265,268}

Future bioinks should therefore be designed around specific pathological processes, including endothelial dysfunction, oxidative stress, inflammatory activation, lipid accumulation, vascular remodeling, and immune-cell interactions. Disease-responsive matrices capable of actively sensing and responding to pathological cues may provide more physiologically relevant platforms than passive structural materials alone.^{267,281}

At the same time, increasing biological complexity will be required. Future atherosclerosis models should integrate endothelial cells, VSMCs, macrophages, ECM remodeling, lipoprotein transport, ROS gradients, hemodynamic stimulation, biosensing modules, and patient-specific vascular geometries within unified experimental systems.^{300,301}

The convergence of smart bioinks, OOC technologies, stem cell engineering, biosensing systems, medical imaging, and computational modeling is expected to drive the next generation of CVD platforms. Graphene-based materials may contribute conductivity and biosensing capability, BP may provide disease-responsive redox regulation, and emerging Xenes may offer additional programmable functionalities.³⁰²

Nevertheless, enthusiasm surrounding advanced nanomaterials should be balanced by rigorous validation. The true value of nanomaterial-enabled bioinks will ultimately depend on whether they improve disease-modeling accuracy, predictive power, and translational utility beyond what can be achieved using conventional hydrogel systems. If these challenges can be successfully addressed, smart bioinks integrated with functional nanomaterials may become powerful tools for investigating atherosclerosis pathogenesis, accelerating therapeutic discovery, and enabling patient-specific CVD modeling.

9. Conclusion

Smart hydrogel bioinks integrated with 2D nanomaterials represent a promising strategy for constructing dynamic and physiologically relevant vascular disease models. Unlike conventional hydrogels that primarily provide structural support, smart bioinks can actively regulate cellular behavior, matrix remodeling, oxidative stress, and disease-associated signaling pathways. By combining responsive hydrogel chemistry with advanced biofabrication technologies, these materials offer new opportunities to reconstruct key features of atherosclerosis, including endothelial dysfunction, inflammation, ECM remodeling, and disturbed-flow-mediated vascular pathology.

Among currently available nanomaterials, graphene-family materials remain the most mature additives for cardiovascular bioinks, providing mechanical reinforcement, electrical conductivity, and biosensing functionality. In contrast, BP exhibits a more disease-matched profile owing to its biodegradability, ROS responsiveness, and redox-regulatory behavior, making it particularly attractive for atherosclerosis-oriented bioink design. Emerging Xenes such as germanene remain at an exploratory stage, with limited evidence supporting cardiovascular bioprinting applications. Consequently, graphene should be viewed primarily as a structural, conductive, and sensing component, whereas BP represents a promising disease-responsive module that still requires rigorous bioprinting and vascular-model validation.

Looking forward, future progress will depend less on the development of increasingly complex materials and more on the establishment of biologically relevant and rigorously validated disease platforms. Key priorities include standardized nanomaterial characterization, validation of bioprinting performance, integration of multicellular and perfusable vascular systems, incorporation of real-time sensing technologies, and benchmarking against human plaque biology. The convergence of smart bioinks, OOC systems, patient-derived vascular cells, biosensing technologies, and computational modeling may ultimately enable predictive and patient-specific platforms for mechanistic investigation, therapeutic screening, and precision cardiovascular medicine.

Acknowledgments

The authors would like to thank Professor Xiangrong Song for providing valuable suggestions and support regarding the conceptual framework and overall structure of this manuscript. This non-funding support helped improve the organization and clarity of the review. The authors also appreciate the constructive input from colleagues who contributed helpful discussions during manuscript preparation.

Funding

This study was supported by a key research project grant from the Chengdu Technology Bureau (2026-YF05-00796-SN) and the Sichuan Province Science and Technology Major Special Project (2025ZDZX0039).

Conflict of interest

The authors declare they have no competing interests.

Author contributions

Conceptualization: Shengbin Liu, Jiangzhou Chu, Zhongshan He

Visualization: Zhi Liu

Writing–original draft: Jiangzhou Chu, Shengbin Liu

Writing–review & editing: Shengbin Liu, Zhongshan He

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of data

Not applicable.

Further disclosure

ChatGPT (OpenAI, GPT-5.5 Thinking) was used during manuscript revision solely for language polishing, formatting assistance, and literature-related support. The authors directed all prompts, critically reviewed and edited all AI-assisted outputs, verified the scientific accuracy of the manuscript, and take full responsibility for the final content. BioRender.com and ChatGPT.55 image-generation assistance were used for preliminary figure drafting, visual refinement, and layout optimization. All figure concepts, annotations, labels, and final visual content were reviewed, revised, and approved by the authors. No third-party copyrighted images were reused.

References

- Porsch F, Binder CJ. Autoimmune diseases and atherosclerotic cardiovascular disease. *Nat Rev Cardiol.* 2024;21(11):780–807.
doi: 10.1038/s41569-024-01045-7
- Shoaran M, Maffia P. Tackling inflammation in atherosclerosis. *Nat Rev Cardiol.* 2024;21(7):442.
doi: 10.1038/s41569-024-01007-z
- Libby P, Buring JE, Badimon L, *et al.* Atherosclerosis. *Nat Rev Dis Prim.* 2019;5(1):56.
doi: 10.1038/s41572-019-0106-z
- Mach F, Baigent C, Catapano AL, *et al.* 2019 ESC/EAS guidelines for the management of dyslipidaemias: Lipid modification to reduce cardiovascular risk. *Atherosclerosis.* 2019;290(1):140–205.
doi: 10.1016/j.atherosclerosis.2019.08.014
- Zhang L, Zhou J, Kong W. Extracellular matrix in vascular homeostasis and disease. *Nat Rev Cardiol.* 2025;22(5):333–353.
doi: 10.1038/S41569-024-01103-0
- Lin A, Miano JM, Fisher EA, Misra A. Chronic inflammation and vascular cell plasticity in atherosclerosis. *Nat Cardiovasc Res.* 2024;3(12):1408–1423.
doi: 10.1038/S44161-024-00569-Y
- Döring Y, van der Vorst EPC, Weber C. Targeting immune cell recruitment in atherosclerosis. *Nat Rev Cardiol.* 2024;21(11):824–840.
doi: 10.1038/S41569-024-01023-Z
- Stroope C, Nettersheim FS, Coon B, *et al.* Dysregulated cellular metabolism in atherosclerosis: mediators and therapeutic opportunities. *Nat Metab.* 2024;6(4):617.
doi: 10.1038/S42255-024-01015-W
- Tamargo IA, Baek KI, Kim Y, Park C, Jo H. Flow-induced reprogramming of endothelial cells in atherosclerosis. *Nat Rev Cardiol.* 2023;20(11):738–753.
doi: 10.1038/S41569-023-00883-1
- Nam U, Lee S, Ahmad A, Yi HG, Jeon JS. Microphysiological Systems as Organ-Specific In Vitro Vascular Models for Disease Modeling. *BioChip J.* 2024;18(3):345–356.
doi: 10.1007/S13206-024-00152-4
- Zhao Y, Landau S, Okhovatian S, *et al.* Integrating organoids and organ-on-a-chip devices. *Nat Rev Bioeng.* 2024;2(7):588–608.
doi: 10.1038/s44222-024-00207-z
- Khosravi R, Radisic M. Heart-on-a-chip and vasculature-on-a-chip platforms as models of cardiovascular disease. *Nat Rev Cardiol.* 2026;23(8):537–554.
doi: 10.1038/s41569-026-01255-1
- Huang MS, Christakopoulos F, Roth JG, Heilshorn SC. Organoid bioprinting: from cells to functional tissues. *Nat Rev Bioeng.* 2025;3(2):126–142.
doi: 10.1038/s44222-024-00268-0
- Khanna A, Ayan B, Undieh AA, Yang YP, Huang NF. Advances in three-dimensional bioprinted stem cell-based tissue engineering for cardiovascular regeneration. *J Mol Cell Cardiol.* 2022;169:13–27.
doi: 10.1016/j.yjmcc.2022.04.017
- Kim MS, Choi Y, Lee KY. Three-Dimensional Printing and Bioprinting Strategies for Cardiovascular Constructs: From Printing Inks to Vascularization. *Polym.* 2025;17(17):2337.
doi: 10.3390/POLYM17172337
- Wang B, Zhang J, Jiang J, Cheng C, Zhao Y. Current trends, applications, and challenges in three-dimensional bioprinting for cardiovascular disease models and therapies. *iScience.* 2026;29(5):115519.
doi: 10.1016/j.isci.2026.115519
- Ma D, Liu J, Lu WW, Liu W, Ruan C. Dynamic bioinks for tissue/organ bioprinting: Principle, challenge, and perspective. *Prog Mater Sci.* 2026;155:101527.

- doi: 10.1016/j.pmatsci.2025.101527
18. Chandarana C, Sane D, Mishra S, Chaubey A, Gohil U, Prajapati B. Recent Advances in Bioink Research for Biomedical Applications. *Biomed Mater Devices*. 2026;4(4):3950–3981.
doi: 10.1007/S44174-025-00478-Z
19. Malekmohammadi S, Aminabad NS, Sabzi A, *et al*. Smart and Biomimetic 3D and 4D Printed Composite Hydrogels: Opportunities for Different Biomedical Applications. *Biomedicines*. 2021;9(11):1–46.
doi: 10.3390/biomedicines9111537
20. El-Husseiny HM, Mady EA, Hamabe L, *et al*. Smart/stimuli-responsive hydrogels: Cutting-edge platforms for tissue engineering and other biomedical applications. *Mater Today Bio*. 2022;13:100186.
doi: 10.1016/j.mtbio.2021.100186
21. Rahimnejad M, Jahangiri S, Zirak Hassan Kiadeh S, *et al*. Stimuli-responsive biomaterials: smart avenue toward 4D bioprinting. *Crit Rev Biotechnol*. 2024;44(5):860–891.
doi: 10.1080/07388551.2023.2213398
22. Kim J, D A G, Debnath P, Saha P. Smart Multi-Responsive Biomaterials and Their Applications for 4D Bioprinting. *Biomimetics*. 2024;9(8):484.
doi: 10.3390/biomimetics9080484
23. Yarali E, Mirzaali MJ, Ghalayanesfahani A, Accardo A, Diaz-Payno PJ, Zadpoor AA. 4D Printing for Biomedical Applications. *Adv Mater*. 2024;36(31):e2402301.
doi: 10.1002/adma.202402301
24. Kandasamy M, Vijayananth K, Parasuraman A, Ayrilmis N. 3D Bioprinting of Biomaterials: A Review of Advances in Techniques, Materials, and Applications. *Polym Adv Technol*. 2025;36(9):1–17.
doi: 10.1002/pat.70324
25. Batty M, Bennett MR, Yu E. The Role of Oxidative Stress in Atherosclerosis. *Cells*. 2022;11(23):3843.
doi: 10.3390/cells11233843
26. Pu M, Cao H, Zhang H, *et al*. ROS-responsive hydrogels: from design and additive manufacturing to biomedical applications. *Mater Horizons*. 2024;11(16):3721–3746.
doi: 10.1039/D4MH00289J
27. Gaihre B, Potes MA, Serdiuk V, Tilton M, Liu X, Lu L. Two-dimensional nanomaterials-added dynamism in 3D printing and bioprinting of biomedical platforms: Unique opportunities and challenges. *Biomaterials*. 2022;284:121507.
doi: 10.1016/j.biomaterials.2022.121507
28. Zhang J, Eyisoğlu H, Qin XH, Rubert M, Müller R. 3D bioprinting of graphene oxide-incorporated cell-laden bone mimicking scaffolds for promoting scaffold fidelity, osteogenic differentiation and mineralization. *Acta Biomater*. 2021;121:637–652.
doi: 10.1016/j.actbio.2020.12.026
29. Patil R, Alimperti S. Graphene in 3D Bioprinting. *J Funct Biomater*. 2024;15(4):82.
doi: 10.3390/jfb15040082
30. Edrisi F, Baheiraei N, Razavi M, Roshanbinfar K, Imani R, Jalilnejad N. Potential of graphene-based nanomaterials for cardiac tissue engineering. *J Mater Chem B*. 2023;11(31):7280–7299.
doi: 10.1039/D3TB00654A
31. Memarian P, Bagher Z, Asghari S, Aleemardani M, Seifalian A. Emergence of graphene as a novel nanomaterial for cardiovascular applications. *Nanoscale*. 2024;16(27):12793–12819.
doi: 10.1039/d4nr00018h
32. He Z, Chen W, Hu K, *et al*. Resolvin D1 delivery to lesional macrophages using antioxidative black phosphorus nanosheets for atherosclerosis treatment. *Nat Nanotechnol*. 2024;19(9):1386–1398.
doi: 10.1038/S41565-024-01687-1
33. He M, Zhang X, Ran X, *et al*. Black Phosphorus Nanosheets Protect Neurons by Degrading Aggregative α -syn and Clearing ROS in Parkinson's Disease. *Adv Mater*. 2024;36(30):e2404576.
doi: 10.1002/adma.202404576
34. Huang W, Hu L, Tang Y, Xie Z, Zhang H. Recent Advances in Functional 2D MXene-Based Nanostructures for Next-Generation Devices. *Adv Funct Mater*. 2020;30(49):2005223.
doi: 10.1002/adfm.202005223
35. Wang M, Huang W. Emerging Xene-Related Nanostructures for Versatile Applications. *Nanomaterials*. 2023;13(3):517.
doi: 10.3390/nano13030517
36. Wang X, Sun X, Bu T, *et al*. Germanene-modified chitosan hydrogel for treating bacterial wound infection: An ingenious hydrogel-assisted photothermal therapy strategy. *Int J Biol Macromol*. 2022;221:1558–1571.
doi: 10.1016/j.ijbiomac.2022.09.128
37. Kang MS, Jang HJ, Jo HJ, Raja IS, Han DW. MXene and Xene: promising frontier beyond graphene in tissue engineering and regenerative medicine. *Nanoscale Horizons*. 2024;9(1):93–117.
doi: 10.1039/D3NH00428G
38. Bennett MR, Sinha S, Owens GK. Vascular Smooth Muscle Cells in Atherosclerosis. *Circ Res*. 2016;118(4):692–702.
doi: 10.1161/CIRCRESAHA.115.306361

39. Lambert J, Jørgensen HF. Epigenetic regulation of vascular smooth muscle cell phenotypes in atherosclerosis. *Atherosclerosis*. 2025;401:119085.
doi: 10.1016/j.atherosclerosis.2024.119085
40. Grootaert MOJ, Bennett MR. Vascular smooth muscle cells in atherosclerosis: time for a re-assessment. *Cardiovasc Res*. 2021;117(11):2326-2339.
doi: 10.1093/CVR/CVAB046
41. Chen R, McVey DG, Shen D, Huang X, Ye S. Phenotypic Switching of Vascular Smooth Muscle Cells in Atherosclerosis. *J Am Heart Assoc*. 2023;12(20):31121.
doi: 10.1161/JAHA.123.031121
42. Saraswathibhatla A, Indana D, Chaudhuri O. Cell-extracellular matrix mechanotransduction in 3D. *Nat Rev Mol Cell Biol*. 2023;24(7):495-516.
doi: 10.1038/S41580-023-00583-1
43. Kong P, Cui ZY, Huang XF, Zhang DD, Guo RJ, Han M. Inflammation and atherosclerosis: signaling pathways and therapeutic intervention. *Signal Transduct Target Ther*. 2022;7(1):131.
doi: 10.1038/S41392-022-00955-7
44. Ajoolabady A, Pratico D, Lin L, et al. Inflammation in atherosclerosis: pathophysiology and mechanisms. *Cell Death Dis*. 2024;15(11):817.
doi: 10.1038/S41419-024-07166-8
45. Hou P, Fang J, Liu Z, et al. Macrophage polarization and metabolism in atherosclerosis. *Cell Death Dis*. 2023;14(10):691.
doi: 10.1038/S41419-023-06206-Z
46. Chen R, Zhang H, Tang B, et al. Macrophages in cardiovascular diseases: molecular mechanisms and therapeutic targets. *Signal Transduct Target Ther*. 2024;9(1):130.
doi: 10.1038/S41392-024-01840-1
47. Wieland EB, Kempen LJAP, Donners MMPC, Biessen EAL, Goossens P. Macrophage heterogeneity in atherosclerosis: A matter of context. *Eur J Immunol*. 2024;54(1):e2350464.
doi: 10.1002/EJI.202350464
48. Li Q, Yu S, Wang Y, et al. Multiscale bioprinted arterial models recapitulate synergistic microenvironmental interactions in vascular disease. *Cell Biomater*. 2026;2(5):100257.
doi: 10.1016/j.celbio.2025.100257
49. Förstermann U, Xia N, Li H. Roles of Vascular Oxidative Stress and Nitric Oxide in the Pathogenesis of Atherosclerosis. *Circ Res*. 2017;120(4):713-735.
doi: 10.1161/circresaha.116.309326
50. Susser LI, Rayner KJ. Through the layers: how macrophages drive atherosclerosis across the vessel wall. *J Clin Invest*. 2022;132(9):e157011.
doi: 10.1172/jci157011
51. Gastanadui MG, Margaroli C, Litovsky S, et al. Spatial Transcriptomic Approach to Understanding Coronary Atherosclerotic Plaque Stability. *Arterioscler Thromb Vasc Biol*. 2024;44(11):e264-e276.
doi: 10.1161/atvbaha.123.320330
52. Cole JE, Monaco C. Spatial Transcriptomics: A New Frontier in Atherosclerosis Research? *Arterioscler Thromb Vasc Biol*. 2024;44(11):2291-2293.
doi: 10.1161/atvbaha.124.321652
53. Yu H, Gao R, Liu Y, Fu L, Zhou J, Li L. Stimulus-Responsive Hydrogels as Drug Delivery Systems for Inflammation Targeted Therapy. *Adv Sci*. 2024;11(1):1-48.
doi: 10.1002/advs.202306152
54. Zhang J, Sun D, Liao Y, et al. Time-Released Black Phosphorus Hydrogel Accelerates Myocardial Repairing through Antioxidant and Motivates Macrophage Polarization Properties. *Biomater Res*. 2024;28:0029.
doi: 10.34133/BMR.0029
55. Zhang J, Sun D, Guo Y, et al. Targeted delivery of black phosphorus nanosheets by ROS responsive complex hydrogel based on angiogenesis and antioxidant promotes myocardial infarction repair. *J Nanobiotechnology*. 2024;22(1):433.
doi: 10.1186/s12951-024-02685-0
56. Di Nubila A, Dilella G, Simone R, Barbieri SS. Vascular Extracellular Matrix in Atherosclerosis. *Int J Mol Sci*. 2024;25(22):12017.
doi: 10.3390/ijms252212017
57. Alonso-Herranz L, Albarrán-Juárez J, Bentzon JF. Mechanisms of fibrous cap formation in atherosclerosis. *Front Cardiovasc Med*. 2023;10:1-7.
doi: 10.3389/fcvm.2023.1254114
58. Yan A, Gotlieb AI. The microenvironment of the atheroma expresses phenotypes of plaque instability. *Cardiovasc Pathol*. 2023;67:107572.
doi: 10.1016/j.carpath.2023.107572
59. Ruddy JM, Ikonomidis JS, Jones JA. Multidimensional Contribution of Matrix Metalloproteinases to Atherosclerotic Plaque Vulnerability: Multiple Mechanisms of Inhibition to Promote Stability. *J Vasc Res*. 2016;53(1-2):1-16.
doi: 10.1159/000446703
60. Wang X, Shen Y, Shang M, Liu X, Munn LL. Endothelial mechanobiology in atherosclerosis. *Cardiovasc Res*. 2023;119(8):1656-1675.
doi: 10.1093/cvr/cvad076

61. Cheng H, Zhong W, Wang L, *et al.* Effects of shear stress on vascular endothelial functions in atherosclerosis and potential therapeutic approaches. *Biomed Pharmacother.* 2023;158:114198.
doi: 10.1016/j.biopha.2022.114198
62. Wang WL, Shih YT, Wei SY, Chiu JJ. Impacts of aging and fluid shear stress on vascular endothelial metabolism and atherosclerosis development. *J Biomed Sci.* 2025;32(1):83.
doi: 10.1186/s12929-025-01177-z
63. He L, Zhang CL, Chen Q, Wang L, Huang Y. Endothelial shear stress signal transduction and atherogenesis: From mechanisms to therapeutics. *Pharmacol Ther.* 2022;235:108152.
doi: 10.1016/j.pharmthera.2022.108152
64. Liu Y, Huang T, Yap NA, Lim K, Ju LA. Harnessing the power of bioprinting for the development of next-generation models of thrombosis. *Bioact Mater.* 2024;42:328-344.
doi: 10.1016/j.bioactmat.2024.08.040
65. Maringanti R, Van Dijk CGM, Meijer EM, *et al.* Atherosclerosis on a Chip: A 3-Dimensional Microfluidic Model of Early Arterial Events in Human Plaques. *Arterioscler Thromb Vasc Biol.* 2024;44(12):2453-2472.
doi: 10.1161/atvbaha.124.321332
66. Gong S, Li Y, Yan K, *et al.* The Crosstalk Between Endothelial Cells, Smooth Muscle Cells, and Macrophages in Atherosclerosis. *Int J Mol Sci.* 2025;26(4):1457.
doi: 10.3390/ijms26041457
67. Vuong TNAM, Bartolf-Kopp M, Anđelović K, *et al.* Integrating Computational and Biological Hemodynamic Approaches to Improve Modeling of Atherosclerotic Arteries. *Adv Sci (Weinh).* 2024;11(26):e2307627.
doi: 10.1002/advs.202307627
68. Kim JJ, Cho DW. Advanced strategies in 3D bioprinting for vascular tissue engineering and disease modelling using smart bioinks. *Virtual Phys Prototyp.* 2024;19(1):e2395470.
doi: 10.1080/17452759.2024.2395470
69. Lu P, Ruan D, Huang M, *et al.* Harnessing the potential of hydrogels for advanced therapeutic applications: current achievements and future directions. *Signal Transduct Target Ther.* 2024;9(1):166.
doi: 10.1038/S41392-024-01852-X
70. Neumann M, di Marco G, Iudin D, *et al.* Stimuli-Responsive Hydrogels: The Dynamic Smart Biomaterials of Tomorrow. *Macromolecules.* 2023;56(21):8377-8392.
doi: 10.1021/acs.macromol.3C00967
71. Ikram M, Mahmud MAP, Kalyar AA, Alomayri T, Almahri A, Hussain D. 3D-bioprinting of MXenes: Developments, medical applications, challenges, and future roadmap. *Colloids Surfaces B Biointerfaces.* 2025;251:114568.
doi: 10.1016/j.colsurfb.2025.114568
72. Choi JH, Haizan I, Choi JW. Recent advances in two-dimensional materials for the diagnosis and treatment of neurodegenerative diseases. *Discov Nano.* 2024;19(1):151.
doi: 10.1186/S11671-024-04099-1
73. Cao H, Duan L, Zhang Y, Cao J, Zhang K. Current hydrogel advances in physicochemical and biological response-driven biomedical application diversity. *Signal Transduct Target Ther.* 2021;6(1):426.
doi: 10.1038/S41392-021-00830-X
74. Miao Y, Chen Y, Luo J, *et al.* Black phosphorus nanosheets-enabled DNA hydrogel integrating 3D-printed scaffold for promoting vascularized bone regeneration. *Bioact Mater.* 2023;21:97-109.
doi: 10.1016/j.bioactmat.2022.08.005
75. Wolf D, Ley K. Immunity and Inflammation in Atherosclerosis. *Circ Res.* 2019;124(2):315-327.
doi: 10.1161/CIRCRESAHA.118.313591
76. Su C, Menon NV, Xu X, *et al.* A novel human arterial wall-on-a-chip to study endothelial inflammation and vascular smooth muscle cell migration in early atherosclerosis. *Lab Chip.* 2021;21(12):2359-2371.
doi: 10.1039/D1LC00131K
77. Kort-Mascort J, Flores-Torres S, Peza-Chavez O, *et al.* Decellularized ECM hydrogels: prior use considerations, applications, and opportunities in tissue engineering and biofabrication. *Biomater Sci.* 2023;11(2):400-431.
doi: 10.1039/D2BM01273A
78. Wang Z, Hu C, Zhang W, *et al.* Dynamically crosslinked ECM-like hydrogels loaded with ROS-responsive drug nanoparticles for treating inflammation in myocardial infarction and stroke. *Compos Part B Eng.* 2024;285:111734.
doi: 10.1016/j.compositesb.2024.111734
79. Shakeri A, Wang Y, Zhao Y, *et al.* Engineering Organ-on-a-Chip Systems for Vascular Diseases. *Arterioscler Thromb Vasc Biol.* 2023;43(12):2241-2255.
doi: 10.1161/atvbaha.123.318233
80. Basatemur GL, Jørgensen HF, Clarke MCH, Bennett MR, Mallat Z. Vascular smooth muscle cells in atherosclerosis. *Nat Rev Cardiol.* 2019;16(12):727-744.
doi: 10.1038/s41569-019-0227-9
81. Synofzik J, Heene S, Jonczyk R, Blume C. Ink-structuring the future of vascular tissue engineering: a review of the physiological bioink design. *Bio-Design Manuf.* 2024;7(2):181-205.
doi: 10.1007/S42242-024-00270-W

82. Zhou Z, Tang W, Yang J, Fan C. Application of 4D printing and bioprinting in cardiovascular tissue engineering. *Biomater Sci.* 2023;11(19):6403-6420.
doi: 10.1039/D3BM00312D
83. Motta I, Soccio M, Guidotti G, Lotti N, Pasquinelli G. Hydrogels for Cardio and Vascular Tissue Repair and Regeneration. *Gels (Basel, Switzerland).* 2024;10(3):196.
doi: 10.3390/GELS10030196
84. El-Husseiny HM, Mady EA, El-Dakrouy WA, Doghish AS, Tanaka R. Stimuli-responsive hydrogels: smart state-of-the-art platforms for cardiac tissue engineering. *Front Bioeng Biotechnol.* 2023;11(Jun 28):1-30.
doi: 10.3389/fbioe.2023.1174075
85. Kalhori D, Zakeri N, Zafar-Jafarzadeh M, Moroni L, Solati-Hashjin M. Cardiovascular 3D bioprinting: A review on cardiac tissue development. *Bioprinting.* 2022;28(2022):e00221.
doi: 10.1016/j.bprint.2022.e00221
86. Schwab A, Levato R, D'Este M, Piluso S, Eglin D, Malda J. Printability and Shape Fidelity of Bioinks in 3D Bioprinting. *Chem Rev.* 2020;120(19):11028-11055.
doi: 10.1021/acs.chemrev.0C00084
87. Kamboj D, Choudhary B, Singh G, Kumar N, Narwal S, Dhingra AK. Oxidative Stress in Cardiovascular Diseases: Mechanisms and Exploring Advanced Therapies. *Cardiovasc Hematol Agents Med Chem.* 2025;23(3):183-201.
doi: 10.2174/0118715257344485250207074727
88. Ungvari Z, Tarantini S, Donato AJ, Galvan V, Csiszar A. Mechanisms of vascular aging. *Circ Res.* 2018;123(7):849-867.
doi: 10.1161/CIRCRESAHA.118.31137
89. Ungvari Z, Tarantini S, Sorond F, Merkely B, Csiszar A. Mechanisms of Vascular Aging, A Geroscience Perspective: JACC Focus Seminar. *J Am Coll Cardiol.* 2020;75(8):931.
doi: 10.1016/j.jacc.2019.11.061
90. Yuan Z, Li Y, Sun M, *et al.* Recent progress in ROS-responsive biomaterials for the diagnosis and treatment of cardiovascular diseases. *Theranostics.* 2025;15(11):5172-5219.
doi: 10.7150/thno.106991
91. Tapeinos C, Gao H, Bauleth-Ramos T, Santos HA. Progress in Stimuli-Responsive Biomaterials for Treating Cardiovascular and Cerebrovascular Diseases. *Small.* 2022;18(36):e2200291.
doi: 10.1002/sml.202200291
92. Zhang Y, Jiang M, Wang T. Reactive oxygen species (ROS)-responsive biomaterials for treating myocardial ischemia-reperfusion injury. *Front Bioeng Biotechnol.* 2024;12:1469393.
doi: 10.3389/fbioe.2024.1469393
93. Shen Z, Guo Z, Tan T, Hu J, Zhang Y. Reactive Oxygen Species Scavenging and Biodegradable Peptide Hydrogel as 3D Culture Scaffold for Cardiomyocytes. *ACS Biomater Sci Eng.* 2020;6(7):3957-3966.
doi: 10.1021/acsbiomaterials.0c00340
94. Tong S, Chen J, Li Y, Zhao W. Emerging Gel Technologies for Atherosclerosis Research and Intervention. *Gels.* 2026;12(1):80.
doi: 10.3390/GELS12010080
95. Fu Y, Zhou Y, Wang K, Li Z, Kong W. Extracellular Matrix Interactome in Modulating Vascular Homeostasis and Remodeling. *Circ Res.* 2024;134(7):931-949.
doi: 10.1161/CIRCRESAHA.123.324055
96. Bräuninger H, Krüger S, Bacmeister L, *et al.* Matrix metalloproteinases in coronary artery disease and myocardial infarction. *Basic Res Cardiol.* 2023;118(1):18.
doi: 10.1007/S00395-023-00987-2
97. Chen W, Wang C, Liu W, *et al.* A Matrix-Metalloproteinase-Responsive Hydrogel System for Modulating the Immune Microenvironment in Myocardial Infarction. *Adv Mater.* 2023;35(13):2209041.
doi: 10.1002/adma.202209041
98. Pi Y, Ganabady K, Celiz AD. Enzyme-responsive biomaterials for biomedical applications. *Commun Mater.* 2025;6(1):263.
doi: 10.1038/s43246-025-00983-0
99. Zustiak SP, Leach JB. Hydrolytically degradable poly(ethylene glycol) hydrogel scaffolds with tunable degradation and mechanical properties. *Biomacromolecules.* 2010;11(5):1348-1357.
doi: 10.1021/bm100137q
100. Marder M, Remmert C, Perschel JA, *et al.* Stem cell-derived vessels-on-chip for cardiovascular disease modeling. *Cell Rep.* 2024;43(4):114008.
doi: 10.1016/j.celrep.2024.114008
101. Ajiteru O, Sultan MT, Lee YJ, *et al.* A 3D Printable Electroconductive Biocomposite Bioink Based on Silk Fibroin-Conjugated Graphene Oxide. *Nano Lett.* 2020;20(9):6873-6883.
doi: 10.1021/acs.nanolett.0c02986
102. Lee M, Park J, Choe G, *et al.* A Conductive and Adhesive Hydrogel Composed of MXene Nanoflakes as a Printable Cardiac Patch for Infarcted Heart Repair. *ACS Nano.* 2023;17(13):12290-12304.
doi: 10.1021/acsnano.3c00933
103. Chong J, Sung C, Nam KS, *et al.* Highly conductive tissue-

- like hydrogel interface through template-directed assembly. *Nat Commun.* 2023;14(1):2206.
doi: 10.1038/S41467-023-37948-1
104. Grzelak KA, Westerfield AD, Kumar V, *et al.* Electrical stimulation directs formation of perfused vasculature in engineered tissues. *bioRxiv Prepr Serv Biol.* Published online September 3, 2025.
doi: 10.1101/2025.08.28.672965
105. Qian Y, Yao Y. Mechanobiological regulation of endothelial vascularization after myocardial infarction: matrix mechanics, hydrogel strategies and applications. *Biomater Sci.* Published online. 2026
doi: 10.1039/D5BM01885D
106. Tao Y, Chan HF, Shi B, Li M, Leong KW. Light: A Magical Tool for Controlled Drug Delivery. *Adv Funct Mater.* 2020;30(49):2005029.
doi: 10.1002/adfm.202005029
107. Luo R, Xiang X, Jiao Q, Hua H, Chen Y. Photoresponsive Hydrogels for Tissue Engineering. *ACS Biomater Sci Eng.* 2024;10(6):3612-3630.
doi: 10.1021/acsbomaterials.4c00314
108. Zhang Y, Kumar P, Lv S, *et al.* Recent advances in 3D bioprinting of vascularized tissues. *Mater Des.* 2021;199:109398.
doi: 10.1016/j.matdes.2020.109398
109. Xian R, Xian H, Dong H, *et al.* Black Phosphorus-Loaded Gelatin Methacryloyl Hydrogels Enhance Angiogenesis via Activation of the PEAK1-MAPK Pathway. *ACS Appl Mater Interfaces.* 2025;17(18):26371-26385.
doi: 10.1021/acsam.5C02054
110. Zhang Y, Qi G, Zhang L, *et al.* Black phosphorus-based photothermal-responsive hydrogel enhanced osteoporotic bone injury regeneration by alleviating oxidative stress and remodeling bone homeostasis. *J Nanobiotechnology.* 2026;24(1):238.
doi: 10.1186/s12951-026-04097-8
111. Picos A, Seoane N, Campos-Toimil M, Viña D. Vascular senescence and aging: mechanisms, clinical implications, and therapeutic prospects. *Biogerontology.* 2025;26(3):118.
doi: 10.1007/S10522-025-10256-5
112. Herzog MJ, Müller P, Lechner K, *et al.* Arterial stiffness and vascular aging: mechanisms, prevention, and therapy. *Signal Transduct Target Ther.* 2025;10(1):282.
doi: 10.1038/s41392-025-02346-0
113. Zheng Y, Hong X, Wang J, *et al.* 2D Nanomaterials for Tissue Engineering and Regenerative Nanomedicines: Recent Advances and Future Challenges. *Adv Healthc Mater.* 2021;10(7):e2001743.
doi: 10.1002/adhm.202001743
114. Zorrón M, Cabrera AL, Sharma R, *et al.* Emerging 2D Nanomaterials-Integrated Hydrogels: Advancements in Designing Theragenerative Materials for Bone Regeneration and Disease Therapy. *Adv Sci.* 2024;11(31):e2403204-e2403204.
doi: 10.1002/advs.202403204
115. Qi W, Zhang R, Wang Z, *et al.* Advances in the Application of Black Phosphorus-Based Composite Biomedical Materials in the Field of Tissue Engineering. *Pharmaceuticals (Basel).* 2024;17(2):242.
doi: 10.3390/ph17020242
116. Lee H, Kim KS, Zare I, *et al.* Smart nanomaterials for multimodal theranostics and tissue regeneration. *Coord Chem Rev.* 2025;541:216801.
doi: 10.1016/j.ccr.2025.216801
117. Chen J, Deng M, Wang J, *et al.* Recent advances in injectable hydrogels for osteoarthritis treatments. *Front Bioeng Biotechnol.* 2025;13:1644222.
doi: 10.3389/fbioe.2025.1644222
118. Jalilnejad N, Rabiee M, Baheiraei N, *et al.* Electrically conductive carbon-based (bio)-nanomaterials for cardiac tissue engineering. *Bioeng Transl Med.* 2023;8(1):e10347.
doi: 10.1002/btm2.10347
119. Lee M, Kim MC, Lee JY. Nanomaterial-Based Electrically Conductive Hydrogels for Cardiac Tissue Repair. *Int J Nanomedicine.* 2022;17:6181-6200.
doi: 10.2147/ijn.s386763
120. Lisboa ES, Serafim C, Santana W, *et al.* Nanomaterials-combined methacrylated gelatin hydrogels (GelMA) for cardiac tissue constructs. *J Control Release.* 2024;365:617-639.
doi: 10.1016/j.jconrel.2023.11.056
121. Byrne R, Carrico A, Lettieri M, Rajan AK, Forster RJ, Cumba LR. Bioinks and biofabrication techniques for biosensors development: A review. *Mater today Bio.* 2024;28:101185.
doi: 10.1016/j.mtbio.2024.101185
122. Wang Z, Zhao J, Tang W, *et al.* Multifunctional Nanoengineered Hydrogels Consisting of Black Phosphorus Nanosheets Upregulate Bone Formation. *Small.* 2019;15(41):e1901560.
doi: 10.1002/smll.201901560
123. Li J, Liu X, Crook JM, Wallace GG. Development of 3D printable graphene oxide based bio-ink for cell support and tissue engineering. *Front Bioeng Biotechnol.* 2022;10:994776.
doi: 10.3389/fbioe.2022.994776
124. Shin SR, Zihlmann C, Akbari M, *et al.* Reduced Graphene Oxide-GelMA Hybrid Hydrogels as Scaffolds for Cardiac

- Tissue Engineering. *Small*. 2016;12(27):3677-3689.
doi: 10.1002/sml.201600178
125. Chen XB, Fazel Anvari-Yazdi A, Duan X, *et al*. Biomaterials / bioinks and extrusion bioprinting. *Bioact Mater*. 2023;28:511-536.
doi: 10.1016/j.bioactmat.2023.06.006
126. Santana MDV, Magulas MBS, Brito GC, *et al*. Cryogenic 3D Printing of GelMA/Graphene Bioinks: Improved Mechanical Strength and Structural Properties for Tissue Engineering. *Int J Nanomedicine*. 2024;19:10745-10765.
doi: 10.2147/ijn.s486868
127. Ni F, Chen Y, Wang Z, *et al*. Graphene derivative based hydrogels in biomedical applications. *J Tissue Eng*. 2024;15:20417314241282131.
doi: 10.1177/20417314241282131
128. Diez-Aldama I, Garcia-Villen F, Saenz-Del-Burgo L, Scaini D, Pedraz JL. Graphene Oxide Modified Bioink for 3D-Bioprinting of Vascular Graft. *ACS Appl bio Mater*. 2025;8(5):3858-3872.
doi: 10.1021/acsabm.5c00004
129. Liu N, Ye X, Yao B, *et al*. Advances in 3D bioprinting technology for cardiac tissue engineering and regeneration. *Bioact Mater*. 2021;6(5):1388-1401.
doi: 10.1016/j.bioactmat.2020.10.021
130. Shin SR, Aghaei-Ghareh-Bolagh B, Dang TT, *et al*. Cell-laden microengineered and mechanically tunable hybrid hydrogels of gelatin and graphene oxide. *Adv Mater*. 2013;25(44):6385-6391.
doi: 10.1002/adma.201301082
131. Zhou J, Yang X, Liu W, *et al*. Injectable OPF/graphene oxide hydrogels provide mechanical support and enhance cell electrical signaling after implantation into myocardial infarct. *Theranostics*. 2018;8(12):3317-3330.
doi: 10.7150/THNO.25504
132. Noor N, Shapira A, Edri R, Gal I, Wertheim L, Dvir T. 3D Printing of Personalized Thick and Perfusable Cardiac Patches and Hearts. *Adv Sci*. 2019;6(11):1900344.
doi: 10.1002/advs.201900344
133. Dai X, Zhang Y, Gao L, *et al*. A mechanically strong, highly stable, thermoplastic, and self-healable supramolecular polymer hydrogel. *Adv Mater*. 2015;27(23):3566-3571.
doi: 10.1002/adma.201500534
134. Bellet P, Gasparotto M, Pressi S, *et al*. Graphene-Based Scaffolds for Regenerative Medicine. *Nanomater (Basel, Switzerland)*. 2021;11(2):1-41.
doi: 10.3390/nano11020404
135. Oubari H, Berkane Y, Jeljeli M, Lellouch AG, Smadja DM. Engineering the Future of Stem Cells in Vascular Reconstruction: A Leap Towards Functional Endothelialized Tissue-Engineered Vascular Conduits. *Stem cell Rev reports*. 2025;21(8):2796-2806.
doi: 10.1007/s12015-025-10968-8
136. Paul A, Hasan A, Kindi H Al, *et al*. Injectable graphene oxide/hydrogel-based angiogenic gene delivery system for vasculogenesis and cardiac repair. *ACS Nano*. 2014;8(8):8050-8062.
doi: 10.1021/nn5020787
137. Landau S, Okhovatian S, Zhao Y, *et al*. Bioengineering vascularization. *Dev*. 2024;151(23):dev204455.
doi: 10.1242/dev.204455/363262
138. Wang X, He B. Endothelial dysfunction: molecular mechanisms and clinical implications. *MedComm*. 2024;5(8):e651.
doi: 10.1002/mcO2.651
139. Lu Y, Yang G, Wang S, *et al*. Stretchable graphene-hydrogel interfaces for wearable and implantable bioelectronics. *Nat Electron*. 2024;7(1):51-65.
doi: 10.1038/s41928-023-01091-y
140. Cognetti JS, Moen MT, Brewer MG, *et al*. A photonic biosensor-integrated tissue chip platform for real-time sensing of lung epithelial inflammatory markers. *Lab Chip*. 2023;23(2):239-250.
doi: 10.1039/d2lc00864e
141. Shinde A, Illath K, Kasiviswanathan U, *et al*. Recent Advances of Biosensor-Integrated Organ-on-a-Chip Technologies for Diagnostics and Therapeutics. *Anal Chem*. 2023;95(6):3121-3146.
doi: 10.1021/acs.analchem.2C05036
142. Yang W, Li T, Liao S, Zhou J, Huang L. Organ-on-a-chip platforms integrated with biosensors for precise monitoring of the cells and cellular microenvironment. *TrAC Trends Anal Chem*. 2024;172:117569.
doi: 10.1016/j.trac.2024.117569
143. Kosowska K, Korycka P, Jankowska-Snopkiewicz K, *et al*. Graphene Oxide (GO)-Based Bioink with Enhanced 3D Printability and Mechanical Properties for Tissue Engineering Applications. *Nanomater (Basel, Switzerland)*. 2024;14(9):760.
doi: 10.3390/nano14090760
144. Lee YJ, Ajiteru O, Lee JS, *et al*. Highly conductive, stretchable, and biocompatible graphene oxide biocomposite hydrogel for advanced tissue engineering. *Biofabrication*. 2024;16(4):045032.
doi: 10.1088/1758-5090/ad6cf7
145. Chen X, Manshahi F, Tioran K, *et al*. Wearable biosensors for

- cardiovascular monitoring leveraging nanomaterials. *Adv Compos Hybrid Mater.* 2024;7(3):97.
doi: 10.1007/s42114-024-00906-6
146. Coronado Reyes OI, Téllez Anguiano A del C, Gutiérrez Gneccchi JA, Castro Pimentel LA, García Rodríguez E. Comparison between mathematical methods to estimate blood glucose levels from ECG signals. *Biosens Bioelectron X.* 2024;18:100474.
doi: 10.1016/j.biosx.2024.100474
147. Li J, Zeng H, Zeng Z, Zeng Y, Xie T. Promising Graphene-Based Nanomaterials and Their Biomedical Applications and Potential Risks: A Comprehensive Review. *ACS Biomater Sci Eng.* 2021;7(12):5363-5396.
doi: 10.1021/acsbomaterials.1c00875
148. Lin H, Buerki-Thurnherr T, Kaur J, *et al.* Environmental and Health Impacts of Graphene and Other Two-Dimensional Materials: A Graphene Flagship Perspective. *ACS Nano.* 2024;18(8):6038-6094.
doi: 10.1021/acsnano.3c09699
149. Donato KZ, Tan HL, Marangoni VS, *et al.* Graphene oxide classification and standardization. *Sci Rep.* 2023;13(1):6064.
doi: 10.1038/S41598-023-33350-5
150. Loret T, de Luna LAV, Lucherelli MA, *et al.* Lung Persistence, Biodegradation, and Elimination of Graphene-Based Materials are Predominantly Size-Dependent and Mediated by Alveolar Phagocytes. *Small.* 2023;19(39):e2301201.
doi: 10.1002/sml.202301201
151. Zhang Z, Schniepp HC, Adamson DH. Characterization of graphene oxide: Variations in reported approaches. *Carbon N Y.* 2019;154:510-521.
doi: 10.1016/j.carbon.2019.07.103
152. Tao W, Zhu X, Yu X, *et al.* Black Phosphorus Nanosheets as a Robust Delivery Platform for Cancer Theranostics. *Adv Mater.* 2017;29(1):1603276.
doi: 10.1002/adma.201603276
153. Wu J, Xu R, Shao M, Zhao L, Xu W, Guo Y. Phosphorus-Based Nanomaterials for Biomedical Applications: A Review. *ACS Appl Nano Mater.* 2024;7(10):11022-11036.
doi: 10.1021/acsnm.4c00015
154. Guo Y, Huang C, Zhang J, Wang Z, Guo X, Wei Y. Injectable Conductive Hydrogel Containing Black Phosphorus Nanosheets Inhibits the Oxidative Stress-Inflammation Chain during Myocardial Infarction. *ACS Appl Nano Mater.* 2023;6(18):16749-16767.
doi: 10.1021/acsnm.3c02956
155. Ling X, Wang H, Huang S, Xia F, Dresselhaus MS. The renaissance of black phosphorus. *Proc Natl Acad Sci U S A.* 2015;112(15):4523-4530.
doi: 10.1073/pnas.1416581112
156. Qian X, Gu Z, Chen Y. Two-dimensional black phosphorus nanosheets for theranostic nanomedicine. *Mater Horizons.* 2017;4(5):800-816.
doi: 10.1039/c7mh00305f
157. Sun Z, Xie H, Tang S, *et al.* Ultrasmall Black Phosphorus Quantum Dots: Synthesis and Use as Photothermal Agents. *Angew Chem Int Ed Engl.* 2015;54(39):11526-11530.
doi: 10.1002/anie.201506154
158. Shao J, Xie H, Huang H, *et al.* Biodegradable black phosphorus-based nanospheres for in vivo photothermal cancer therapy. *Nat Commun.* 2016;7:12967.
doi: 10.1038/ncomms12967
159. Li H xuan, Zhao K chi, Jiang J jia, Zhu Q san. Research progress on black phosphorus hybrids hydrogel platforms for biomedical applications. *J Biol Eng.* 2023;17(1):8.
doi: 10.1186/s13036-023-00328-w
160. Choi JR, Yong KW, Choi JY, *et al.* Black Phosphorus and its Biomedical Applications. *Theranostics.* 2018;8(4):1005-1026.
doi: 10.7150/THNO.22573
161. Bai X, Wang R, Hu X, *et al.* Two-Dimensional Biodegradable Black Phosphorus Nanosheets Promote Large Full-Thickness Wound Healing through In Situ Regeneration Therapy. *ACS Nano.* 2024;18(4):3553-3574.
doi: 10.1021/acsnano.3c11177
162. Wu N, Wang X, Das CM, *et al.* Bioengineering applications of black phosphorus and their toxicity assessment. *Environ Sci Nano.* 2021;8(12):3452-3477.
doi: 10.1039/d1en00273b
163. Zhang T, Wan Y, Xie H, *et al.* Degradation Chemistry and Stabilization of Exfoliated Few-Layer Black Phosphorus in Water. *J Am Chem Soc.* 2018;140(24):7561-7567.
doi: 10.1021/jacs.8b02156
164. Qiu Y, Yu C, Yue Z, *et al.* Chronological-Programmed Black Phosphorus Hydrogel for Responsive Modulation of the Pathological Microenvironment in Myocardial Infarction. *ACS Appl Mater Interfaces.* 2024;16(14):17323-17338.
doi: 10.1021/acsnm.4c01956
165. Xiang Q, Yi X, Zhu XH, Wei X, Jiang DS. Regulated cell death in myocardial ischemia-reperfusion injury. *Trends Endocrinol Metab.* 2024;35(3):219-234.
doi: 10.1016/j.tem.2023.10.010
166. Qin L, Ling G, Peng F, *et al.* Black phosphorus nanosheets and gemcitabine encapsulated thermo-sensitive hydrogel for synergistic photothermal-chemotherapy. *J Colloid Interface Sci.* 2019;556:232-238.

- doi: 10.1016/j.jcis.2019.08.058
167. Qiu M, Wang D, Liang W, *et al.* Novel concept of the smart NIR-light-controlled drug release of black phosphorus nanostructure for cancer therapy. *Proc Natl Acad Sci U S A.* 2018;115(3):501-506.
doi: 10.1073/pnas.1714421115
168. Chen J, Huan W, Mao L, *et al.* Impaired barrier integrity of endothelial cells induced by PEGylated black phosphorus nanosheets. *Sci Total Environ.* 2023;861:160645.
doi: 10.1016/j.scitotenv.2022.160645
169. Xu Y, Xu C, He L, *et al.* Stratified-structural hydrogel incorporated with magnesium-ion-modified black phosphorus nanosheets for promoting neuro-vascularized bone regeneration. *Bioact Mater.* 2022;16:271-284.
doi: 10.1016/j.bioactmat.2022.02.024
170. Cui H, Yu ZX, Huang Y, *et al.* 3D printing of thick myocardial tissue constructs with anisotropic myofibers and perfusable vascular channels. *Biomater Adv.* 2023;153:213579.
doi: 10.1016/j.bioadv.2023.213579
171. Sakamoto A, Suwa K, Kawakami R, *et al.* Significance of Intra-plaque Hemorrhage for the Development of High-Risk Vulnerable Plaque: Current Understanding from Basic to Clinical Points of View. *Int J Mol Sci.* 2023;24(17):13298.
doi: 10.3390/ijms241713298
172. Ugunusman A, Hisam NSN, Othman NS, *et al.* Pharmacological interventions for intraplaque neovascularization in atherosclerosis. *Pharmacol Ther.* 2024;261:108685.
doi: 10.1016/j.pharmthera.2024.108685
173. Wang H, Gui B, Chen Y, *et al.* Black-Phosphorus-Reinforced Injectable Conductive Biodegradable Hydrogel for the Delivery of ADSC-Derived Exosomes to Repair Myocardial Infarction. *ACS Appl Mater Interfaces.* 2024;16(43):58286-58298.
doi: 10.1021/acsami.4c12285
174. Hasan MM, Ahmad A, Akter MZ, Choi YJ, Yi HG. Bioinks for bioprinting using plant-derived biomaterials. *Biofabrication.* 2024;16(4):042004.
doi: 10.1088/1758-5090/ad6932
175. Emebu S, Ogunleye RO, Achbergerová E, Vítková L, Ponížil P, Martinez CM. Review and proposition for model-based multivariable-multiobjective optimisation of extrusion-based bioprinting. *Appl Mater Today.* 2023;34:101914.
doi: 10.1016/j.apmt.2023.101914
176. Wang J, Cui Z, Maniruzzaman M. Bioprinting: A focus on improving bioink printability and cell performance based on different process parameters. *Int J Pharm.* 2023;640:123020.
doi: 10.1016/j.ijpharm.2023.123020
177. Cooke ME, Rosenzweig DH. The rheology of direct and suspended extrusion bioprinting. *APL Bioeng.* 2021;5(1):011502.
doi: 10.1063/5.0031475
178. Ribeiro A, Blokzijl MM, Levato R, *et al.* Assessing bioink shape fidelity to aid material development in 3D bioprinting. *Biofabrication.* 2018;10(1):014102.
doi: 10.1088/1758-5090/aa90e2
179. Groll J, Burdick JA, Cho DW, *et al.* A definition of bioinks and their distinction from biomaterial inks. *Biofabrication.* 2019;11(1):013001.
doi: 10.1088/1758-5090/aaec52
180. Guo J, Du L. An update on ox-LDL-inducing vascular smooth muscle cell-derived foam cells in atherosclerosis. *Front Cell Dev Biol.* 2024;12:1-12.
doi: 10.3389/fcell.2024.1481505
181. Kong Z, Wang X. Bioprinting Technologies and Bioinks for Vascular Model Establishment. *Int J Mol Sci.* 2023;24(1):891.
doi: 10.3390/ijms24010891
182. Ashtari K, Nazari H, Ko H, *et al.* Electrically conductive nanomaterials for cardiac tissue engineering. *Adv Drug Deliv Rev.* 2019;144:162-179.
doi: 10.1016/j.addr.2019.06.001
183. Luo X, Zhang M, Dai W, *et al.* Targeted nanoparticles triggered by plaque microenvironment for atherosclerosis treatment through cascade effects of reactive oxygen species scavenging and anti-inflammation. *J Nanobiotechnology.* 2024;22(1):440.
doi: 10.1186/s12951-024-02652-9
184. Chen X, Sun Z, Peng X, *et al.* Graphene Oxide/Black Phosphorus Functionalized Collagen Scaffolds with Enhanced Near-Infrared Controlled In Situ Biomineralization for Promoting Infectious Bone Defect Repair through PI3K/Akt Pathway. *ACS Appl Mater Interfaces.* 2024;16(38):50369-50388.
doi: 10.1021/acsami.4c10284
185. Akther F, Sajin D, Moonshi SS, *et al.* An intimal-lumen model in a microfluidic device: potential platform for atherosclerosis-related studies. *Lab Chip.* 2025;25(3):354-369.
doi: 10.1039/d4lc00868e
186. Chegel R, Behzad S. Tunable Electronic, Optical, and Thermal Properties of two-dimensional Germanene via an external electric field. *Sci Rep.* 2020;10(1):704.
doi: 10.1038/S41598-020-57558-x
187. Rohaizad N, Mayorga-Martinez CC, Fojtů M, Latiff NM, Pumera M. Two-dimensional materials in biomedical, biosensing and sensing applications. *Chem Soc Rev.* 2021;50(1):619-657.

- doi: 10.1039/d0CS00150c
188. Tao W, Kong N, Ji X, *et al.* Emerging two-dimensional monoelemental materials (Xenes) for biomedical applications. *Chem Soc Rev.* 2019;48(11):2891-2912.
doi: 10.1039/c8cs00823j
189. Kang Y, Zhang H, Chen L, *et al.* The marriage of Xenes and hydrogels: Fundamentals, applications, and outlook. *Innov.* 2022;3(6):100327.
doi: 10.1016/j.xinn.2022.100327
190. Garg M, Thakur A. A review: Biomedical applications of phosphorene, antimonene, and germanene-based 2D material/hydrogel complexes. *J Mater Sci.* 2023;58(1):34-45.
doi: 10.1007/S10853-022-07954-7
191. Ge M, Zong M, Xu D, *et al.* Freestanding germanene nanosheets for rapid degradation and photothermal conversion. *Mater Today Nano.* 2021;15:100119.
doi: 10.1016/j.mtnano.2021.100119
192. Liu N, Bo G, Liu Y, Xu X, Du Y, Dou SX. Recent Progress on Germanene and Functionalized Germanene: Preparation, Characterizations, Applications, and Challenges. *Small.* 2019;15(32):1805147.
doi: 10.1002/sml.201805147
193. Ng S, Pumera M. 2D Functionalized Germanenes: Synthesis and Applications. *Adv Mater.* 2023;35(7):1-37.
doi: 10.1002/adma.202207196
194. Kupchak I, Bechstedt F, Pulci O, Gori P. Tuning the optical absorption and exciton bound states of germanene by chemical functionalization. *Sci Rep.* 2024;14(1):25182.
doi: 10.1038/S41598-024-75620-W
195. Kovalska E, Antonatos N, Luxa J, Sofer Z. Edge-Hydrogenated Germanene by Electrochemical Decalcification-Exfoliation of CaGe₂: Germanene-Enabled Vapor Sensor. *ACS Nano.* 2021;15(10):16709-16718.
doi: 10.1021/acsnano.1c06675
196. Miri AK, Khalilpour A, Cecen B, Maharjan S, Shin SR, Khademhosseini A. Multiscale bioprinting of vascularized models. *Biomaterials.* 2019;198:204-216.
doi: 10.1016/j.biomaterials.2018.08.006
197. Samal S, Sahoo SP, Acharya B. Nanotechnology-Driven cardiac tissue engineering and 3D bioprinting: Mechanistic insights into myocardial repair and regeneration. *Nano Trends.* 2025;12:100155.
doi: 10.1016/j.nwnano.2025.100155
198. Karvinen J, Kellomäki M. Design aspects and characterization of hydrogel-based bioinks for extrusion-based bioprinting. *Bioprinting.* 2023;32:e00274.
doi: 10.1016/j.bprint.2023.e00274
199. Suzuki S, Katsube D, Yano M, *et al.* Germanene Reformation from Oxidized Germanene on Ag(111)/Ge(111) by Vacuum Annealing. *Small methods.* 2025;9(3):e2400863.
doi: 10.1002/smt.202400863
200. Ouyang J, Feng C, Ji X, *et al.* 2D Monoelemental Germanene Quantum Dots: Synthesis as Robust Photothermal Agents for Photonic Cancer Nanomedicine. *Angew Chem Int Ed Engl.* 2019;58(38):13405-13410.
doi: 10.1002/ANIE.201908377
201. Geevarghese R, Żur-Pińska J, Parisi D, Włodarczyk-Biegun MK. A comprehensive protocol for hydrogel-based bioink design: balancing printability, stability, and biocompatibility. *J Mater Chem B.* 2025;13(42):13750-13768.
doi: 10.1039/d5tb00737b
202. Resende MAA, Rigo ECS, Vercik A. Printability of Bioinks: A Consolidated Definition for Additive Manufacturing. *ACS omega.* 2025;10(48):58110-58122.
doi: 10.1021/acsomega.5c00727
203. Bianco E, Butler S, Jiang S, Restrepo OD, Windl W, Goldberger JE. Stability and Exfoliation of Germanene: A Germanium Graphene Analogue. *ACS Nano.* 2013;7(5):4414-4421.
doi: 10.1021/nn4009406
204. Wang H, Wang C, Zhang Y, *et al.* Recent Advances in Xenes Based FET for Biosensing Applications. *Adv Sci (Weinh).* 2025;12(21):e2500752.
doi: 10.1002/advs.202500752
205. Duan X, Liu Z, Xie Z, *et al.* Emerging monoelemental 2D materials (Xenes) for biosensor applications. *Nano Res.* 2023;16(5):7030-7052.
doi: 10.1007/S12274-023-5418-3
206. Dixit M, Shrestha LK, Ariga K, Pati F. Carbon-Based Nanoarchitectonics in Advancing Cardiac Tissue Bioprinting: A Review. *ACS Appl Nano Mater.* 2024;7(21):24638-24652.
doi: 10.1021/acsanm.4C04441
207. Ding Q, Sun T, Su W, *et al.* Bioinspired Multifunctional Black Phosphorus Hydrogel with Antibacterial and Antioxidant Properties: A Stepwise Countermeasure for Diabetic Skin Wound Healing. *Adv Healthc Mater.* 2022;11(12):e2102791.
doi: 10.1002/adhm.202102791
208. Wu Y, Okesola BO, Xu J, *et al.* Disordered protein-graphene oxide co-assembly and supramolecular biofabrication of functional fluidic devices. *Nat Commun.* 2020;11(1):1182.
doi: 10.1038/S41467-020-14716-Z
209. Kaviani S, Talebi A, Labbaf S, Karimzadeh F. Conductive GelMA/alginate/polypyrrole/graphene hydrogel as a potential scaffold for cardiac tissue engineering: Physiochemical, mechanical, and biological evaluations. *Int J Biol Macromol.* 2024;259(Pt 2):129276.

- doi: 10.1016/j.ijbiomac.2024.129276
210. Liu F, Ding N, Huo D, *et al.* Surface-Engineered Monocyte Inhibits Atherosclerotic Plaque Destabilization via Graphene Quantum Dot-Mediated MicroRNA Delivery. *Adv Healthc Mater.* 2019;8(15):e1900386.
doi: 10.1002/adhm.201900386
211. Chung S, Revia RA, Zhang M. Graphene Quantum Dots and Their Applications in Bioimaging, Biosensing, and Therapy. *Adv Mater.* 2021;33(22):e1904362.
doi: 10.1002/adma.201904362
212. Yan Z, Liu Z, Yang B, Zhu X, Song E, Song Y. Long-term exposure of molybdenum disulfide nanosheets leads to hepatic lipid accumulation and atherogenesis in apolipoprotein E deficient mice. *NanoImpact.* 2023;30:100462.
doi: 10.1016/j.impact.2023.100462
213. Ge X, Cui H, Kong J, *et al.* A Non-Invasive Nanoprobe for In Vivo Photoacoustic Imaging of Vulnerable Atherosclerotic Plaque. *Adv Mater.* 2020;32(38):e2000037.
doi: 10.1002/adma.202000037
214. Bai Q, Lao X, Pang SY, *et al.* Plaque-Targeted Delivery of Fluoride-Free MXene Nanozyme for Alleviating Atherosclerosis via Sonocatalytic Therapy. *Adv Mater.* 2025;37(48):1-16.
doi: 10.1002/adma.202420189
215. Leung CM, de Haan P, Ronaldson-Bouchard K, *et al.* A guide to the organ-on-a-chip. *Nat Rev Methods Prim.* 2022;2(1):33.
doi: 10.1038/s43586-022-00118-6
216. Augustin HG, Koh GY. Organotypic vasculature: From descriptive heterogeneity to functional pathophysiology. *Science.* 2017;357(6353):1-12.
doi: 10.1126/science.aal2379
217. Li J, Chen X, Hu M, *et al.* The application of composite scaffold materials based on decellularized vascular matrix in tissue engineering: a review. *Biomed Eng Online.* 2023;22(1):62.
doi: 10.1186/S12938-023-01120-z
218. Ouyang Z, Zhong J, Shen J, Zeng Y. The cell origins of foam cell and lipid metabolism regulated by mechanical stress in atherosclerosis. *Front Physiol.* 2023;14:01-09.
doi: 10.3389/fphys.2023.1179828
219. Fernandez DM, Rahman AH, Fernandez NF, *et al.* Single-cell immune landscape of human atherosclerotic plaques. *Nat Med.* 2019;25(10):1576-1588.
doi: 10.1038/S41591-019-0590-4
220. La Chica Lhoëst MT, Martinez A, Claudi L, *et al.* Mechanisms modulating foam cell formation in the arterial intima: exploring new therapeutic opportunities in atherosclerosis. *Front Cardiovasc Med.* 2024;11:1381520.
doi: 10.3389/fcvm.2024.1381520
221. Chen J, Zhang X, Millican R, *et al.* Recent Progress in in vitro Models for Atherosclerosis Studies. *Front Cardiovasc Med.* 2021;8:1-23.
doi: 10.3389/fcvm.2021.790529
222. Wang R, Zhang H, Li S, *et al.* Current progress of in vitro vascular models on microfluidic chips. *Biofabrication.* 2025;17(2):022004.
doi: 10.1088/1758-5090/adb182
223. Garcia-Sabaté A, Mohamed WKE, Sapudom J, Alatoom A, Al Safadi L, Teo JCM. Biomimetic 3D Models for Investigating the Role of Monocytes and Macrophages in Atherosclerosis. *Bioeng (Basel, Switzerland).* 2020;7(3):1-20.
doi: 10.3390/bioengineering7030113
224. Akther F, Sajin D, Moonshi SS, Wu Y, Vazquez-Prada KX, Ta HT. Modeling Foam Cell Formation in A Hydrogel-Based 3D-Intimal Model: A Study of The Role of Multi-Diseases During Early Atherosclerosis. *Adv Biol.* 2024;8(4):e2300463.
doi: 10.1002/adbi.202300463
225. Naiyeju I, Lehoux S, Tabrizian M. Key parameters for designing robust 2D and 3D spheroid models for in vitro atherosclerosis research. *Bioeng Transl Med.* 2025;10(3):e10736.
doi: 10.1002/btm2.10736
226. Krug A, Inserra G, Drewes R, *et al.* Three-dimensional spheroid models for cardiovascular biology and pathology. *Mechanobiol Med.* 2025;3(3):100144.
doi: 10.1016/j.mbm.2025.100144
227. Parma L, Sachs N, Sobczak N, *et al.* CXCL12 Derived From ACKR1+ Intraplaque Neovessels Mediates CD8+ T Cell Recruitment in Human Atherosclerosis. *Circulation.* 2025;151(8):581-584.
doi: 10.1161/CIRCULATIONAHA.124.072560
228. Mallone A, Stenger C, Von Eckardstein A, Hoerstrup SP, Weber B. Biofabricating atherosclerotic plaques: In vitro engineering of a three-dimensional human fibroatheroma model. *Biomaterials.* 2018;150:49-59.
doi: 10.1016/j.biomaterials.2017.09.034
229. Wang Y, Liu A, Zhang X, *et al.* Microfluidic organ-on-a-chip for modeling coronary artery disease: Recent applications, limitations and potential. *J Tissue Eng.* 2025;16:20417314251394447.
doi: 10.1177/20417314251394447
230. Quintard C, Tubbs E, Jonsson G, *et al.* A microfluidic platform integrating functional vascularized organoids-on-chip. *Nat Commun.* 2024;15(1):1452.
doi: 10.1038/s41467-024-45710-4

231. Chandra Sekar N, Khoshmanesh K, Baratchi S. Bioengineered models of cardiovascular diseases. *Atherosclerosis*. 2024;393:117565.
doi: 10.1016/j.atherosclerosis.2024.117565
232. Chang SQ, Qiao L, Abodunrin OD, Zou L, Huang NP. Advanced technologies of artery-on-a-chip: a review of construction strategies and disease models. *Angiogenesis*. 2026;29(2):17.
doi: 10.1007/S10456-026-10030-2
233. Yarbrough D, Chen R, Katsouleas K, *et al.* Artery-on-Chip Demonstrates Mechanical and Functional Features of Healthy and Diseased Living Smooth Muscle Tissue. *Adv Funct Mater*. Published online July 1, 2026. 2026:e76324.
doi: 10.1002/adfm.76324
234. Gold KA, Saha B, Rajeeva Pandian NK, *et al.* 3D Bioprinted Multicellular Vascular Models. *Adv Healthc Mater*. 2021;10(21):e2101141.
doi: 10.1002/adhm.202101141
235. Wang Z, Lee SJ, Cheng HJ, Yoo JJ, Atala A. 3D bioprinted functional and contractile cardiac tissue constructs. *Acta Biomater*. 2018;70:48-56.
doi: 10.1016/j.actbio.2018.02.007
236. Dell AC, Maresca J, Davis BA, Isaji T, Dardik A, Geibel JP. Development and deployment of a functional 3D-bioprinted blood vessel. *Sci Reports*. 2025;15(1):11668.
doi: 10.1038/s41598-025-93276-y
237. Krajewska U, Chechlińska C, Kurzyk A, Vascularisation in 3D bioprinted models: emerging solutions engineering functional tissues and tumour models. *Biofabrication*. 2026;18(2):022001.
doi: 10.1088/1758-5090/AE2F02
238. Liao Y, Gallegos-Martínez S, Kuang X, Du Y, Zhang YS, Zhang Y. Hybrid bioprinting of hierarchical vascular networks at capillary-scale resolution. *Nat Chem Eng*. 2026;3(6):328-339.
doi: 10.1038/s44286-026-00396-x
239. Chandra DK, Reis RL, Kundu SC, Kumar A, Mahapatra C. Nanomaterials-Based Hybrid Bioink Platforms in Advancing 3D Bioprinting Technologies for Regenerative Medicine. *ACS Biomater Sci Eng*. 2024;10(7):4145-4174.
doi: 10.1021/acsbomaterials.4c00166
240. Jin X, Li Y, Ran H, Zhang Z, Cheng P, Wu Y. Smart nanomaterial-crosslinked hydrogels for biomedical applications. *Smart Mater Med*. 2025;6(3):417-433.
doi: 10.1016/j.smaim.2025.11.001
241. Ribezzi D, Zegwaart JP, Van Gansbeke T, *et al.* Multi-material Volumetric Bioprinting and Plug-and-play Suspension Bath Biofabrication via Bioresin Molecular Weight Tuning and via Multiwavelength Alignment Optics. *Adv Mater*. 2025;37(13):e2409355.
doi: 10.1002/adma.202409355
242. Aizarna-Lopetegui U, Größbacher G, Herrero-Ruiz A, *et al.* Hybrid Plasmonic Bioresins and dECM-Based Materials for Volumetric Bioprinting of Vascular-Inspired Architectures. *ACS Appl Mater Interfaces*. 2025;17(25):36982-36991.
doi: 10.1021/acscami.5c03880
243. Kong D, Ryu JC, Shin N, *et al.* In Vitro Modeling of Atherosclerosis Using iPSC-Derived Blood Vessel Organoids. *Adv Healthc Mater*. 2025;14(1):e2400919.
doi: 10.1002/adhm.202400919
244. Taylor CA, Figueroa CA. Patient-specific modeling of cardiovascular mechanics. *Annu Rev Biomed Eng*. 2009;11:109-134.
doi: 10.1146/annurev.bioeng.10.061807.160521
245. Jackson ML, Bond AR, George SJ. Mechanobiology of the endothelium in vascular health and disease: in vitro shear stress models. *Cardiovasc drugs Ther*. 2023;37(5):997-1010.
doi: 10.1007/S10557-022-07385-1
246. Aitken C, Mehta V, Schwartz MA, Tzima E. Mechanisms of endothelial flow sensing. *Nat Cardiovasc Res*. 2023;2(6):517-529.
doi: 10.1038/S44161-023-00276-0
247. Tabas I, Bornfeldt KE. Macrophage Phenotype and Function in Different Stages of Atherosclerosis. *Circ Res*. 2016;118(4):653-667.
doi: 10.1161/CIRCRESAHA.115.306256
248. De Meyer GRY, Zurek M, Puylaert P, Martinet W. Programmed death of macrophages in atherosclerosis: mechanisms and therapeutic targets. *Nat Rev Cardiol*. 2024;21(5):312-325.
doi: 10.1038/S41569-023-00957-0
249. Lee J, Choi JH. Deciphering Macrophage Phenotypes upon Lipid Uptake and Atherosclerosis. *Immune Netw*. 2020;20(3):1-21.
doi: 10.4110/IN.2020.20.E22
250. Moore KJ, Sheedy FJ, Fisher EA. Macrophages in atherosclerosis: a dynamic balance. *Nat Rev Immunol*. 2013;13(10):709-721.
doi: 10.1038/NRI3520
251. Roy P, Orecchioni M, Ley K. How the immune system shapes atherosclerosis: roles of innate and adaptive immunity. *Nat Rev Immunol*. 2022;22(4):251-265.
doi: 10.1038/S41577-021-00584-1
252. Guan Y, Racioppi L, Gerecht S. Engineering biomaterials to tailor the microenvironment for macrophage-endothelium

- interactions. *Nat Rev Mater*. 2023;8(10):688-699.
doi: 10.1038/s41578-023-00591-9
253. Imhof BA, Aurrand-Lions M. Adhesion mechanisms regulating the migration of monocytes. *Nat Rev Immunol*. 2004;4(6):432-444.
doi: 10.1038/nri1375
254. Whitworth CP, Polacheck WJ. Vascular organs-on-chip made with patient-derived endothelial cells: technologies to transform drug discovery and disease modeling. *Expert Opin Drug Discov*. 2024;19(3):339-351.
doi: 10.1080/17460441.2023.2294947
255. Jiang H, Li X, Chen T, *et al*. Bioprinted vascular tissue: Assessing functions from cellular, tissue to organ levels. *Mater today Bio*. 2023;23:100846.
doi: 10.1016/j.mtbio.2023.100846
256. Tabish TA, Zhu Y, Shukla S, *et al*. Graphene nanocomposites for real-time electrochemical sensing of nitric oxide in biological systems. *Appl Phys Rev*. 2023;10(4):041310.
doi: 10.1063/5.0162640
257. Ochieng BO, Zhao L, Ye Z. Three-Dimensional Bioprinting in Vascular Tissue Engineering and Tissue Vascularization of Cardiovascular Diseases. *Tissue Eng Part B Rev*. 2024;30(3):340-358.
doi: 10.1089/TEN.TEB.2023.0175
258. Lai J, Liu Y, Lu G, *et al*. 4D bioprinting of programmed dynamic tissues. *Bioact Mater*. 2024;37:348-377.
doi: 10.1016/j.bioactmat.2024.03.033
259. Liu J, Zhu M, Bai N, *et al*. Microfluidic Perfusable Pathological Vasculature for Atherosclerosis Drug Screening. *Res (Washington, DC)*. 2025;8:0902.
doi: 10.34133/research.0902
260. Chae S, Ha DH, Lee H. 3D bioprinting strategy for engineering vascularized tissue models. *Int J bioprinting*. 2023;9(5):748.
doi: 10.18063/ijb.748
261. Kizhisseri M, Gharaie S, Schluter J. An analytical method informed by clinical imaging data for estimating outlet boundary conditions in computational fluid dynamics analysis of carotid artery blood flow. *Sci Rep*. 2023;13(1):14973.
doi: 10.1038/S41598-023-42004-5
262. Mir A, Lee E, Shih W, *et al*. 3D Bioprinting for Vascularization. *Bioeng (Basel, Switzerland)*. 2023;10(5):606.
doi: 10.3390/bioengineering10050606
263. Zhou Y, Sekar NC, Thurgood P, *et al*. Bioengineered Vascular Model of Foam Cell Formation. *ACS Biomater Sci Eng*. 2023;9(12):6947-6955.
doi: 10.1021/acsbiomaterials.3c01308
264. Agarwal T, Onesto V, Banerjee D, *et al*. 3D bioprinting in tissue engineering: current state-of-the-art and challenges towards system standardization and clinical translation. *Biofabrication*. 2025;17(4):045001.
doi: 10.1088/1758-5090/ade47a
265. Wang Z, Hao Z, Yu S, Huang C, Pan Y, Zhao X. A Wearable and Deformable Graphene-Based Affinity Nanosensor for Monitoring of Cytokines in Biofluids. *Nanomater (Basel, Switzerland)*. 2020;10(8):1-9.
doi: 10.3390/nano10081503
266. Sun Z, Zhao J, Leung E, *et al*. Three-Dimensional Bioprinting in Cardiovascular Disease: Current Status and Future Directions. *Biomolecules*. 2023;13(8):1180.
doi: 10.3390/biom13081180
267. Kim J. Characterization of Biocompatibility of Functional Bioinks for 3D Bioprinting. *Bioeng (Basel, Switzerland)*. 2023;10(4):457.
doi: 10.3390/bioengineering10040457
268. Alves T, Mota WS, Barros C, *et al*. Review of scientific literature and standard guidelines for the characterization of graphene-based materials. *J Mater Sci*. 2024;59(32):14948-14980.
doi: 10.1007/S10853-024-10061-4
269. Bai M, Wan H, Zhang Y, *et al*. Two-dimensional nanomaterials based on rare earth elements for biomedical applications. *Chem Sci*. 2024;15(41):16887-16907.
doi: 10.1039/d4sc02625j
270. Wang Z, Guo H, Zhang J, Qian Y, Liu Y. Two-Dimensional Nanomaterials in Hydrogels and Their Potential Bio-Applications. *Lubr*. 2024;12(5):149.
doi: 10.3390/lubricants12050149
271. Libby P. The changing landscape of atherosclerosis. *Nature*. 2021;592(7855):524-533.
doi: 10.1038/S41586-021-03392-8
272. Ou L, Tan X, Qiao S, *et al*. Graphene-Based Material-Mediated Immunomodulation in Tissue Engineering and Regeneration: Mechanism and Significance. *ACS Nano*. 2023;17(19):18669-18687.
doi: 10.1021/acsnano.3C03857
273. Ding X, Pu Y, Tang M, Zhang T. Pulmonary hazard identifications of Graphene family nanomaterials: Adverse outcome pathways framework based on toxicity mechanisms. *Sci Total Environ*. 2023;857(Pt 1):159329.
doi: 10.1016/j.scitotenv.2022.159329
274. Smith LS, Haidari H, Amsalu A, *et al*. Black Phosphorus Nanoflakes: An Emerging Nanomaterial for Clinical Wound Management and Biomedical Applications. *Int J Mol Sci*.

- 2024;25(23):12824.
doi: 10.3390/ijms252312824
275. Yedgar S, Barshtein G, Gural A. Hemolytic Activity of Nanoparticles as a Marker of Their Hemocompatibility. *Micromachines*. 2022;13(12):2091.
doi: 10.3390/mi13122091
276. Kim Y, Chica-Carrillo EC, Lee HJ. Microfabricated sensors for non-invasive, real-time monitoring of organoids. *Micro Nano Syst Lett*. 2024;12(1):26.
doi: 10.1186/S40486-024-00216-y
277. Liu S, Kumari S, He H, *et al*. Biosensors integrated 3D organoid/organ-on-a-chip system: A real-time biomechanical, biophysical, and biochemical monitoring and characterization. *Biosens Bioelectron*. 2023;231:115285.
doi: 10.1016/j.bios.2023.115285
278. Flynn CD, Chang D, Mahmud A, *et al*. Biomolecular sensors for advanced physiological monitoring. *Nat Rev Bioeng*. 2023;1(8):560-575.
doi: 10.1038/s44222-023-00067-z
279. Wu J, Liu H, Chen W, Ma B, Ju H. Device integration of electrochemical biosensors. *Nat Rev Bioeng*. 2023;1(5):346-360.
doi: 10.1038/S44222-023-00032-W
280. Chen S, Sun Y, Fan X, *et al*. Review on two-dimensional material-based field-effect transistor biosensors: accomplishments, mechanisms, and perspectives. *J Nanobiotechnology*. 2023;21(1):144.
doi: 10.1186/S12951-023-01898-z
281. He W, Deng J, Ma B, *et al*. Recent Advancements of Bioinks for 3D Bioprinting of Human Tissues and Organs. *ACS Appl Bio Mater*. 2023;7(1):17-43.
doi: 10.1021/acsabm.3C00806
282. Havelikar U, Ghorpade KB, Kumar A, *et al*. Comprehensive insights into mechanism of nanotoxicity, assessment methods and regulatory challenges of nanomedicines. *Discov Nano*. 2024;19(1):165.
doi: 10.1186/S11671-024-04118-1
283. Shyam R, Palaniappan A. Effect of sterilization techniques on biomaterial inks' properties and 3D bioprinting parameters. *Bioprinting*. 2023;33:e00294.
doi: 10.1016/j.bprint.2023.E00294
284. Liang K. Tissue Bioprinting: Promise and Challenges. *Bioeng*. 2023;10(12):1400.
doi: 10.3390/bioengineering10121400
285. Nahon DM, Moerkens R, Aydogmus H, *et al*. Standardizing designed and emergent quantitative features in microphysiological systems. *Nat Biomed Eng*. 2024;8(8):941-962.
doi: 10.1038/S41551-024-01236-0
286. Wang Y, Zhang X, Yue H. Two-dimensional nanomaterials induced nano-bio interfacial effects and biomedical applications in cancer treatment. *J Nanobiotechnology*. 2024;22(1):67.
doi: 10.1186/S12951-024-02319-5
287. Xu Y, Chen S, Zhang Y, *et al*. Antibacterial black phosphorus nanosheets for biomedical applications. *J Mater Chem B*. 2023;11(30):7069-7093.
doi: 10.1039/D3TB00723E
288. Chen X, He W, Liang Y, *et al*. Enhanced degradation of few-layer black phosphorus by fulvic acid: Processes and mechanisms. *Water Res*. 2023;238:120014.
doi: 10.1016/j.watres.2023.120014
289. Mandrycky CJ, Howard CC, Rayner SG, Shin YJ, Zheng Y. Organ-on-a-chip systems for vascular biology. *J Mol Cell Cardiol*. 2021;159:1-13.
doi: 10.1016/j.yjmcc.2021.06.002
290. Ingber DE. Human organs-on-chips for disease modelling, drug development and personalized medicine. *Nat Rev Genet*. 2022;23(8):467-491.
doi: 10.1038/S41576-022-00466-9
291. Wang D, Maharjan S, Kuang X, *et al*. Microfluidic bioprinting of tough hydrogel-based vascular conduits for functional blood vessels. *Sci Adv*. 2022;8(43):eabq6900.
doi: 10.1126/sciadv.abq6900
292. Grebenyuk S, Abdel Fattah AR, Kumar M, *et al*. Large-scale perfused tissues via synthetic 3D soft microfluidics. *Nat Commun*. 2023;14(1):193.
doi: 10.1038/S41467-022-35619-1
293. Huang Y, Liu T, Huang Q, Wang Y. From Organ-on-a-Chip to Human-on-a-Chip: A Review of Research Progress and Latest Applications. *ACS Sensors*. 2024;9(7):3466-3488.
doi: 10.1021/acssensors.4c00004
294. Zhang B, Korolj A, Lai BFL, Radisic M. Advances in organ-on-a-chip engineering. *Nat Rev Mater*. 2018;3(8):257-278.
doi: 10.1038/s41578-018-0034-7
295. Ugodnikov A, Persson H, Simmons CA. Bridging barriers: advances and challenges in modeling biological barriers and measuring barrier integrity in organ-on-chip systems. *Lab Chip*. 2024;24(13):3199-3225.
doi: 10.1039/d3lc01027a
296. Liu J, Han X, Zhang T, Tian K, Li Z, Luo F. Reactive oxygen species (ROS) scavenging biomaterials for anti-inflammatory diseases: from mechanism to therapy. *J Hematol Oncol*. 2023;16(1):116.

- doi: 10.1186/S13045-023-01512-7
297. Liu J, Jia B, Li Z, Li W. Reactive oxygen species-responsive polymer drug delivery systems. *Front Bioeng Biotechnol.* 2023;11:01-11.
doi: 10.3389/fbioe.2023.1115603
298. Amirthalingam S, Rajendran AK, Moon YG, Hwang NS. Stimuli-responsive dynamic hydrogels: design, properties and tissue engineering applications. *Mater Horizons.* 2023;10(9):3325-3350.
doi: 10.1039/d3mh00399j
299. Chen S, Qiao Z, Niu Y, *et al.* Wearable flexible microfluidic sensing technologies. *Nat Rev Bioeng.* 2023;1(12):950-971.
doi: 10.1038/s44222-023-00094-w
300. Song JW, Munn LL. Fluid forces control endothelial sprouting. *Proc Natl Acad Sci U S A.* 2011;108(37):15342-15347.
doi: 10.1073/pnas.1105316108
301. Zhu W, Ma X, Gou M, Mei D, Zhang K, Chen S. 3D printing of functional biomaterials for tissue engineering. *Curr Opin Biotechnol.* 2016;40:103-112.
doi: 10.1016/j.copbio.2016.03.014
302. Wang Q, Zhang Y, Ma Y, Wang M, Pan G. Nano-crosslinked dynamic hydrogels for biomedical applications. *Mater Today Bio.* 2023;20:100640.
doi: 10.1016/j.mtbio.2023.100640